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Marine molluscs from Cyprus: new data and checklist

Bilal Öztürk, Giovanni Buzzurro & H. Avni Benli

KEY WORDS: Mollusca, Recent, Checklist, Cyprus, Mediterranean Sea.

ABSTRACT

This study has been carried out along the North Cypriot coasts on May 10th-19th 1997 and July 12th-21st 1998. As a result of the sampling in the shore region and in deepwater, depths ranging from 0 to 600 m, 304 species, for a total number of 4786 individuals, included in the classes of Polyplacophora, Gastropoda, Bivalvia and Scaphopoda have been identified. Among the 304 taxa examined, 18 species from Cyprus Island are here reported for the first time. In particular, 16 Gastropoda: Nodilittorina (Echinolittorina) punctata (Gmelin, 1791), Rissoa splendida Eichwald, 1830, Alvania fractospira Oberling, 1970, Alvania testae (Aradas & Maggiore, 1844), Hydrobia acuta (Draparnaud, 1805), Protaslanta souleyeti (Smith E. A., 1888), Cerithiopsis scalaris Locard, 1892, ex Monterosato ms., Nassarius circumcictus (Adams A., 1852), Granulina boucheti Gofas, 1992, Gymno bela subaraneosa (Dautzenberg & Fischer H., 1896), Teretia teres (Reeve, 1844), Chrysallida juliae (de Folin, 1872), Folinella gbisottii Aartsen, 1984, Tragula fenestrata (Jeffreys, 1848), Eulimella bogii Aartsen, 1994, Turbonilla acutissima Monterosato, 1884, and 2 Bivalvia: Limaria loscombi (Sowerby G. B. I, 1823), Kelliella abyssicola (Forbes, 1844). Furthermore, six species are reported for the Eastern basin for the first time. All the species reported for the first time from Cyprus in this paper are figured and discussed. A total amount of 696 species is here reported in a newly formed checklist of Molluscs of Cyprus, considering all the previous works concerned about the malacological fauna occurring in the area under investigation and the additional species we have identified. Furthermore, the list, which is taking into account the many nomenclatural changes recently introduced in the class Mollusca, also deals with alien species.

RIASSUNTO

Questo studio è stato condotto lungo le coste della parte Nord dell'isola di Cipro tra il 10 e il 19 Maggio del 1997 e il 12 e il 21 Luglio del 1998. Sono stati raccolti campioni fino alla profondità massima di 600 m, individuando 4786 esemplari distribuiti tra 304 specie appartenenti alle classi Poliplacophora, Gastropoda, Bivalvia e Scaphopoda. Di queste 304 specie 18 risultano inedite per Cipro, 16 Gastropoda: Nodilittorina (Echinolittorina) punctata (Gmelin, 1791), Rissoa splendida Eichwald, 1830, Alvania fractospira Oberling, 1970, Alvania testae (Aradas & Maggiore, 1844), Hydrobia acuta (Draparnaud, 1805), Protatlanta souleyeti (Smith E. A., 1888), Cerithiopsis scalaris Locard, 1892, ex Monterosato ms., Nassarius circumcictus (Adams A., 1852), Granulina boucheti Gofas, 1992, Gymnobela subaraneosa (Dautzenberg & Fischer H., 1896), Teretia teres (Reeve, 1844), Chrysallida juliae (de Folin, 1872), Folinella ghisottii Aartsen, 1984, Tragula fenestrata (Jeffreys, 1848), Eulinella bogii Aartsen, 1994, Turbonilla acutissima Monterosato, 1884 e 2 Bivalvia: Limaria loscombi (Sowerby G. B. I, 1823), Kelliella abyssicola (Forbes, 1844). Tra queste, 6 specie sono inedite per il bacino orientale del Mar Mediterraneo. Di tutte le specie trattate si dà anche una rappresentazione iconografica. Viene presentata inoltre una nuova lista aggiornata comprendente tutte le specie precedentemente segnalate in Letteratura e quelle ulteriormente identificate da noi, per un totale di 696 taxa. Questa lista, corredata da relative note integrative ed in accordo con gli ultimi aggiornamenti tassonomici, evidenzia anche le specie di provenienza indopacifica.

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INTRODUCTION the complicated taxonomy of some groups, and several species were listed as separate taxa in spite of being synonyms accord-The island of Cyprus has special ecological characteristics due ing to the latest taxonomy. In this paper we present an up to date checklist of Mollusca species occurring in the Island of Cyprus. With this report, the total number of Mollusca occurring along the Cyprus coasts has been enriched with 16 Gastropoda and 2 Bivalvia species.

MATERIAL AND METHODS

Specimens and shells identified in this study were collected during the expeditions along Northern Cyprus coasts on the dates of May 10th-19th, 1997 and July 12nd-21st, 1998 by R/V " K. Piri Reis". Samples were taken from a depth ranging from 0 to 10 m along the shore region and from 20 to 600 m offshore (Fig.1). Sampling have been randomly selected in the coastal regions having both sandy and rocky habitat and in the substrates consisting of algae (Padina pavonica (L) Thivy, 1960, Cystoseira crinita (Desf.) Bory, 1846, Cystoseira sp., Sargassum vulgare C. Agardh, 1821, Udotea petiolata (Turra) Boerg, 1915, Caulerpa prolifera (Forskkal) Lamaroux, 1809, Caulerpa racemosa (Forskkal) J. Agardh), phanerogam (Posidonia oceanica (L.) Delile, 1813) and sponge (Sarcotragus sp.) (Table 1). Samples were collected directly by hand. A total of 70 samplings were carried out at 12 stations. Furthermore, in the open sea, trawl-

to its narrow sublittoral and littoral but wide bathyal zones. This island, with a coastline of 750 km in length, is an important biogeographical spot in the Levantine basin where Red Sea originated Lessepsian migrants establish. There are many studies so far carried out by different scientists on the biological diversity Cyprus' bottoms. The first report on the Mollusca from the coasts of this island was by MONTEROSATO (1899). Thereafter, many works were carried out especially in the second half of century. Among the others, we cite TSURNAMAL (1969), Demetropoulos (1969, 1971), Barash & Lewinsohn (1971), Demetropoulos & Hadjichristophorou (1976), STAVRINOS (1984), TORNARITIS (1987), BARASH & DANIN (1989), Bogi et al. (1989), Fischer (1993a, b; 1994), Cecalupo & QUADRI (1994, 1995, 1996) and HADJICHRISTOPHOROU et al. (1997). As a result of these studies, 585 Mollusca species have been reported from Cyprus (CECALUPO & QUADRI, 1996). More recently, Buzzurro & Greppi (1997) and Salman et al. (1998) have reported 42 and 10 additional species, respectively.

Several checklists of the molluscan fauna of Cyprus have been published in the past by different Authors (DEMETROPOULOS, 1971; BOGI et al., 1989; FISCHER, 1994 and CECALUPO & QUADRI, 1996). However, some of these early reports suffered



ing and dredging have been utilized. Samples were sieved with 0.5 mm mesh size.

Systematics of the identified species are according to SABELLI *et al.* (1990-1992). Moreover, species names reported in the past by several Authors have been reconsidered according to SABELLI *et al.* (1990-1992) during the preparation of this new checklist. Thus, the list was cleaned up of synonym species. The many

changes occurred in the nomenclature of Mollusca after 1990 have been also considered. Furthermore, systematic information about some species was adopted by visiting the web-site Check List of European Marine Mollusca (CLEMAM: home page, http://www.mnhn.fr/base/malaco/html).

The materials collected were deposited in the Department of Hydrobiology, Ege University (Izmir, Turkey).

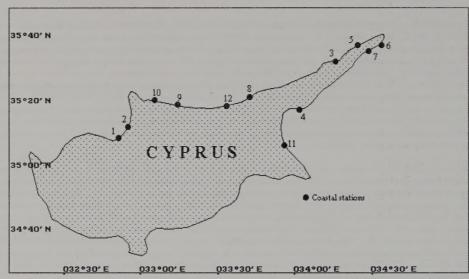


Fig. 1. Map of studied area with locations of shallow water stations.

Fig. 1. Cartina dell'area studiata con l'indicazione delle stazioni di acque poco profonde.

Table 1. Coordinates,	localities, depths.	dates and biotor	characterizations	of the samples.

	BOTTOM-TRAWL									
St.	Date	Coordinates or Localities	Depth (m)	Biotopes						
		Start / Finish ·								
T1	10.5.97	35°21'7"N-33°07'5"E / 35°23'9"N-33°09'0"E	560-576	Mud						
T2	10.5.97	35°22'7"N-33°02'6"E / 35°22'7"N-33°04'2"E	33-35	Hard substratum and Sarcotragus sp.						
T3	11.5.97	35°20′58"N-33°23′4"E / 35°20′7"N-33°22′5"E	40-46	Posidonia oceanica						
T4	12.5.97	35°20′7"N-33°28′6"E / 35°20′6"N-33°27′5"E	28-38	Posidonia oceanica						
T5	12.5.97	35°22'9"N-33°41'0"E / 35°22'4"N-33°39'3"E	38-45	Posidonia oceanica and Caulerpa prolifera						
T7	13.5.97	35°19'2"N-32°54'6"E / 35°17'7"N-32°54'4"E	28-45	Posidonia oceanica						
T9	13.5.97	35°12′5″N-32°52′3″E / 35°11′2″N-32°51′8″E	75-85	Muddy sand with Caulerpa racemosa						
T10	16.5.97	35°40'3"N-34°39'1"E / 35°39'7"N-34°37'8"E	275-337	Mud						
T11	16.5.97	35°40′9″N-34°35′4″E / 35°40′6″N-34°35′3″E	27-45	Posidonia oceanica						
T12	17.5.97	35°26'7"N-34°10'4"E / 35°25'0"N-34°08'4"E	32-95	Posidonia oceanica						
T13	17.5.97	35°25'1"N-34°11'1"E / 35°26'3"N-34°11'7"E	37-69	Posidonia oceanica						
T14	17.5.97	35°21′5″N-34°10′4″E / 35°21′9″N-34°08′9″E	187-155	Mud						
T17	185.97	35°13'9"N-33°56'9"E / 35°15'2"N-33°57'0"E	100-145	Hard substratum						
T18	18.5.97	35°13'6"N-33°56'0"E / 35°15'0"N-33°55'8"E	62-70	Muddy sand with Caulerpa racemosa						
T19	18.5.97	35°12'0"N-33°55'6"E / 35°13'3"N-33°55'1"E	37-38	Muddy-sand with Caulerpa racemosa and Udotea petiolata						
T20	19.5.97	35°11'1"N-33°56'7"E / 35°10'3"N-33°55'9"E	50-100	Mud and Sarcotragus sp.						
T21	19.5.97	35°11'1"N-33°58'0"E / 35°10'0"N-33°57'0"E	66-150	Muddy sand with Caulerpa racemosa and Udotea petiolata						
T22	14.7.98	35°19'1"N-32°49'0"E / 35°18'6"N-32°49'9"E	223-227	Mud						
T23	17.7.98	35°47'7"N-34°43'8"E / 35°47'9"N-34°44'1"E	113-114	Mud						
T24	19.7.98	35°06'4"N-34°00'5"E / 35°07'6"N-33°59'6"E	90-89	Mud						
T25	19.7.98	35°09'4"N-33°59'1"E / 35°10'4"N-33°58'5"E	105-131	Mud with shell fragments						
T26	19.7.98	35°11'8"N-33°56'3"E / 35°10'6"N-33°57'2"E	59-70	Mud						



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St.	Date	Coordinates or Localities Start / Finish	Depth (m)	Biotopes
D1	10.5.97	35°29′1"N-33°58′0"E	50	Hard substratum
D2	12.5.97	35°22'4"N-33°39'9"E	35	Sandy mud with shell fragments
D3	12.5.97	35°20'7"N-33°28'3"E	35	Sandy mud with shell fragments
D4	15.5.97	35°37'8"N-34°21'1"E	35	Sand with Brachiostoma lanceolatum and Posidonia oceanic
D7	16.5.97	35°35'7"N-34°27'4"E	70	Mud with Ascidia sp.
D8	16.5.97	35°34'8"N-34°25'9"E	35	Posidonia oceanica
D9	16.5.97	35°34'4"N-34°26'9"E	140	Mud
	16.5.97	35°34'8"N-34°28'3"E	210	Mud
	19.5.97	35°10'0"N-33°59'4"E	25	Posidonia oceanica
	19.5.97	35°08'7"N-33°57'6"E	92	Hard substratum with Sargassum vulgare
	12.7.98	35°30'0"N-33°09'9"E	600	Mud
	14.7.98	35°10'0"N-32°50'0"E	69	Sandy mud with shell fragments
	14.7.98	35°19'9"N-32°50'1"E	210	Mud
D17	16.7.98	35°43'8"N-34°37'3"E	250-300	Mud with shell fragments
D18	16.7.98	35°33'8"N-34°13'0"E	32	Posidonia oceanica
D19		35°24'8"N-34°10'1"E	50	Muddy sand with Caulerpa racemosa
D20	18.7.98	35°08'7"N-34°00'1"E	120	Mud
D21	19.7.98	35°09'0"N-33°57'0"E	20	Posidonia oceanica
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St.	Date	Coordinates or Localities	Depth (m)	Biotopes
		Start / Finish	1	
C1	14.7.98	Guzelyurt Bay	0-10	Cystoseira crinita, Posidonia oceanica, rocks and sand
C2	14.7.98	Guzelyurt Bay	0-10	Cystoseira crinita and sand
C3	18.7.98	Karpas Cape	0-10	Cystoseira crinita, Posidonia oceanica and rocks
C4	18.7.98	Famagusta Bay	0-10	Posidonia oceanica and rocks
C5	19.7.98	Karpas Cape	0-10	Cystoseira sp., Posidonia oceanica and rocks
C6	19.7.98	Karpas Cape	0-10	Padina pavonica, Posidonia oceanica and rocks
C7	19.7.98	Karpas Cape	0-10	Cystoseira sp. and rocks
C8	21.7.98	Girne	0-10	Cystoseira crinita, Posidonia oceanica and rocks
C9	21.7.98	Malazgirt	0-10	Padina pavonica, Cystoseira crinita, Posidonia oceanica and rocks
	21.7.98	Kormakiti Cape	0-10	Cystoseira crinita, Posidonia oceanica and sand
C11	21.7.98	Famagusta Harbour	0-3	Rocks with Cystoseira crinita
C12	13.7.98	Girne Harbour	0-1	Padina pavonica

RESULTS

During the study carried out in the Northern Cypriot coasts, According to the literature so far analyzed, 6 of these 18 species 4786 specimens were collected. As a result of evaluation of this sampling, 9 Polyplacophora, 183 Gastropoda, 110 Bivalvia and 2 i.e.: Rissoa splendida Eichwald, 1830, Protatlanta souleyeti (Smith Scaphopoda species were identified (Table 2).

here reported for the first time from Cyprus (Table 3).

have never been reported before for the entire Levantine basin, E. A., 1888), Cerithiopsis scalaris Locard, 1892, ex Monterosato Out of 304 species identified during this study, 18 species are ms., Granulina boucheti Gofas, 1992, Teretia teres (Reeve, 1844) and Turbonilla acutissima (Monterosato, 1884).

Table 2: Taxonomical distribution of the species examined.

1				
Classis	Ordo	Familia	Genus	Species
Polyplacophora	1	4	6	9
Gastropoda	.12	59	109	183
Bivalvia	10	38	83	110
Scaphopoda	_	2	2	2
Total	23	103	200	304
1 1-				



Table 3: Molluscs species known from the Cyprus Island (examined material and literature).

L: literature (Lx: Authors from the literature); FR: first record for the area; N: notes (*indopacific origin); R: findings from our material; Z: figures.

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SPECIES POLYPLA COPHORA	L	N	R	Z
POLYPLACOPHORA				
Leptochitonidae	T 2			
Lepidopleurus cajetanus (Poli, 1791)	L3	1	+	
Lepidopleurus bedullii (Dell'Angelo & Palazzi, 1986)	L23	1		
Lepidopleurus cancellatus (Sowerby G. B. II, 1840)	L3			
Ischnochitonidae	TE			
Ischnochiton rissoi (Payraudeau, 1826)	L5		+	
Callochiton septemvalvis (Montagu, 1803)	L2		+	
Lepidochitona cinerea (Linnaeus, 1767)	L23		+	
Lepidochitona caprearum (Scacchi, 1836)	L5		+	
Lepidochitona monterosatoi Kaas & Van Belle, 1981	L11		+	
Chitonidae				
Chiton corallinus (Risso, 1826)	L5		+	
Chiton olivaceus Spengler, 1797	L2		+	
Chiton phaseolinus Monterosato, 1879	L23			
Acanthochitonidae		2		
Acanthochitona crinita (Pennant, 1777)	L11	3		
Acanthochitona fascicularis (Linnaeus, 1767)	L2		+	
GASTROPODA				
Patellidae				
Patella caerulea Linnaeus, 1758	L1		+	
Patella rustica Linnaeus, 1758	L1		+	
Patella ulyssiponensis Gmelin, 1791	L9		+	
Lottiidae				
Tectura virginea (Müller O. F., 1776)	L2			
Lepetidae				
Propilidium scabrosum Jeffreys, 1883	L4			
Addisoniidae				
Addisonia lateralis (Réquien, 1848)	L9			
Neritidae				
Smaragdia souverbiana (Montrouzier, 1863)	L24	*		
Smaragdia viridis (Linnaeus, 1758)	L2		+	
Fissurellidae				
Fissurella nubecula (Linnaeus, 1758)	L4			
Diodora dorsata (Monterosato, 1878)	L23			
Diodora gibberula (Lamarck, 1822)	L4		+	
Diodora graeca (Linnaeus, 1758)	L4		+	
Diodora italica (Defrance, 1820)	L2		+	
Emarginula octaviana Coen, 1939	L2			
Emarginula sicula Gray, 1825	L2			
Emarginella huzardii (Payraudeau, 1826)	L9		+	
Scissurellidae				
Scissurella costata D'Orbigny, 1824	L11		+	
Anatoma crispata Fleming, 1828	L24		+	
Sinezona cingulata (Costa O. G., 1861)	L11			
Haliotidae				
Haliotis tuberculata tuberculata Linnaeus, 1758	L1	4	+	
Trochidae	~-			
Trochus erythraeus Brocchi, 1826	L9	*	+	
Clanculus (Clanculus) corallinus (Gmelin, 1791)	L2		+	
Clanculus (Clanculopsis) cruciatus (Linnaeus, 1758)	L3		_	
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Table 5 continued				
SPECIES	L	N	R	Z
Clanculus (Clanculopsis) jussieui (Payraudeau, 1826)	L4			
Calliostoma conulus (Linnaeus, 1758)	L9		+	
Calliostoma laugieri laugieri (Payraudeau, 1826)	L2		+	
Calliostoma zizyphinum (Linnaeus, 1758)	L2			
Calliostoma granulatum (Von Born, 1778)	L2		+	
Gibbula ardens (Von Salis, 1793)	L1		+	
Gibbula magus (Linnaeus, 1758)	L3		+	
Gibbula adansonii (Payraudeau, 1826)	L4		+	
Gibbula turbinoides (Deshayes, 1835)	L9		+	
Gibbula fanulum (Gmelin, 1791)	L4		+	
Gibbula guttadauri (Philippi, 1836)	L9			
Gibbula leucophaea (Philippi, 1836)	L9			
Gibbula philberti (Recluz, 1843)	L11			
Gibbula richardi (Payraudeau, 1826)	L5		+	
Gibbula varia (Linnaeus, 1758)	L2		+	
Gibbula divaricata (Linnaeus, 1758)	L4		+	
Gibbula rarilineata (Michaud, 1829)	L9			
Gibbula umbilicaris nebulosa (Philippi, 1848)	L1	5	+	
Osilinus articulatus Lamarck, 1822	L1		+	
Osilinus mutabilis (Philippi, 1846)	L10			
Osilinus turbinatus (Von Born, 1778)	L1		+	
Clelandella miliaris (Brocchi, 1814)	L24			
Jujubinus exasperatus (Pennant, 1777)	L2		+	
Jujubinus striatus striatus (Linnaeus, 1758)	L3		+	
Colloniidae	22		·	
Cantrainea peloritana (Cantraine, 1835)	L9			
Homalopoma carmelae Oliverio & Buzzurro, 1994	L19			
Homalopoma sanguineum (Linnaeus, 1758)	L2		+	
Tricoliidae	1.2		1	
Tricolia pullus pullus (Linnaeus, 1758)	L2		+	
Tricolia speciosa (Von Muehlfeldt, 1824)	L2			
Tricolia tenuis (Michaud, 1829)	L9		+	
Turbinidae	L		1	
Bolma rugosa (Linnaeus, 1758)	L7		+	
Cerithiidae	L/		-	
Cerithium alucaster (Brocchi, 1814)	L9			
	L24			
Cerithium haustellum Monterosato in Crema, 1903	L24 L24			
Cerithium lividulum Risso, 1826	L11	*		
Cerithium nesioticum Pilsbry & Vanatta, 1906	L24			
Cerithium protractum Bivona Ant. in Bivona And., 1838	L24 L1		+	
Cerithium rupestre Risso, 1826	L15	*	+	
Cerithium scabridum Philippi, 1848		,	+	
Cerithium vulgatum Bruguière, 1792	L1		+	
Bittium jadertinum (Brusina, 1865)	L11		+	
Bittium latreillii (Payraudeau, 1826)	L9		+	
Bittium reticulatum (Da Costa, 1778)	L2		+	
Bittium scabrum (Olivi, 1792)	L23	*		
Rhinoclavis kochi (Philippi, 1848)	L4	4.		
Fossaridae	T 1 1			
Fossarus ambiguus (Linnaeus, 1758)	L11			
Obtortionidae	102	*		
Finella pupoides Adams, A., 1860	L23	*	+	
Clathrofenella fusca (Adams, A., 1860)	L21	-4,	+	
Potamididae				



Table 3 continued				
SPECIES	L	N	R	Z
Pirenella conica (Blainville, 1826)	L9		+	
Siliquariidae				
Tenagodus obtusus (Schumacher, 1817)	L3			
Turritellidae				
Turritella communis Risso, 1826	L2		+	
Turritella turbona Monterosato, 1877	L2			
Littorinidae	22			
Melarhaphe neritoides (Linnaeus, 1758)	L1	6	+	
Nodilittorina (Echinolittorina) punctata (Gmelin, 1791)	FR	· ·	+	2
Cingulopsidae (Cingulopsidae)	110		,	4
Eatonina fulgida (Adams, J., 1797)	L11			
Rissoidae	1211			
Rissoa angustior (Monterosato, 1917)	L23			
Rissoa auriformis pseudomonodonta Verduin, 1983	L12			
Rissoa auriscalpium (Linnaeus, 1758)	L12			
Rissoa decorata Philippi, 1846	L23			
Rissoa guerinii Récluz, 1843	L9			
Rissoa italiensis Verduin, 1985	L23		+	
Rissoa labiosa (Montagu, 1803)	L23			
Rissoa lia (Monterosato, 1884, ex Benoit ms.)	L9			
Rissoa monodonta Philippi, 1836	L9		+	
Rissoa rodhensis Verduin, 1985	L23		+	
Rissoa scurra (Monterosato, 1917)	L11		+	
Rissoa similis Scacchi, 1836	L9		+	2
Rissoa splendida Eichwald, 1830	FR		+	3
Rissoa variabilis (Von Muehlfeldt, 1824)	L9		+	
Rissoa ventricosa Desmarest, 1814	L2		+	
Rissoa violacea violacea Récluz, 1843	L9	_	+	
Alvania amatii Oliverio, 1986	L11	7	+	
Alvania aspera (Philippi, 1844)	L9			
Alvania beani (Hanley in Thorpe, 1844)	L9			
Alvania beniamina (Monterosato, 1884)	L6			
Alvania cancellata (Da Costa, 1778)	L18		+	
Alvania chiarellii Cecalupo & Quadri, 1995	L21			
Alvania cimex (Linnaeus, 1758)	L1		+	
Alvania colossophilus Oberling, 1970	L11			
Alvania datchaensis Amati & Oliverio, 1987	L12			
Alvania discors (Allan, 1818)	L9		+	
Alvania dorbignyi (Audouin, 1827)	L11	8		
Alvania fractospira Oberling, 1970	FR		+	4
Alvania geryonia (Nardo, 1847, ex Chiereghini ms.)	L11		+	
Alvania cf. hallgassi Amati & Oliverio, 1985	L11	25		
Alvania hispidula (Monterosato, 1884)	L24			
Alvania lactea (Michaud, 1832)	L18			
Alvania lanciae (Calcara, 1841)	L9			
Alvania lineata Risso, 1826	L9		+	
Alvania mamillata Risso, 1826	L8			
Alvania oranica (Pallary, 1900)	L11			
Alvania paupercula (Jeffreys, 1867)	L9			
Alvania rudis (Philippi, 1844)	L23			
Alvania scabra (Philippi, 1844)	L23		+	
Alvania sculptilis (Monterosato, 1877)	L23			
Alvania testae (Aradas & Maggiore, 1844)	FR		+	- 5
Cingula trifasciata (Adams J., 1800)	L9			



SPECIES		L	N	R Z
Manzonia crassa (Kanmacher, 1798)		L11		+
Manzonia weinkauffi jacobusi Oliverio, Amati & Nofroni, 1	986	L23		
Obtusella macilenta (Monterosato, 1880)		L23		
Peringiella elegans (Locard, 1892)		L23		
Pusillina diversa (Nordsieck, 1972)		L23		
Pusillina inconspicua (Alder, 1844)		L23		
Pusillina lineolata (Michaud, 1832)		L9		
Pusillina margiminia (Nordsieck, 1972)		L23 ⁻		
Pusillina marginata (Michaud, 1832)		L23		
Pusillina munda (Monterosato, 1884)		L23		
Pusillina philippi (Aradas & Maggiore, 1844)		L23		+
Pusillina radiata (Philippi, 1836)		L11		
Setia ambigua (Brugnone, 1873)		L23		
Setia fusca (Philippi, 1841)		L23	9	
Setia slikorum (Verduin, 1984)		L23		
Setia turriculata Monterosato, 1884		L11		
Rissoina bruguieri (Payraudeau, 1826)		L9		+
Rissoina bertholleti Issel, 1869		L11	*	
Adeorbidae				
Circulus striatus (Philippi, 1836)		L9		
Anabathridae				
Pisinna glabrata (Von Muhelfeldt, 1824)		L11		
Assimineidae				
Paludinella littorina (Delle Chiaje, 1828)		L11		
Paludinella cf. sicana (Brugnone, 1876)		L23		
Barleeidae				
Barleeia unifasciata (Montagu, 1803)		L9		
Caecidae				
Caecum auriculatum de Folin, 1868		L11		+
Caecum trachea (Montagu, 1803)		L11		+
Hydrobiidae				
Hydrobia acuta (Draparnaud, 1805)		FR		+ 6
Tornidae				
Tornus subcarinatus (Montagu, 1803)		L9		+
Truncatellidae				
Truncatella subcylindrica (Linnaeus, 1767)		L2		+
Strombidae				
Strombus persicus Swainson, 1821		L9	*	+
Aporrhaiidae				
Aporrhais pespelecani (Linnaeus, 1767)		L2		+
Crepidulidae			10	
Crepidula unguiformis Lamarck, 1822		L2		
Capulidae				
Capulus hungaricus (Linnaeus, 1758)		L2		
Vermetidae				
Vermetus cristatus Biondi, 1857		L9		
Vermetus rugulosus Monterosato 1878		L23		
Vermetus triquetrus Bivona Ant., 1832		L2		+
Vermetus cf. granulatus (Gravenhorst, 1831)		L24		
Vermetus cf. semisurrectus Bivona Ant., 1832		L24		
Vermetus sp.		L23		
Dendropoma anguliferum (Monterosato, 1884)		L11		
Dendropoma petraeum (Monterosato, 1884)		L2		
Petaloconchus glomeratus (Linnaeus, 1758)		L7		
Townsonous growns (Minnetts, 1770)				



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SPECIES	L	N	R	Z
Serpulorbis arenaria (Linnaeus, 1767)	L1		+	
Cypraeidae		11		
Erosaria spurca (Linnaeus, 1758)	L2		+	
Luria lurida (Linnaeus, 1758)	L2			
Lamellariidae				
Lamellaria latens (Müller O. F., 1776)	L9			
Lamellaria perspicua (Linnaeus, 1758)	L3			
Triviidae				
Trivia arctica (Pulteney, 1789)	L1			
Trivia levantina Smriglio, Mariottini & Buzzurro, 1998	L27		+	
Trivia pulex (Solander in Gray, 1828)	L2			
Naticidae				
Natica dillwynii Payraudeau, 1826	L1		+	
Natica hebraea (Martyn, 1784)	L7		+	
Natica stercusmuscarum (Gmelin, 1791)	L7		+	
Tecnonatica filosa (Philippi, 1845)	L18			
Tecnonatica rizzae (Philippi, 1844)	L9			
Euspira guillemini (Payraudeau, 1826)	L18			
Euspira pulchella (Risso, 1826)	L7		+	
Neverita josephinia Risso, 1826	L7		+	
Payraudeautia intricata (Donovan, 1804)	L9			
Tonnidae				
Tonna galea (Linnaeus, 1758)	L2		+	
Cassidae				
Galeodea echinophora (Linnaeus 1758)	L2		+	
Galeodea rugosa (Linné, 1771)	L3			
Phalium granulatum (Von Born, 1778)	L3		+	
Phalium saburon (Bruguière, 1792)	L2			
Ranellidae				
Ranella olearia (Linnaeus, 1758)	L9			
Cymatium corrugatum corrugatum (Lamarck, 1816)	L2			
Cymatium parthenopeum parthenopeum (Von Salis, 1793)	L9			
Cabestana cutacea cutacea (Linnaeus, 1767)	L9			
Charonia lampas lampas (Linnaeus, 1758)	L3			
Charonia variegata (Lamarck, 1816)	L2	12	+	
Atlantidae				
Atlanta peronii Lesueur, 1817	L24			
Oxygyrus keraudrenii (Lesueur, 1817)	L24			
Protatlanta souleyeti (Smith E. A., 1888)	FR		+	7
Firolidae				
Firoloida desmarestia Lesueur, 1817	L13			
Pterotrachea hippocampus Philippi, 1836	L13			
Pterotrachea minuta Bonnevie, 1920	L13			
Triphoridae				
Cheirodonta pallescens (Jeffreys, 1867)	L23			
Marshallora adversa (Montagu 1803)	L15		+	
Monophorus perversus (Linnaeus, 1758)	L2		+	
Similiphora similior (Bouchet & Guillemot, 1978)	L23			
Metaxia bacillum (Issel, 1869)	L21	*		
Metaxia metaxa (Delle Chiaje, 1828)	L13			
Cerithiopsidae				
Cerithiopsis diadema Monterosato, 1864, ex Watson ms.	L24			
Cerithiopsis fayalensis Watson, 1886	L23			
Cerithiopsis minima (Brusina, 1865)	L23		+	



Table 3 continued				
SPECIES	L	N	R	Z
Cerithiopsis nana Jeffreys, 1867	L23			
Cerithiopsis pulvis (Issel, 1869)	L11	*		
Cerithiopsis scalaris Locard, 1892, ex Monterosato ms.	FR		+	8
Cerithiopsis tenthrenois (Melvill, 1869)	L24	*		
Cerithiopsis tubercularis (Montagu, 1803)	L11		+	
Krachia tiara Monterosato, 1874, ex Watson ms.	L23			
Dizoniopsis bilineata (Hörnes, 1848)	L11			
Dizoniopsis coppolae (Aradas, 1870)	L23			
Seila trilineata (Philippi, 1836)	L9			
Janthinidae				
Janthina janthina (Linnaeus, 1758)	L9			
Janthina nitens Menke, 1828	L2			
Cimidae				
Cima cylindrica (Jeffreys, 1858)	L11			
Epitoniidae				
Epitonium algerianum (Weinkauff, 1866)	L23			
Epitonium clathratulum (Kanmacher, 1798)	L3			
Epitonium commune (Lamarck, 1822)	L9		+	
Epitonium turtoni (Turton, 1819)	L4			
Cycloscala hyalina (Sowerby G. B. II, 1844)	L17	*		
Gyroscala lamellosa (Lamarck, 1822)	L2			
Opalia hellenica (Forbes, 1844)	L23		+	
Eulimidae				
Crinophtheiros comatulicola (Graff, 1875)	L11			
Ersilia mediterranea (Monterosato, 1869)	L11			
Eulima bilineata Alder, 1848	L25		+	
Eulima glabra (Da Costa, 1778)	L2		+	
Melanella boscii (Payraudeau, 1827)	L23			
Melanella monterosatoi (Monterosato, 1890, ex De Boury ms.)	L23			
Melanella polita Linnaeus, 1758)	L11			
Parvioris ibizenca (Nordsieck, 1968)	L23			
Nanobalcis nana (Monterosato, 1878)	L24			
Sabinella piriformis Brugnone, 1873	L24			
Sticteulima lentiginosa (Adams A., 1861)	L24	*		
Sticteulima jeffreysiana (Brusina, 1869)	L23			
Vitreolina curva (Monterosato, 1874, ex Jeffreys ms.)	L9		+	
Vitreolina devians Monterosato, 1875	L24			
Vitreolina incurva (B. D. D., 1883)	L11			
Vitreolina levantina Oliverio, Buzzurro & Villa, 1994	L20			
Vitreolina perminima (Jeffreys, 1883)	L9			
Vitreolina philippi (Rayneval & Ponzi, 1854)	L9		+	
Muricidae				
Aspella anceps (Lamarck, 1822)	L4	*		
Bolinus brandaris (Linnaeus, 1758)	L2		+	
Dermomurex scalaroides (Blainville, 1829)	L23			
Hadriania oretea (De Gregorio, 1885)	L4			
Hexaplex trunculus (Linnaeus, 1758)	L2		+	
Murexsul aradasii (Poirier, 1883, ex Monterosato ms.)	L9			
? Muricopsis cevikeri Houart, 2000	L28			
Muricopsis cristata (Brocchi, 1814)	L2		+	
Ocenebra erinaceus (Linnaeus, 1758)	L4		+	
Ocinebrina aciculata (Lamarck, 1822)	L3		+	
Ocinebrina edwardsii (Payraudeau, 1826)	L4		+	
Ocinebrina hispidula (Pallary, 1904)	L23			



Table 3 continued				
SPECIES	L	N	R	Z
Ocinebrina hybrida (Aradas & Benoit, 1876)	L11			
Typhinellus sowerbyi (Broderip, 1833)	L4			
Ergalatax obscura Houart, 1996	L24	*		
Thais lacera (Born, 1778)	L28	*		
Stramonita haemastoma (Linnaeus, 1766)	L2		+	
Coralliophila meyendorffii (Calcara, 1845)	L4		·	
Hirtonurex squamosa (Bivona And., 1838)	L9			
Babelomurex babelis (Requién, 1848)	L3			
Buccinidae	11)			
Buccinulum corneum (Linnaeus, 1758)	L2		+	
Chauvetia candidissima (Philippi, 1836)	L18			
Chauvetia recondita (Brugnone, 1873)	L23			
Engina leucozona (Philippi, 1843)	L25			
Pisania striata (Gmelin, 1791)	L4 L2	`	+	
	L7		+	
Pollia dorbignyi (Payraudeau, 1826)			+	
Pollia scabra Locard, 1886	L9			
Pollia scacchiana (Philippi, 1844)	L9		+	
Colubraria reticulata (Blainville, 1826)	L3			
Columbellidae	Υ 1			
Columbella rustica (Linnaeus, 1758)	L1	414	+	
Anachis savignyi (Moazzo, 1939, ex Jousseaume ms.)	L24	*	+	
Anachis troglodytes (Sowerbie G. B. I & Montrouzier, 1866)	L23	*		
Mitrella cf. bruggeni Aartsen, Menkhorst & Gittenberg, 1984	L23			
Mitrella gervillii (Payraudeau, 1826)	L9		+	
Mitrella minor (Scacchi, 1836)	L11			
Mitrella scripta (Linnaeus, 1758)	L2		+	
Nassariidae				
Nassarius corniculus (Olivi, 1792)	L2			
Nassarius incrassatus (Stroem, 1768)	L7		+	
Nassarius reticulatus (Linnaeus, 1758)	L2			
Nassarius circumcictus (Adams A., 1852)	FR		+	9
Nassarius gibbosulus (Linnaeus, 1758)	L2		+	
Nassarius mutabilis (Linnaeus, 1758)	L2		+	
Nassarius cuvierii (Payraudeau, 1826)	L9		+	
Nassarius louisi (Pallary, 1912)	L9		+	
Nassarius lima (Dillwyn, 1817)	L9			
Cyclope neritea (Linnaeus, 1758)	L2		+	
Fasciolariidae				
Fasciolaria lignaria (Linnaeus, 1758)	L2		+	
Fusinus sanctaeluciae (Von Salis, 1793)	L2		+	
Fusinus rudis (Philippi, 1844)	L9			
Fusinus syracusanus (Linnaeus, 1758)	L2		+	
Costellariidae		13		
Vexillum ebenus (Lamarck, 1811)	L2		+	
Vexillum littorale (Forbes, 1844)	L9		+	
Vexillum savignyi (Payraudeau, 1826)	L9			
Vexillum tricolor (Gmelin, 1790)	L3		+	
Marginellidae				
Volvarina mitrella (Risso, 1826)	L4		+	
Cystiscidae				
Gibberula miliaria (Linnaeus, 1758)	L2		+	
Gibberula philippii (Monterosato, 1878)	L11		+	
Granulina boucheti Gofas, 1992	FR		+	10
Granulina marginata (Bivona, 1832)	L23	14		



SPECIES	L	N	R	Z
Granulina occulta (Monterosato, 1869)	L11	~ 1		-
Mitridae				
Mitra cornicula (Linnaeus, 1758)	L2		+	
Mitra zonata Marryat, 1818	L9			
Cancellariidae				
Cancellaria cancellata (Linnaeus, 1767)	L4			
Turridae				
Haedropleura septangularis (Montagu, 1803)	L18			
Conidae				
Conus mediterraneus Hwass in Bruguière, 1792	L1		+	
Bela cf. cycladensis (Reeve, 1845)	L24			
Bela laevigata (Philippi, 1836)	L15		+	
Bela menkhorsti Aartsen, 1988	L15			
Bela nebula (Montagu, 1803)	L3		+	
Bela ornata (Locard, 1897)	L23			
Bela taprurensis (Pallary, 1904)	L11		+	
Clathromangelia granum (Philippi, 1844)	L9	15		
Gymnobela subaraneosa (Dautzenberg & Fischer H., 1896)	FR		+	11
Mangelia attenuata (Montagu, 1803)	L3		+	
Mangelia barashi (Aartsen & Fehr de Wal, 1978)	L11		+	
Mangelia bertrandii (Payraudeau, 1826)	L11			
Mangelia costulata (Blainville, 1829)	L3			
Mangelia fieldeni (Aartsen & Fehr de Wal, 1978, ex Mont. ms.)	L11	16	+	
Mangelia cf. pallary (Nordsieck, 1977)	L11			
Mangelia sandrii (Brusina, 1865)	L23			
Mangelia secreta (Aartsen & Fehr de Wal, 1978, ex Mont. ms.)	L23			
Mangelia sicula Reeve, 1846	L23			
Mangelia taeniata (Deshayes, 1835)	L3		+	
Mangelia unifasciata (Deshayes, 1835)	L7		+	
Mangelia vauquelini (Payraudeau, 1826)	L9			
Taranis moerchi (Malm, 1863)	L24			
Mitrolumna olivoidea (Cantraine, 1835)	L3		+	
Raphitoma alternans (Monterosato, 1884)	L24			
Raphitoma cf. atropurpurea (Locard & Caziot, 1900, ex Mont. ms.				
Raphitoma bicolor (Risso, 1826)	L7			
Raphitoma concinna (Scacchi, 1836)	L9			
Raphitoma cordieri (Payraudeau, 1826)	L24			
Raphitoma echinata (Brocchi, 1814)	L9			
Raphitoma laviae (Philippi, 1844)	L23		+	
Raphitoma linearis (Montagu, 1803)	L9		+	
Raphitoma papillosa (Pallary, 1904)	L24			
Raphitoma philberti (Michaud, 1829)	L11		+	
Raphitoma pruinosa (Pallary, 1906)	L24			
Raphitoma pupoides (Monterosato, 1884)	L23			
Leufroya leufroyi (Michaud, 1828)	L11			
Comarmondia gracilis (Montagu, 1803) Teretia teres (Reeve, 1844)	L13 FR		+	12
Drilliidae	LIX		+	12
	L9			
Crassopleura incrassata (Dujardin, 1837) Mathildidae	Ly		+	
Mathilda gemmulata Semper, 1865	L9			
Rissoellidae	11)			
Rissoella inflata Locard, 1892	L11			
Omalogyridae	LII			
- Charles Addition				



Table 3 continued				
SPECIES	L	N	R	Z
Omalogyra atomus (Philippi, 1841)	L11			
Ammonicera fischeriana (Monterosato, 1869)	L11		+	
Pyramidellidae				
Chrysallida clathrata (Jeffreys, 1848)	L9			
Chrysallida decorata (Philippi, 1849)	L24	*17		
Chrysallida decussata (Montagu, 1803)	L24			
Chrysallida emaciata (Brusina, 1866)	L24			
Chrysallida indistincta (Montagu, 1808)	L11		+	
Chrysallida intermixta (Monterosato, 1884)	L11			
Chrysallida jeffreysiana (Monterosato, 1884, ex Seguenza G. ms.)	L11			
Chrysallida juliae (de Folin, 1872)	FR		+	13
Chrysallida limitum (Brusina, 1876)	L23			
Chrysallida obtusa (Brown T., 1827)	L11		+	
Chrysallida sp. B	L14			
Chrysallida suturalis (Philippi, 1844)	L23			
Chrysallida terebellum (Philippi, 1844)	L14			
Clathrella clathrata (Philippi, 1844)	L18			
Euparthenia bulinea (Lowe, 1841)	L9			
Euparthenia humboldti (Risso, 1826)	L11		+	
Folinella excavata (Philippi, 1836)	L9		+	
Folinella ghisottii Aartsen, 1984	FR		+	14
Odostomella doliolum (Philippi, 1844)	L9			
Tragula fenestrata (Jeffreys, 1848)	FR		+	15
Adelactaeon amoenus (Adams A., 1851)	L23	*		
Eulimella acicula (Philippi, 1836)	L23		+	
Eulimella bogii Aartsen, 1984	FR		+	16
Eulimella cerullii (Cossmann, 1915)	L15			
Eulimella ventricosa (Forbes, 1844)	L24			
"Syrnola" fasciata Jickeli, 1882	L22	*		
Odostomia carrozzai Aartsen, 1987	L11			
Odostomia erjaveciana Brusina, 1869	L23			
Odostomia imponderabilior Oberling, 1970	L23		+	
Odostomia lukisii Jeffreys, 1859	L11			
Odostomia nofronii Buzzurro, 2002	L29		+	
Odostomia plicata (Montagu, 1803)	L11			
Odostomia scalaris Mac Gillivray, 1843	L23			
Odostomia striolata Forbes & Hanley, 1850	L23			
Odostomia turriculata Monterosato, 1869	L23			
Odostomia turrita Hanley, 1844	L11			
Odostomia unidentata (Montagu, 1803)	L11		+	
Megastomia conoidea (Brocchi, 1814)	L9	18	+	
Megastomia sicula Philippi, 1851	L23	18		
Liostomia clavula (Lovén, 1846)	L11			
Ondina crystallina Locard, 1892	L23			
Ondina vitrea (Brusina, 1866)	L23			
Ondina warreni (Thompson, 1845)	L11			
Turbonilla acuta (Donovan, 1804)	L24			
Turbonilla acutissima Monterosato, 1884	FR		+	17
Turbonilla delicata Monterosato, 1874	L10			
Turbonilla edgari (Melvill, 1896)	L23	*		
Turbonilla gradata B. D. D., 1883	L23			
Turbonilla lactea (Linnaeus, 1758)	L2			
Turbonilla pumila Seguenza, G., 1876	L23			
Turbonilla pusilla (Philippi, 1844)	L10			
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Table J continued				
SPECIES	L	N	R	2
Turbonilla cf. rosewateri Corgan & Aartsen, 1993	L11			
Turbonilla rufa (Philippi, 1836)	L9		+	
Turbonilla sinuosa (Jeffreys, 1884)			т	
	L15			
Turbonilla striatula (Linnaeus, 1758)	L10		+	
Turbonilla cf. syrtensis Aartsen, 1981	L11			
Anisocyclidae				
Anisocycla folini (Fischer P. in de Folin, 1869)	L11	19		
Anisocycla nitidissima (Montagu, 1803)	L24			
Anisocycla pointeli (de Folin, 1867)	L11			
Acteonidae				
Acteon tornatilis (Linnaeus, 1758)	L2		+	
Crenilabium exile (Jeffreys, 1870, ex Forbes ms.)	L18	20		
Retusidae				
Pyrunculus fourierii (Audouin, 1826)	L23	*		
Retusa leptoeneilema (Brusina, 1866)	L11			
Retusa mammillata (Philippi, 1836)	L15			
Retusa truncatula (Bruguière, 1792)	L3		+	
Ventomnestia girardi (Audouin, 1826)	L23	*		
Cylichnina crebrisculpta Monterosato, 1884	L23			
Cylichnina laevisculpta (Granata-Grillo, 1877)	L23			
Cylichnina multiquadrata (Oberling, 1970)	L24	21		
Cylichnina umbilicata (Montagu, 1803)	L2			
Volvulella acuminata (Bruguière, 1792)	L9			
Ringiculidae	-/			
Ringicula auriculata (Ménard de la Groye, 1811)	L9			
Ringicula conformis Monterosato, 1877	L4		+	
Bullidae	Li		'	
Bulla striata Bruguière, 1792	L2		+	
Haminoeidae	L2		т	
	L2			
Haminoea hydatis (Linnaeus, 1758)			,	
Haminoea navicula (Da Costa, 1778)	L9			
Haminoea orbignyana (Férrussac, 1822)	L11			
Atys jeffreysi (Weinkauff, 1868)	L23		+	
Weinkauffia turgidula (Forbes, 1844)	L11		+	
Smaragdinellidae	T 00			
Smaragdinella algirae (Adams A. in Sowerby G. B. II, 1850)	L23			
Philinidae				
Philine aperta (Linnaeus, 1767)	L2		+	
Philine catena (Montagu, 1803)	L11			
Philine punctata (Adams J., 1800)	L10			
Aglajidae				
Philinopsis depicta (Renier, 1804)	L10			
Akeridae				
Akera bullata Müller O. F., 1776	L4			
Cylichnidae				
Cylichna crossei B. D. D., 1886	L10			
Cylichna cylindracea (Pennant, 1777)	L9		+	
Acteocina mucronata (Philippi, 1849)	L17	*		
Roxania utriculus (Brocchi, 1814)	L10			
Scaphander lignarius (Linnaeus, 1758)	L25		+	
Gastropteridae				
Gastropteron meckeli Kosse, 1813	L3			
Cavolinidae				
Cavolinia gibbosa gibbosa (D' Orbigny, 1835, ex Rang ms.)	L13			



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Lahi	P	4	continued

Table 3 continued				
SPECIES	L	N	R	Z
Cavolinia tridentata (Niebuhr, 1775, ex Forskâl ms.)	L9		+	
Clio cuspidata (Bosc, 1802)	L9			
Clio pyramidata lanceolata (Leseur, 1813)	L9			
Creseis acicula Rang, 1828	L13			
Hyalocylis striata (Rang, 1828)	L13			
Styliola subula (Quoy & Gaimard, 1827)	L10			
Limacinidae				
Limacina trochiformis (D' Orbigny, 1836)	L11			
Limacina inflata (D' Orbigny, 1836)	L11		+	
Oxynoidae	LII		T	
·	L4			
Oxynoe olivacea Rafinesque, 1814	L4 L25			
Lobiger serradifalci (Calcara, 1840)	L2)			
Volvatellidae	T 0 /			
Ascobulla fragilis (Jeffreys, 1856)	L24			
Elyssidae				
Elysia timida (Risso, 1818)	L10			
Thuridilla hopei (Vérany, 1853)	L4			
Umbraculidae				
Umbraculum umbraculum (Röding, 1798)	L2		+	
Tylodinidae				
Tylodina perversa (Gmelin, 1791)	L23			
Pleurobranchidae				
Pleurobranchus membranaceus (Montagu, 1815)	L9			
Berthella ocellata (Delle Chiaje, 1830)	L10			
Berthellina citrina (Rueppell & Leuckart, 1830)	L11			
Berthellina engeli Gardiner, 1936	L10			
Pleurobranchaea meckelii Meckel in Leue, 1813	L2			
Pleurobranchus testudinarius Cantraine, 1835	L10			
Aplysidae	LIU			
Aplysia depilans Gmelin, 1791	L2			
Aplysia parvula Guilding in Mörch, 1863	L10		+	
Aplysia punctata (Cuvier, 1803)	L10			
Aplysia fasciata Poiret, 1789	L4			
Dolabriferidae	T 0.2			
Petalifera gravieri (Vayssière, 1906)	L23			
Notarchus punctatus Philippi, 1836	L10			
Aegiretidae				
Aegires punctilucens (D' Orbigny, 1837)	L10			
Triophidae				
Kaloplocamus ramosus (Cantraine, 1835)	L10			
Chromodorididae				
Hypselodoris tricolor (Cantraine, 1835)	L4			
Hypselodoris villafranca (Risso, 1818)	L4			
Discodorididae				
Discodoris atromaculata (Bergh, 1880)	L2			
Dendrodorididae				
Dendrodoris grandiflora (Rapp, 1827)	L4			
Doriopsilla areolata Bergh, 1880	L10			
Tethyidae				
Tethys fimbria Linnaeus, 1767	L10			
Zephyrinidae				
Janolus cristatus (Delle Chiaje, 1841)	L4			
Flabellinidae				
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Tab	10 4	conti	nned

SPECIES	L	N	R	Z
Flabellina affinis (Gmelin, 1791)	L4			
Siphonariidae				
Siphonaria pectinata (Linnaeus, 1758)	L9			
Williamia gussonii (Costa O. G., 1829)	L11		+	
Trimusculidae				
Trimusculus mammillaris (Linnaeus, 1758)	L9			
Melampidae				
Auriculinella erosa (Jeffreys, 1830)	L18			
Ovatella firminii (Payraudeau, 1826)	L11		+	
Ovatella denticulata (Montagu, 1803)	L23			
Ovatella myosotis (Draparnaud, 1801)	L2			
BIVALVIA				
Solemyidae				
Solemya togata (Poli, 1791)	L9		+	
Nuculidae				
Nucula nitidosa Winckworth, 1931	L11		+	
Nucula nucleus (Linnaeus, 1758)	L3		+	
Nucula sulcata Bronn, 1831	L4		+	
Nuculoma tenuis (Montagu, 1808)	L11			
Nuculanidae 1750)	T 0			
Lembulus pellus (Linnaeus, 1758)	L3		+	
Saccella commutata (Philippi, 1844)	L3		+	
Arcidae	Т 1			
Arca noae Linnaeus, 1758	L1		+	
Arca tetragona Poli, 1795	L9			
Barbatia barbata (Linnaeus, 1758)	L1		+	
Acar plicata (Dillwyn, 1817)	L18			
Anadara corbuloides (Monterosato, 1878)	L9			
Anadara polii (Mayer, 1868)	L3 L24			
Bathyarca pectunculoides (Scacchi, 1834) Striarca lactea (Linnaeus, 1758)	L24 L3		,	
Glycymerididae	13		+	
Glycymeris bimaculata (Poli, 1795)	L9		+	
Glycymeris olimacutatu (1011, 1795) Glycymeris glycymeris (Linnaeus, 1758)	L3		+	
Glycymeris insubrica (Brocchi, 1814)	L3		+	
Mytilidae	1			
Mytilus galloprovincialis Lamarck, 1819	L3		+	
Brachidontes pharaonis (Fischer P., 1870)	L9	*	+	
Mytilaster minimus (Poli, 1795)	L11		+	
Crenella arenaria Monterosato, 1875, ex Martin H. ms.	L11		+	
Gregariella petagnae (Scacchi, 1832)	L9		+	
Modiolarca subpicta (Cantraine, 1835)	L4		+	
Musculus costulatus (Risso, 1826)	L4		+	
Lithophaga lithophaga (Linnaeus, 1758)	L3		+	
Modiolus barbatus (Linnaeus, 1758)	L3		+	
Modiolus adriaticus (Lamarck, 1819)	L 3		+	
Modiolula phaseolina (Philippi, 1844)	L20		+	
Pinnidae				
Pinna nobilis Linnaeus, 1758	L3		+	
Pteriidae				
Pteria hirundo (Linnaeus, 1758)	L4		+	
Pinctada radiata (Leach, 1814)	L3	*	+	
Malleidae				
Malleus regula (Forskâl, 1775)	L3	*22	+	



Table 3 continued				
SPECIES	L	N	R	Z
Malleus sp.	L23	*		
Pectinidae				
Pecten jacobaeus (Linnaeus, 1758)	L3		+	
Aequipecten opercularis (Linnaeus, 1758)	L7		+	
Pseudamussium septemradiatum (Müller O. F., 1776)	L3			
Pseudamussium clavatum (Poli, 1795)	L9		+	
Similpecten similis (Laskey, 1811)	L13			
Palliolum incomparabile (Risso, 1826)	L9		+	
Crassadoma multistriata (Poli, 1795)	L9			
Minachlamys varia (Linnaeus, 1758)	L3		+	
Manupecten pesfelis (Linnaeus, 1758)	L3			
Flexopecten flexuosus (Poli, 1795)	L3		+	
Flexopecten glaber (Linnaeus, 1758)	L9		+	
Lissopecten hyalinus (Poli, 1795)	L3			
Spondylidae				
Spondylus gaederopus Linnaeus, 1758	L3		+	
Anomiidae				
Anomia ephippium Linnaeus, 1758	L3		+	
Limidae				
Lima lima (Linnaeus, 1758)	L3		+	
Limaria hians (Gmelin, 1791)	L3		+	
Limaria loscombi (Sowerby G. B. I, 1823)	FR		+	18
Limaria tuberculata (Olivi, 1792)	L23			
Limatula gwyni (Sykes, 1903)	L9			
Limatula subauriculata (Montagu, 1808)	L18			
Ostreidae .				
Ostrea edulis Linnaeus, 1758	L3		+	
Ostreola stentina (Payraudeau, 1826)	L24			
Gryphaeidae				
Neopycnodonte cochlear (Poli, 1795)	L9			
Lucinidae				
Ctena decussata (Costa O. G., 1829)	L4		+	
Loripes lucinalis (Lamarck, 1818)	L3		+	
Megaxinus unguiculinus Pallary, 1904	L12			
Divaricella angulifera Von Martens, 1880	L16			
Lucinella divaricata (Linnaeus, 1758)	L9		+	
Anadontia fragilis (Philippi, 1836)	L9		+	
Myrtea spinifera (Montagu, 1803)	L3		+	
Thyasiridae				
Thyasira biplicata (Philippi,1836)	L3		+	
Thyasira planata (Jeffreys, 1882)	L25			
Axinulus croulinensis (Jeffreys, 1847)	L25			
Axinulus cycladius (Wood S., 1848)	L9			
Leptaxinus cf. subovatus (Jeffreys, 1881)	L24			
Ungulinidae				
Diplodonta apicalis Philippi, 1836	L23			
Chamidae				
Chama gryphoides Linnaeus, 1758	L3		+	
Pseudochama corbierei (Jonas, 1846)	L3 -	*		
Pseudochama gryphina (Lamarck, 1819)	L9		+	
Kelliidae				
Bornia sebetia (Costa O. G., 1829)	L9			
Montacutidae				
Tellimya ferruginosa (Montagu, 1808)	L25			



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SPECIES	L	N	R	Z
Carditidae				
Cardita calyculata (Linnaeus, 1758)	L3		+	
Glans trapezia (Linnaeus, 1767)	L3		+	
Glans elegans (Réquien, 1848)	L4		+	
Pteromeris jozinae Aartsen, 1984	L11			
Pteromeris minuta (Scacchi, 1836)	L23			
Cardites antiquatus (Linnaeus, 1758)	L3		+	
Astartidae				
Astarte fusca (Poli, 1791)	L3		+	
Astarte sulcata (Da Costa, 1778)	L9			
Cardiidae				
Acanthocardia aculeata (Linnaeus, 1758)	L4		+	
Acanthocardia echinata (Linnaeus, 1758)	L3		+	
Acanthocardia paucicostata (Sowerby G. B. II, 1834)	L1		+	
Acanthocardia tuberculata (Linnaeus, 1758)	L1		+	
Parvicardium exiguum (Gmelin, 1791)	L4		+	
Parvicardium exiguum (Gilletiii, 1791) Parvicardium ovale (Sowerby G. B. II, 1840)	L3		T	
	L18			
Parvicardium scabrum (Philippi, 1844)				
Parvicardium scriptum (B. D. D., 1892)	L11			
Parvicardium trapezium Cecalupo & Quadri, 1996	L23			
Plagiocardium papillosum (Poli, 1791)	L3		+	
Laevicardium crassum (Gmelin, 1791)	L9			
Laevicardium oblongum (Gmelin, 1791)	L3		+	
Cerastoderma edule (Linnaeus, 1758)	L3		+	
Cerastoderma glaucum (Poiret, 1789)	L3			
Fulvia fragilis (Forskâl in Niebuhr, 1775)	L16	*		
Mactridae				
Mactra stultorum (Linnaeus, 1758)	L3		+	
Mesodesmatidae				
Donacilla cornea (Poli, 1791)	L7		+	
Solenidae				
Solen marginatus Pulteney, 1799	L3		+	
Pharidae				
Ensis arcuatus (Jeffreys, 1865)	L18			
Ensis ensis (Linnaeus, 1758)	L3		+	
Ensis minor (Chenu, 1843)	L9		+	
Phaxas adriaticus (Coen, 1933)	L25			
Phaxas pellucidus (Pennant, 1777)	L25			
Tellinidae	1127			
Tellina compressa Brocchi, 1814	L23			
~	L25			
Tellina distorta Poli, 1791			,	
Tellina donacina Linnaeus, 1758	L3		+	
Tellina fabula Gmelin, 1791	L9			
Tellina incarnata Linnaeus, 1758	L3		+	
Tellina nitida Poli, 1791	L3		+	
Tellina pygmaea Lovén, 1846	L25			
Tellina planata Linnaeus, 1758	L7		+	
Tellina pulchella Lamarck, 1818	L3		+	
Tellina serrata Brocchi, 1814	L13		+	
Tellina tenuis Da Costa, 1778	L9			
Arcopagia balaustina (Linnaeus, 1758)	L3		+	
Macoma cf. balthica (Linnaeus, 1758)	L24	23		
Gastrana fragilis (Linnaeus, 1758)	L3		+	
Donacidae				



Table 3 continued				
SPECIES	L	N	R	Z
Donax semistriatus Poli, 1795	L3		+	
Donax venustus Poli, 1795	L3			
Donax trunculus Linnaeus, 1758	L3			
Psammobiidae				
Gari depressa (Pennant, 1777)	L7		+	
Semelidae				
Abra nitida (Müller O. F., 1776)	L18			
Abra prismatica (Montagu, 1808)	L25			
Abra segmentum (Récluz, 1843)	L25			
Abra tenuis (Montagu, 1803)	L3		+	
Abra alba (Wood W., 1802)	L9		+	
Abra longicallus (Scacchi, 1834)	- L25			
Solecurtidae				
Solecurtus strigilatus (Linnaeus, 1758)	L3		+	
Azorinus chamasolen (Da Costa, 1778)	L 3		+	
Kelliellidae				
Kelliella abyssicola (Forbes, 1844)	FR		+	19
Glossidae				
Glossus humanus (Linnaeus, 1758)	L3		+	
Veneridae				
Venus verrucosa Linnaeus, 1758	L3		+	
Venus casina Linnaeus, 1758	L9		+	
Globivenus effosa (Philippi, 1836, ex Bivona ms.)	L25			
Chamelea gallina (Linnaeus, 1758)	L3		+	
Clausinella fasciata (Da Costa, 1778)	L9		+	
Timoclea ovata (Pennant, 1777)	L3		+	
Gouldia minima (Montagu, 1803)	L3		+	
Dosinia lupinus (Linnaeus, 1758)	L3		+	
Dosinia exoleta (Linnaeus, 1758)	L9		+	
Pitar rudis (Poli, 1795)	L9		+	
Callista chione (Linnaeus, 1758)	L3		+	
Ruditapes decussatus (Linnaeus, 1758)	L1	24	+	
Irus irus (Linnaeus, 1758)	L3		+	
Paphia aurea (Gmelin, 1791)	L1		+	
Paphia lucens (Locard, 1886)	L1		+	
Paphia rhomboides (Pennant, 1777)	L9		+	
Venerupis senegalensis (Gmelin, 1791)	L3		+	
Petricolidae				
Petricola lithophaga (Retzius, 1786)	L3		+	
Petricola substriata (Montagu, 1808)	L9			
Petricola lajonkairii (Payraudeau, 1826)	L9		+	
Mysia undata (Pennant, 1777)	L9		+	
Corbulidae				
Corbula gibba (Olivi, 1792)	L7		+	
Gastrochaenidae				
Gastrochaena dubia (Pennant, 1777)	L7		+	
Hiatellidae				
Hiatella arctica (Linnaeus, 1767)	L9		+	
Pholadidae				
Pholas dactylus Linnaeus, 1758	L4		+	
Barnea candida (Linnaeus, 1758)	L3		+	
Teredinidae				
Teredo navalis Linnaeus, 1758	. L9			
Teredo utriculus Gmelin, 1791	L3			



Table 3 continued				
SPECIES	L	N	R	Z
Lyrodus pedicellatus (Quatrefages, 1849)	L13			
Nototeredo norvegica (Spengler, 1792)	L18			
Thraciidae				
Thracia papyracea (Poli, 1791)	L9			
Thracia pubescens (Pulteney, 1799)	L3			
Clavagellidae				
Clavagella melitensis Broderip, 1835	L9			
Pandoridae				
Pandora inaequivalvis (Linnaeus, 1758)	L9			
Lyonsiidae				
Lyonsia norwegica (Gmelin, 1791)	L25			
Cuspidariidae	227			
Cuspidaria cuspidata (Olivi, 1792)	L3		+	
Cuspidaria rostrata (Spengler, 1793)	L3		+	
Cardiomya costellata (Deshayes, 1835)	L3		+	
Caratoniya tostettara (Desnayes, 10)))	11)		'	
SCAPHOPODA				
Dentaliidae				
Antalis dentalis (Linnaeus, 1758)	L3			
Antalis entalis (Linnaeus, 1758)	L23			
Antalis panorma (Chenu, 1842)	L2)			
*	L4 L13			
Antalis rossati (Caprotti, 1966)	L ₁			
Antalis vulgaris (Da Costa, 1778) Fustiariidae	LZ		+	
	L2			
Fustiaria rubescens (Deshayes, 1826) Gadilidae	1.2		+	
	L18			
Dischides politus (Wood S., 1842) Entalinidae	LIO			
	L13			
Entalina tetragona (Brocchi, 1814)	LIJ			
CEPHALOPODA				
Sepiidae				
Sepita elegans Blainville 1827	L2			
Sepia officinalis Linnaeus, 1758	L2 L2			
Sepia orbignyana Férrussac in D' Orbigny, 1826	L26			
	120			
Sepiolidae Sepiolidae Noof 1012	L26			
Sepiola robusta Naef, 1912 Sepiola rondeleti Leach, 1817	L20			
•	L26			
Sepiola steenstrupiana Levy, 1912	L26			
Sepietta oweniana (D' Orbigny, 1840)	L26			
Sepietta neglecta Naef, 1916	120			
Loliginidae	L26			
Loligo forbesii Steenstrup, 1856	L20			
Loligo vulgaris Lamarck, 1798	L26			
Alloteuthis media (Linnaeus, 1758)	L20			
Ommastrephidae	Υ /4			
Illex coindetii (Vérany 1839)	L4			
Todaropsis eblanae (Ball, 1841)	L26			
Todarodes sagittatus (Lamarck, 1798)	L2			
Octopodidae Pi 1026	1.0			
Octopus macropus Risso, 1826	L2			
Octopus salutii Vérany, 1839	L26			
Octopus vulgaris Cuvier, 1798	L2			
Scaeurgus unicirrhus (Delle Chiaje in Férrussac & D' Orbigny, 1840)	L26			



SPECIES	L	N	R	Z
Eledone moschata (Lamarck, 1798)	L2			
Argonautidae				
Argonauta argo Linnaeus, 1758	L2			

Literatur: L1: Monterosato, 1899; L2: Demetropoulos, 1969; L3: Demetropoulos, 1971; L4: Demetropoulos & Hadjichristophorou, 1976; L5: Barash & Danin, 1977; L6: Verduin, 1984; L7: Stavrinos, 1984; L8: Verduin, 1986; L9: Tornaritis, 1987; L10: Barash & Danin, 1989; L11: Bogi et al., 1989; L12: Aartsen & Kinzelbach, 1990; L13: Barash & Danin, 1992; L14: Linden & Eikenboom, 1992; L15: Fischer, 1993a; L16: Fischer, 1993b; L17: Cecalupo & Quadri, 1994; L18: Fischer, 1994; L19: Oliverio & Buzzurro, 1994; L20: Oliverio et al., 1994; L21: Cecalupo & Quadri, 1995; L22: Nofroni & Tringali, 1995; L23: Cecalupo & Quadri, 1996; L24: Buzzurro & Greppi, 1997; L25: Hadjichristophorou et al., 1997; L26: Salman et al., 1998; L27: Smriglio et al., 1998; L28: Houart, 2001, L29: Buzzurro, 2002.

NOTES

1: Lepidopleurus boettgeri Sulc, 1934 for the claimed synonymy with L. bedulli (Dell'Angelo & Palazzi, 1986), reported by CECALUPO & QUADRI (1996), see DELL'ANGELO & SMRIGLIO (1999).

2: Acanthochitona aeneus (Risso, 1826), reported by BOGI et al. (1989) is hard to consider, since the species belonging to this genus are only two according to DELL'ANGELO & SMRIGLIO (1999) and are already present in the checklist, so we prefer not to report it.

3: Acanthochitona crinita oblonga Leloup, 1981 reported by CECALUPO & QUADRI (1996) has been excluded: see DELL'ANGELO & SMRIGLIO (1999).

4: this is the correct name considering that *Haliotis lamellosa* Lamarck, 1822 is a synonym of *H. tuberculata*: see GEIGER (1998).

5: several Authors have reported this species, while others have identified it as *Gibbula umbilicaris* (Linnaeus, 1758). Anatomical studies are needed to definitively solved this taxonomical problem. Due to the lack of data, we are conservatively in line with the interpretation of Monterosato (1899: 398), who wrote: "Forme peu commune, voisine du G. umbilicaris Linné, dont elle diffère par sa forme conique, par ses taches suturales blanches et son ombilic plus étroit".

6: following REID (1996) we use *Melarhaphe* for *neritoides* (Linnaeus, 1758), the type and only living member of this genus.

7: BOGI et al. (1989) reported a record of this species by TORNARITIS (1987), who has nevertheless not reported it in his work.

8: the origin of *Alvania dorbigny* (Audouin, 1827) is still debated by nowadays Authors: we actually follow the opinion of MIENIS (1985) considering this species as non lessepsian.

9: GIANNUZZI-SAVELLI et al. (1997: 15) ascribed this species to the genus *Rudolphosetia* Monterosato, 1917: we follow PONDER (1985: 34) in considering *Rudolphosetia* as a *junior* synonym of *Setia* (H. & A. Adams, 1852).

10: according to Abbott (1974: 139) Calyptraeidae de Blainville, 1824 is a junior synonym of Crepulidae Fleming, 1822

11: Erronea caurica (Linnaeus, 1758) reported by GIANNUZZI-SAVELLI et al. (1997) has been excluded: see BUZZURRO & GREP-

PI (1997).

12: BEU (1998) has demonstrated that the Mediterranean form related to *Charonia tritonis* represents a distinct species instead of a subspecies.

13: Vexillum hypatiae (Pallary, 1912), reported by CECALUPO & QUADRI (1996) is not listed according to TRINGALI & VILLA (1991).

14: for the claimed synonymy reported by CECALUPO & QUADRI (1996) with *Granulina clandestina* (Brocchi, 1814) see GOFAS (1992).

15: in the past it has been identified by many Authors as *Clathromangelia quadrillum* (Dujardin, 1837), but this is a fossil species (OLIVERIO, 1995).

16: Mangelia fieldeni (Aartsen & Fehr de Wal, 1978) = M. angelinae (Cecalupo & Quadri, 1996): see BUZZURRO & GREPPI (1997).

17: Chrysallida decorata (Philippi, 1849) is the correct name for the taxon commonly reported by several Authors as Chrysallida maiae (Hornung & Mermod, 1924) (BUZZURRO & GREPPI, unpublished data).

18: the genus has been assigned following SCHANDER & SUNDBERG (1997).

19: probably the record referred to Anisocycla nitidissima (Montagu, 1803).

20: reported by FISCHER (1994) from shallow water of the littoral zone, this is the only record of this deep-sea species, probably it is a case of misidentification with *Caecilianella acicula* Müller, 1774, a terrestrial species commonly found in our samples: see GAGLINI (1982).

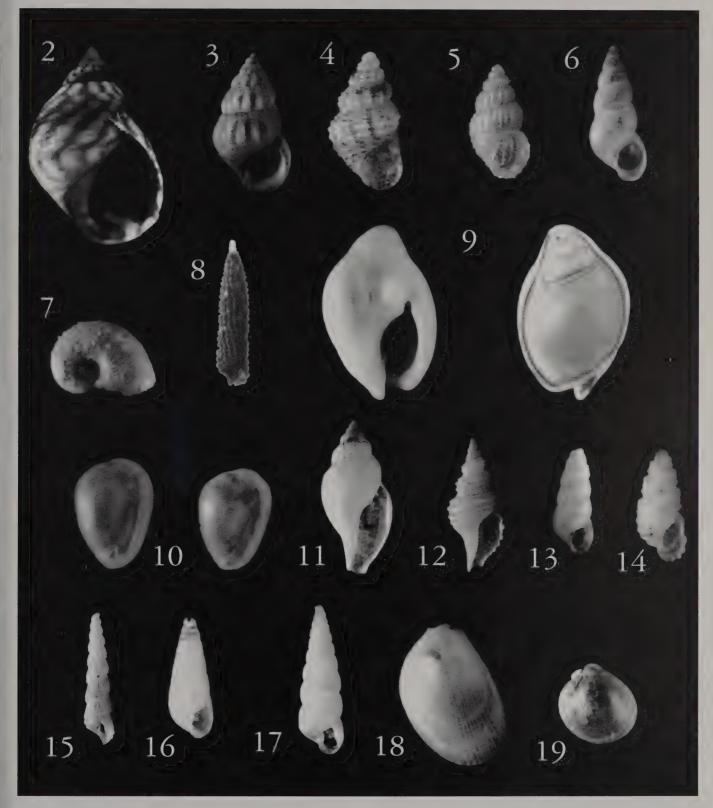
21: for the putative finding of *Cylichnina* cf. *mongii* (Audouin, 1827) reported by CECALUPO & QUADRI (1996) see BUZZURRO & GREPPI (1997).

22: the name of this species comes from the Latin word regula(-ae) = a ruler [and not from regulus(-i) = a small king!].

23: for the presence of *Macoma balthica* (Linnaeus, 1758) in the Eastern Mediterranean Sea we refer to BUZZURRO (unpublished data)

24: European Authors traditionally refer *decussatus* to *Tapes*, whereas they currently use *Ruditapes* for the Japanese carpet shell (*R. philippinarum* (Adams & Reeve, 1850), which has been imported for acquaculture since a number of years. Both species





Figs 2-19. Gastropods and bivalves from Cyprus. 2: Nodilittorina (Echinolittorina) punctata (Gmelin, 1791) mm 6.6: St. C7. 3: Rissoa splendida Eichwald, 1830 mm 4.8: St. C8. 4: Alvania fractospira Oberling, 1970 mm 2.5: St. C10. 5: Alvania testae (Aradas & Maggiore, 1844) mm 2.1: St. D10. 6: Hydrobia acuta (Draparnaud, 1805) mm 3.2: St. C11. 7: Protatlanta souleyeti (Smith E. A., 1888) mm 2.1: St. D17. 8: Cerithiopsis scalaris Locard, 1892, ex Monterosato ms. mm 4.2: St. C8. 9: Nassarius circumcictus (Adams A., 1852) mm 15: St. C10. 10: Granulina boucheti Gofas, 1992 mm 2.2: St. C8. 11: Gymnobela subaraneosa (Dautzenberg & Fischer H., 1896) mm 5.4: St. D17. 12: Teretia teres (Reeve, 1844) mm 4.3: St. D17. 13: Chrysallida juliae (de Folin, 1872) mm 1.9: St. C8. 14: Folinella ghisottii Aartsen, 1984 mm 2.5: St. C8. 15: Tragula fenestrata (Jeffreys, 1848) mm 3.1: St. C8. 16: Eulimella bogii Aartsen, 1994 mm 1.9: St. C8. 17: Turbonilla acutissima Monterosato, 1884 mm 5.3: St. D21. 18: Limaria loscombi (Sowerby G. B. I, 1823) mm 8.9: St. C10. 19: Kelliella abyssicola (Forbes, 1844) mm 1.6: St. D16



belong to *Ruditapes*, as proposed by Fischer-Piette & Métiver (1971) and Matsukuma (1986); in particular, the last author excludes *Ruditapes* from *Tapes*.

25: Alvania sp., under study.

New records of the molluscs fauna of Cyprus

LITTORINIDAE

Nodilittorina (Echinolittorina) punctata (Gmelin, 1791) (Fig. 2)

Turbo punctatus Gmelin, 1791: 3597

Littorina syriaca Philippi, 1847: 165 (23), pl. 3, figs 21-22

Turbo siculus Chiereghin in NARDO, 1847: 78, sp. 20, figs 909-910

Turbo siculus Brugnone in Monterosato, 1872: 33 (nomen nudum)

Littorina punctata, Gmelin in Monterosato, 1872: 33

Littorina (Melaraphe) punctata (Gmelin, 1791) in Priolo, 1953: 124

Littorina punctata (Gmelin, 1791) = Tricolia draparnauldii Audouin, 1826 = Littorina pulchella Dunker, 1853 = Littorina syriaca Philippi, 1847 = Littorina guttata Reeve, 1858 = Littorina perplexa Turton, 1932 in ROSEWATER, 1970: 474

Nodilittorina (Echinolittorina) punctata (Gmelin, 1791) in Reid, 1996: 99.

Remarks: The allocation in the genus *Nodilittorina* von Martens, 1897 recently proposed by REID (1996) for this species is surprising because *Nodilittorina*, type species *L. pyramidalis* Quoy & Gaimard, 1833, which is characterised by a strong ribbed sculpture with nodules, includes mainly Indo-pacific species with very different shell. The modern systematics of the family Littorinidae is primarily based on anatomical characters of the penis, pallial oviduct, radula, sperm nurse cells or spawn. See REID (1986, 1989, 1996).

Material: St. C7, C12, 19 specimens, Supralittoral Zone, rocky habitats.

Distribution: Discontinuous distribution in Mediterranean basin, Portugal and African coasts of Atlantic Ocean.

RISSOIDAE

Rissoa splendida Eichwald, 1830

(Fig. 3)

Rissoa splendida Eichwald, 1830: 219

Rissoa violaestoma Krynicky, 1837 fide CLEMAM

Remarks: Rissoa splendida Eichwald, 1830 figured by GIAN-NUZZI-SAVELLI et al., 1997: 74, figs 203-204.

Material: St. C8, 1 specimen, *Cystoseira* sp., 2-3 m and St. C10, 1 shell, grit near *Posidonia oceanica*, 4 m.

Distribution: Aegean Sea, Central Mediterranean Sea, Adriatic Sea, the Marmara Sea and Black Sea. This species is here reported for the first time from the Levantine Basin.

Alvania fractospira Oberling, 1970

(Fig. 4)

Alvania fractospira Oberling, 1970: 3

Alvania monterosatoi sensu BOGI et al., 1989: 194, fig. 7 (not Fis-

CHER P., 1877)

Remarks: *Alvania fractospira* Oberling, 1970 in MOOLENBEEK *et al.*, 1991: 113, fig. 16 (the lectotype is selected) and in OLIVERIO, 1993: 33 (restored as a valid name).

Material: St. C1, 3 specimens, *Posidonia oceanica*, 5 m; St. C8, 2 shells in grit near *Posidonia oceanica*, 4 m and St. C10, 2 specimens, *Cystoseira* sp., 1-2 m.

Distribution: Eastern Mediterranean Sea and Adriatic Sea (GIANNUZZI-SAVELLI *et al.*, 1997).

Alvania testae (Aradas & Maggiore, 1844)

(Fig. 5)

Rissoa reticulata Philippi, 1836: 156, tav. 10, fig. 14 (not T. reticulata Montagu, 1803)

Rissoa testae Aradas & Maggiore, 1844: 135

Rissoa abyssicola Forbes in FORBES & HANLEY, 1850: 86

Rissoa partschii Hörnes, 1857: 573 (fossil)

Rissoa (Alvania) testae (Aradas & Maggiore, 1844) in ARADAS & BENOIT, 1870a: 199

Rissoa abyssicola var. conformis Jeffreys, 1870: 77

Rissoa (Actonia) testae Aradas & Maggiore in CARUS, 1889: 336
Actonia testae (Aradas & Maggiore, 1844) in MONTEROSATO,
1890: 147

Alvania testae (Aradas & Maggiore, 1844) in LOCARD, 1892: 161 Alvania (Actonia) testae (Aradas & Maggiore) in THIELE, 1931: 163

Turbona testae (Aradas & Maggiore, 1844) in NORDSIECK, 1972: 186

Remarks: Alvania testae (Aradas & Maggiore, 1844) in BOUCHET & WARÉN, 1993: 628, figs 1386-1387, 1400-1405 and in OLIVERIO et al., 1992: 251 (A. testae complex is here analyzed and discussed).

Material: St. D10 (35°34'8"N-34°28'3"E), 3 shells, muddy habitats, 210 m, dredged.

Distribution: Eastern Mediterranean Sea (Aegean Sea), Western Mediterranean Sea and coastal line of the Eastern Atlantic Ocean from Morocco to Norway.

HYDROBIIDAE

Hydrobia acuta (Draparnaud, 1805)

(Fig. 6)

Cyclostoma acutum Draparnaud, 1805: 40, pl. I, fig. 23

Paludestrina acuta Draparnaud in Monterosato, 1878a: 86

Paludestrina glyca Servain, 1880: 151

Hydrobia acuta (Draparnaud, 1805) in GIUSTI & PEZZOLI (1984: 125, tav. 1)

Remarks: Hydrobia acuta (Draparnaud, 1805) figured by GIUSTI et al., 1995: 116, 117, figs 39-42, 50-52. After BOETERS (1984) that designated a lectotype for this species from two syntypes, GIUSTI et al. (1998) demonstrated that they to belong to different species and designate a neotype "in line with the earlier and more widely accepted usage of the name" (1998: 2, figs 3-6).

Material: St. C11, 5 shells, grit, 1 m.

Distribution: Whole Mediterranean basin (BARASH & DANIN, 1992).

ATLANTIDAE



Protatlanta souleyeti (Smith E. A., 1888)

(Fig. 7)

Atlanta souleyeti Smith E. A., 1888: 1-51, pl. XII, fig. 7a, b

Protatlanta souleyeti (Smith E. A., 1888) = Atlanta lamanonii Souleyet, 1852 = Protatlanta sculpta Issel, 1911 = Protatlanta sculpta var. mediterranea Issel, 1911 = Protatlanta mediterranea (Issel) Di Geronimo, 1970 in VATOVA, 1975: 1

Remarks: *Protatlanta souleyeti* (Smith E. A., 1888) figured by GIANNUZZI-SAVELLI *et al.*, 1997: 248, figs 921-923.

Material: St. D17 (35°43'8"N-34°37'3"E), 1 shell, 250-300 m dredged.

Distribution: It has a cosmopolitan distribution due to its pelagic life style, similar to other members of the family. Southern Aegean Sea, Central Mediterranean and Western Mediterranean Sea. It is here reported for the first time in the Levantine basin.

CERITHIOPSIDAE

Cerithiopsis scalaris Locard, 1892, ex Monterosato ms.

Cerithiopsis scalaris Monterosato, 1878a = C. corona var. scalaris Monterosato: 99 (nomen nudum)

Cerithiopsis scalaris Monterosato, 1878 in LOCARD, 1892: 118 Cerithiopsis scalaris Coen, 1937: 31, tav. 3, fig. 26 (nomen nudum) Cerithiopsis scalaris (Monterosato, 1877) in Nordsieck, 1976: 7;

Cerithiopsis scalaris Monterosato, 1877 in Piani, 1980: 136 Cerithiopsis scalaris Monterosato, 1878 (nomen nudum) in GAGLINI, 1992: 148, fig. 139

Cerithiopsis scalaris Locard, 1892, ex Monterosato ms. in Tringali, 1999: 129, 130: figs 1-3

Remarks: Cerithiopsis scalaris Locard, 1892 figured by GIAN-NUZZI-SAVELLI et al., 1999: 41, figs 60-63.

Material: St. C8, 2 shells, grit near Posidonia oceanica, 4 m.

Distribution: Central Mediterranean Sea. It is here reported from the Eastern Mediterranean Sea for the first time.

NASSARIIDAE

Nassarius circumcictus (Adams A., 1852)

(Fig. 9)

Nassa circumcincta Adams A., 1852: 102.

Arcularia circumcincta Adams A. in PALLARY, 1912: 96, pl.XV (I), figg. 14-17

Nassarius (Plicarcularia) circumcictus (A. Adams, 1851) in PIANI, 1980: 149

Nassarius (Plicarcularia) circumcictus (Adams A., 1852) = Naytiopsis granum flammulata Nordsieck, 1972 in Nofroni, 1986: 13

Remarks: Arcularia circumcincta (Adams A., 1851) in SABELLI & SPADA, 1977: 3, figs 5a-5b and Nassarius circumcictus (Adams A., 1851) in OLIVERIO & TRINGALI, 1992: 159, figs 4-8.

Material: St. C10, 2 specimens, sandy habitat, 2 m.

Distribution: Eastern Mediterranean Sea.

CYSTISCIDAE

Granulina boucheti Gofas, 1992

(Fig. 10)

Granulina boucheti Gofas, 1992: 10, fig. 9; 11, fig. 10; 24, fig. 26. Material: St. C8, 2 specimens, alive collected in brown algae, 1 m. Distribution: Sicily, France, Corse, Tunisie and Algerie. It is here reported from the Levantine basin for the first time.

CONIDAE

Gymnobela subaraneosa (Dautzenberg & Fischer H., 1896) (Fig. 11)

Pleurotoma subaraneosa Dautzenberg & Fischer H., 1896: 422, pl. 16, figs 11-12

Gymnobela subaraneosa (Dautzenberg & Fischer H., 1896) in BOUCHET & WARÉN, 1980: 58, figs 114-115

Pleurotomella suharaneosa (Dautzenberg & Fischer H., 1896) in POWELL, 1966: 131.

Material: St. D17 (35°43'8"N-34°37'3"E), 1 shell, muddy bottoms, 250-300 m, dredged.

Distribution: Mediterranean Sea and North Eastern Atlantic Ocean. Reported once from the Eastern Mediterranean Sea (JANSSEN, 1989).

Teretia teres (Reeve, 1844)

(Fig. 12)

Pleurotoma teres Reeve, 1844: 190

Philbertia anceps (Eichwald, 1830) (fossil): 225

Pleurotoma boreale Phil., ms. in LOVÉN, 1846: 14

Pleurotoma fusiforme Réquien, 1848: 101

Pleurotoma anceps in LOCARD, 1866: 109

Raphitoma barbierii Brusina, 1866: 33

Pleurotoma teres Forbes in Monterosato, 1870: 45

Defrancia (Pleurotoma) teres Forbes in Monterosato, 1872: 51

Pleurotoma (Defrancia) anceps Eichwald in Monterosato, 1875: 44

Clathurella anceps V. Crs. in CARUS, 1889: 427

Teretia teres Forbes in MONTEROSATO, 1890: 187

Teretia anceps (Eichwald, 1830) in PIANI, 1980: 157

Teretia teres (Reeve, 1844) in SABELLI et al., 1990: 45

Remarks: Teretia teres (Forbes, 1844) figured by BOUCHET & WARÉN, 1980: 81, fig. 168 and GIRIBET & PEÑAS, 1997: 75, fig. 69.

Material: St. D17 (35°43'8"N-34°37'3"E), 1 shell, muddy bottoms, 250-300 m, dredged.

Distribution: Coastline from Gibraltar Strait to Northern Norway, Western Mediterranean Sea, Adriatic Sea and Aegean Sea. It is here reported from the Levantine basin for the first time.

PYRAMIDELLIDAE

Chrysallida juliae (de Folin, 1872)

(Fig. 13)

Truncatella juliae de Folin, 1872: 49, pl. II, fig. 4

Odostomia (Pyrgulina) nanodea Monterosato, 1878a: 93

Parthenina tenuistriata Milachewitch, 1909: 314

Chrysallida sarsi Nordsieck, 1972: 98, pl. II, fig. 4 in Peñas et al., 1996: 24

Chrysallida juliae (de Folin, 1872) in AARTSEN, 1977: 61, fig. 13. Remarks: Chrysallida juliae (de Folin, 1872) in MICALI et al. (1993: 150): it is there designated the lectotype. Regarding all



the synonyms listed above we are waiting for the revision of these taxa accomplished by I. Nofroni (personal communication, manuscript in preparation).

Material: St. C8, 3 shells, grit near *Posidonia oceanica*, 4 m. Distribution: Mediterranean Sea and Atlantic Ocean including the Canary Islands.

Folinella ghisottii Aartsen, 1984

(Fig. 14)

Odostomia intermedia Brusina, 1869: 237 (non Deshayes, 1861) Odostomia (Pyrgulina) intermedia Brusina in MONTEROSATO, 1878b: 317

Folinella ghisottii Aartsen, 1984: 137:

Material: St. C8, 1 shell, grit near *Posidonia oceanica*, 4 m. Distribution: Mediterranean Sea and Atlantic Ocean including the Cape Verde Islands.

Tragula fenestrata (Jeffreys, 1848)

(Fig. 15)

Odostomia fenestrata Jeffreys, 1848: 345, pl. 6, fig. 17
Chemnitzia fenestrata Jeffreys in Forbes & Hanley, 1853: 249
Odostomia (Pyrgulina) fenestrata, Forbes in Monterosato, 1877: 35
Tragula fenestrata (Jeffreys, 1848) in Monterosato, 1884: 86

Odostomia (Tragula) fenestrata Forbes in PALLARY, 1912: 132, pl. XVI (II), fig. 42

Turbonilla fenestrata (Forbes) in Babio & Thiriot-Quievreux, 1974: 544, pl. VI, H, L

Tragula fenestrata (Forbes, 1848) in GROSSU, 1986: 24 Chrysallida fenestrata (Forbes in Jeffreys, 1848) in AARTSEN, 1981: 80, pl. 1, fig. 1

Chrysallida fenestrata (Jeffreys, 1848) in LINDEN & EIKENBOOM, 1992: 45

Remarks: C. fenestrata (Jeffreys, 1848) figured by Peñas et al., 1996: 27, fig. 54.

Material: St. C8, 1 shell, grit near *Posidonia oceanica*, 4 m. Distribution: Mediterranean Sea and Atlantic Ocean including Corimba.

Eulimella bogii Aartsen, 1994

(Fig. 16)

Eulimella bogii Aartsen, 1994: 89-90, fig. 5.

Material: St. C8, 1 shell, near Posidonia oceanica, 4 m.

Distribution: Mediterranean Sea including the Eastern basin (Nofroni & Tringali, 1995).

Turbonilla acutissima Monterosato, 1884

(Fig. 17)

Turbonilla acutissima Monterosato, 1884: 92

Remarks: Turbonilla acutissima Monterosato, 1884 figured by AARTSEN, 1981: 85, pl. 4, fig. 24 and Peñas et al., 1996: 69, fig. 185.

Material: St. D21(35°09'0"N-33°57'0"E), 1 shell, *Posidonia oceanica*, 20 m, dredge.

Distribution: Mediterranean Sea (POPPE & GOTO, 1990).

LIMIDAE

Limaria loscombi (Sowerby G. B. I, 1823)

(Fig. 18)

Lima loscombi Sowerby G. B. I, 1823: 17, pl. 113, fig. 4

Limea loscombei Sowerby in ARADAS & BENOIT, 1870b: 94

Lima (Limatella) loscombei Leach in Sowerby in Anderson, 1964:

144, figs 27a-c

Lima (Limea) loscombii Sowerby G. B., 1823 in Altena Regteren et al., 1969: 24 (fossil)

Lima (Mantellum) loscombi Sowerby = Lima bullata Turton = Lima clausa Danilo & Sandri in Musetti, 1969: 52

Limaria (Limatulella) loscombi (Sowerby G. B., 1830) in JANSSEN, 1972: 16, 74, figs 4-5 (fossil)

Limea (Limea) loscombi MacGillivray, 1843 in Piani, 1980: 187 Limea loscombi (Sowerby G. B. I, 1823) in Sabelli et al., 1990: 85 Limea (Limatulella) loscombi (Sowerby G. B. I, 1823) in Zenetos, 1996: 88

Limaria loscombii (Sowerby G. B. I, 1823) in SALAS, 1996: 57. Material: St. C10, 1 specimen, muddy–sandy habitats, 1.5 - 2 m. Distribution: This species is common in muddy and muddy-sandy bottoms down to a depth of 3000 m (PARENZAN, 1974), distributed in the Aegean Sea, Adriatic Sea, Western Mediterranean Sea and the coastline of the Eastern Atlantic Ocean from Senegal and to Norway (Jeffreys, 1879; Tebble, 1976; Barash & Danin, 1992).

KELLIELLIDAE

Kelliella abyssicola (Forbes, 1844)

(Fig. 19)

Kellia abyssicola Forbes, 1844: 122

Venus miliaris Philippi, 1844: 36, pl. 14, fig. 15

Kelliella abyssicola Sars in Monterosato, 1870: 39

Kelliella miliaris Philippi in SARS, 1878: 65

Kelliella miliaris Philippi in Monterosato, 1880: 59

Kellyella miliaris (Phil.) in PARENZAN, 1974: 154, 265, fig. 158

Kelliella abyssicola (Forbes, 1844) in SABELLI et al., 1990: 95.

Material: St. D16 (35°19'9"N-32°50'1"E), 1 specimen, muddy habitats, 210 m, dredged.

Distribution: Aegean Sea, Taranto Gulf and Northern Norway coasts (PARENZAN, 1974; POPPE & GOTO, 1993; ZENETOS, 1996). Already reported from the Eastern Mediterranean Sea by Janssen (1989). This species shows a vertical distribution from the continental shelf down to the abyssal zone.

DISCUSSION

There has been a significant increase in the number of molluscan species reported from Cyprus since the first study carried out by Monterosato (1899). In fact, in his report, an initial amount of 29 species was listed. But the number has grown up to 696 to date. Particularly, the studies of Bogi et al. (1989), Fischer (1994), Cecalupo & Quadri (1996) and Buzzurro & Greppi (1997) made important contribution to such increase. Bogi et al. (1989), reported 221 species in total, 89 as new records from this island. Then, Fischer (1994) gave



a list of 123 fossils and 434 living species for Cyprus. This Author, following SABELLI et al. (1990-1992), has reconsidered the systematic positions of several species reported by Demetropoulos (1969, 1971), Demetropoulos & Had-JICHRISTOPHOROU (1976), STAVRINOS (1984) and TORNARITIS (1987). Since there are many different views concerning classification, at the mean time FISCHER (1994) followed his personal conception regarding some cases of synonymy, changing the taxonomical rank of some species. For example, while Emarginula cancellata Philippi, 1836 and Emarginula sicula Gray J. E., 1825 are two synonyms according to SABELLI et al. (1990), on the contrary FISCHER (1994: 87) reported these two taxa as distinct species. In our opinion the sole valid species name is E. sicula. Other controversial cases of synonymy that we had to face making the new checklist are discussed below. Bogi et al. (1989) reported Pirenella conica (Blainville, 1826) and Pirenella cailliaudi (Potiez & Michaud, 1838) as different species, while the latter should be considered just as junior synonym of the first one according to SABELLI et al. (1990-1992). In like manner, Lima lima (Linnaeus, 1758) and Lima inflata (Chemnitz, 1784) should be thought as synonyms, taking as valid species name L. lima. Similarly, MICALI et al. (1993) considered Chrysallida cf. delpretei (Sulliotti, 1889) as junior synonym of Chrysallyda indistincta (Montagu, 1808): both taxa are listed as separate species in BOGI et al. (1989). An another example of systematic confusion is the case of Retusa semisulcata (Philippi, 1836) and Retusa truncatula (Brugière, 1792), both listed in CECALUPO & QUADRI (1996), but re-discussed as synonyms by TRINGALI (1995).

Of a total of 389 species listed by CECALUPO & QUADRI (1996), 134 species were new ones for Cyprus. In their papers, BUZZURRO & GREPPI (1997) and SALMAN et al. (1998) have reported 42 and 10 additional new species, respectively, for the Cyprus coasts. When the lists of BOGI et al. (1989) and CECALUPO & QUADRI (1996) are compared, 6 and 24 species, respectively, given as new records had already been reported previously (Table 4). Additionally, Argonauta argo Linnaeus, 1758 reported by SALMAN et al. (1998) as a new record, was

Table 4. Species previously listed by BOGI et al. (1989) (*) and CECALUPO & QUADRI (1996) (**) as new records and the literature relevant to these species.

0	
100	20100
SPC	ecies

Ischnochiton rissoi (Payraudeau, 1826) *

Lepidochitona corrugata (Reeve, 1848) *

Chiton corallinus (Risso, 1826)**

Gibbula umbilicaris (Linnaeus, 1758)**

Cerithium scabridum Philippi, 1848 **

Rissoa auriformis pseudomonodonta (Verduin, 1983)**

Alvania datchaensis Amati & Oliverio, 1987 **

Alvania mamillata Risso, 1826 **

Alvania beniamina (Monterosato, 1884) **

Strombus persicus Swainson, 1821 **

Petaloconchus glomeratus (Linnaeus, 1758) **

Vitreolina levantina Oliverio, Buzzurro & Villa, 1994 **

Vexillum littorale (Forbes, 1844) *

Bela laevigata (Philippi, 1836) **

Bela menkhorsti Aartsen, 1988 **

Bela nebula (Montagu, 1803) *

Mitrolumna olivoidea (Cantraine, 1835)*

Chrysallida terebellum (Philippi, 1844)**

Turbonilla delicata (Monterosato, 1874) **

Turbonilla striatula (Linnaeus, 1758)**

Retusa mamillata (Philippi, 1836) **

Retusa semisulcata (Philippi, 1836) **

Cylichnina umbilicata (Montagu, 1803) *

Cylichna crossei (B.D.D., 1886) **

Cavolinia tridentata (Niebuhr, 1775 ex Forskäl, ms.) **

Styliola subula (Quoy & Gaimard, 1827) **

Modiolarca subpicta (Cantraine, 1835)*

Lucinella divaricata (Linnaeus, 1758) **

Cerastoderma edule (Linnaeus, 1758) **

Abra nitida (Müller O. F., 1776) **

Gastrochaena dubia (Pennant, 1777) **

Literature giving former records

Barash & Danin (1977: 7)

Barash & Danin (1977: 9)

BARASH & DANIN (1977: 12)

FISCHER (1993A: 38)

FISCHER (1993A: 39)

AARTSEN & KINZELBACH (1990: 109)

AARTSEN & KINZELBACH (1990: 109)

VERDUIN (1986: 30)

VERDUIN (1984: 49)

FISCHER (1993A: 39)

STAVRINOS (1984: 21)

OLIVERIO et al. (1994: 214)

Tornaritis (1987: 103)

FISCHER (1993A: 40)

FISCHER (1993A: 40)

Demetropoulos (1971: 20)

Demetropoulos (1971: 16)

Linden & Eikenboom (1992: 18)

Barash & Danin (1989: 248)

Barash & Danin (1989: 248)

FISCHER (1993A: 40)

FISCHER (1993A: 40)

Demetropoulos (1969: 13)

Barash & Danin (1989: 250)

TORNARITIS (1987: 114)

Barash & Danin (1989: 253)

Demetropoulos & Hadjichristophorou (1971: 79)

FISCHER (1994: 93)

DEMETROPOULOS (1971: 21)

FISCHER (1994: 93)

FISCHER (1994: 93)



previously reported by DEMETROPOULOS (1969). Besides that, *Anachis troglodytes* (Souverbie & Montrouzier, 1866) and *Anachis savignyi* Moazzo, 1939 ex Jousseaume ms. reported as synonyms by CECALUPO & QUADRI (1996), are considered as two distinct species according to TRINGALI & VILLA (1995).

Furthermore, although CECALUPO & QUADRI (1996) discussed that some of the species which had been reported as synonyms by SABELLI et al. (1990) can be accepted as valid taxa, species such as Emarginula crebrisculpta (Coen, 1939), Calliostoma alexandrinum Pallary, 1911, Muricopsis inermis (Philippi, 1836), Mangelia derelicta Reeve, 1846, Raphitoma intermedia Nordsieck, 1977 and Raphitoma lineolata (B.D.D., 1883) are not valid species according to the CLEMAM database.

Some of the species names found in literature referred to the Island of Cyprus have been omitted from the checklist because fossils: Alvania monterosatoi (Fischer P., 1876), Granulina clandestina (Brocchi, 1814), Clathromangelia quadrillum (Dujardin, 1837), Raphitoma histrix Bellardi, 1847, Odostomia bismichaelis Sacco, 1892 and Odostomia cf. clavulina Fischer P., 1877. Also Hydrobia ulvae (Pennant, 1777), most probably confused with other Hydrobia sp., has been deleted since we reasonably think it is not a Mediterranean species. Other cases of shell misidentification reported by TORNARITIS (1987) are: Cerithiopsis tubercularis (Montagu, 1803) which is Dizoniopsis coppolae (Aradas, 1870) (: 58, fig. 80b), Fusinus pulchellus (Philippi, 1844) is just a juvenile form of F. syracusanus (Linnaeus, 1758) (: 98, fig. 146), Mathilda cochlaeformis Brugnone, 1873 which is M. gemmulata Semper, 1865 (:54, fig. 72); Mytilus edulis Linnaeus, 1758 which is M. galloprovincialis Lamarck, 1819, since the former is not an eastern mediterranean species. Hinnites distorta, to allocate within the genus Crassadoma according to WALLER (1993), has been reported only once by DEMETROPOU-LOS (1971), most probably it has been misidentified with C. multistriata. We remove this taxon from the proposed checklist of molluscs of Cyprus, since this species is mainly distributed in the Atlantic Ocean and very seldom reported from the Western Mediterranean basin. FISCHER (1994) has reported over 90 Indo-Pacific species in the Levantine basin, and 8 of them have also been recorded from the Cyprus coasts. Today, the number of the known Indo-Pacific species occurring in the Island of Cyprus increased up to 32 (Table 3). Summing up all the data coming from this investigation and those of the past, a total of 696 Molluscs species are presently known from Cyprus.

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The bathybenthic and pelagic molluscan fauna off the Levantine coast, eastern Mediterranean

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KEY WORDS: New records, bathyal, benthic, pelagic, Levantine Basin, eastern Mediterranean.

ABSTRACT

A total of 4580 molluscan specimens, identified to 43 species, have been collected during eleven cruises off the Mediterranean coast of Israel at depths between 734 and 1558 m. Much of the material examined consists of juvenile specimens and empty shells — only 23 species were represented by living specimens, 11 of these were represented solely by juvenile specimens. Five species constitute new records for the eastern Mediterranean, 8 are newly recorded from the Levantine sea. The deep waters of the eastern Mediterranean are effectively separated from the Atlantic Ocean by the shallow Gibraltar and Sicilian sills. Yet, the most common benthic molluscs in depths greater than 1000 m off the Israeli coast are of Atlanto-Mediterranean and Boreal distribution, either eurybathic species with upper bathymetric range well within the circalittoral, or having epipelagic larvae. Though the benthic molluscan fauna is considerably impoverished compared with the western Mediterranean, 12 of 13 of the Mediterranean pelagic thecosomate species were collected. The Quaternary anoxic events, combined with the barrier posed by the shallow Siculo-Tunisian sill, and the basin's oligotrophy provide a plausible explanation for the sparse and impoverished mollusc fauna in the Levantine bathyal.

RIASSUNTO

Durante undici crociere oceanografiche intraprese al largo delle coste d' Israele a profondità comprese tra 734 e 1558 metri, sono stati raccolti 4580 esemplari di molluschi ascrivibili a 43 specie differenti. Gran parte del materiale esaminato è costituito da esemplari giovanili e conchiglie vuote. Solo 23 specie sono state rinvenute viventi e, tra queste, 11 specie sono rappresentate esclusivamente da esemplari giovanili. Cinque specie sono state raccolte per la prima volta nel Mediterraneo Orientale: Akritogyra conspicua (Monterosato, 1880), Alvania electa (Monterosato, 1874), Turbonilla micans (Monterosato, 1875), Gleba cordata Forskål, 1776, Thyasira oblonga (Monterosato, 1878). Otto specie costituiscono nuovi ritrovamenti per il Mar di Levante: Carinaria lamarckii (Peron & Lesueur, 1810), Xenoskenea pellucida (Monterosato, 1874), Diacria trispinosa (Lesueur, 1821), Yoldiella philippiana (Nyst, 1845), Cyclopecten cf. hoskynsi (Forbes, 1844), Thyasira granulosa (Monterosato, 1874), Thyasira eumyaria (Sats. M., 1870), Allogrammora (Jeffreys, 1882).

Nonostante le acque profonde del Mediterraneo Orientale siano separate dall'Oceano Atlantico da ben due barriere geografiche, le soglie poco profonde di Gibilterra e del Canale di Sicilia, i molluschi bentonici più comuni rinvenuti a profondità maggiori di mille metri al largo delle coste israeliane hanno affinità Atlanto-Mediterranea e Boreale. La loro presenza è spiegabile grazie all'elevata euribatia delle specie, presenti anche nel piano Circalitorale, e grazie alla presenza di larve epipelagiche a sviluppo planctotrofico. Benché la malacofauna dell'area appaia notevolmente impoverita in confronto a quella del Mediterraneo Occidentale, sono state raccolte 12 delle 13 specie pelagiche Mediterranee note di Thecosomata. L'anossia avvenuta nel Quaternario, le condizioni oligotrofiche del bacino, e la presenza della poco profonda soglia Siculo-Tunisina che ha funzionato da barriera, forniscono una plausibile spiegazione alla bassa diversità specifica ed alla bassa densità della malacofauna batiale del Mar di Levante.

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INTRODUCTION

The study of the Levantine Sea bathyal molluscs commenced with the voyages of the "POLA" (1890-1893), the most extensive deepsea expedition to take place in the Levant Basin (STURANY, 1896). The Danish Oceanographical Expedition to the Mediterranean, aboard the "THOR", sampled nine sites along the western limits of the Levantine Sea, from Cyrenaica to Rhodes, in 1910. In 1987, the German research vessel "METEOR" [cruise 5] collected benthic samples at sites between Crete and Israel at depths ranging between 95 and 4396 m (JANSSEN, 1989). In 1993, the "METE-OR" [cruise 25] obtained deep-sea biota from the region between Crete, Cyprus and Egypt at depths ranging between 200 and 2900 m (HIEKE et al., 1994). The fauna of Eratosthenes Seamount, south of Cyprus, was briefly sampled by the "POSEIDON" [cruise 201/2] on February 1994 (GALIL & ZIBROWIUS, 1998). Thus, when discussing the Mediterranean deep sea gastropod fauna BOUCHET & TAVIANI (1992) declared that the "Eastern Mediterranean is the least adequately sampled region".

A series of eleven cruises conducted between 1994 and 1999 as part of pollution monitoring surveys by the Israel Oceanographic and Limnological Research (IOLR) off the northern coast of Israel,

at depths between 734 and 1558 m, added to our knowledge of the Levantine bathyal mollusc fauna.

STUDY AREA

The Levantine Sea at the easternmost Mediterranean, east of the line connecting Rhodes and Cyrenaica is isolated from the deep Atlantic and western Mediterranean waters by the shallow Gibraltar Straits and the Siculo-Tunisian sill.

The Levantine deep water mass is distinguished by salinity and temperature values that are higher than in the rest of the Mediterranean (T< 13.8°C, S < 38.74, below 700 m) (Hecht et al., 1988). The Levantine Sea is ultra-oligotrophic (Berman et al., 1984; Krom et al., 1991). Chlorophyll-a concentrations are as low as 0.4 μ g l-1 nearshore, and decrease offshore to 0.05 μ g l-1 (Berman et al., 1986; Yacobi et al., 1995).

The area investigated is located off the northern coast of Israel, at depths between 734 and 1558 m (Plate 1, table 1).

MATERIALS AND METHODS

The material was collected during some monitoring surveys of two deepwater waste-dumping sites: an acidic sludge disposal site off



Haifa, coal fly ash site off Hadera, and a control site off Atlit. The samples were collected using a 0.5 mm plankton net secured atop a Marinovich-type deep water trawl. The sample was preserved in 10% buffered formalin aboard ship. In the laboratory samples were washed and sieved through a 500 μm mesh, preserved in 70% alcohol, and stained in Rose Bengal. The sorted material was sent to Livorno for identification. The material is deposited in the collection of the first author and the National collection, Tel Aviv University. Taxonomy follows the Check List of European Marine Mollusca (http://www.mnhn.fr/mnhn/bimm/clemam).

For each species only records within the Levantine sea and adjacent areas (southern Aegean sea, eastern Ionian sea) are cited, or, if none known, the easternmost Mediterranean record. Only synonyms used in the cited records are given. The number of specimens in each sample is given in parenthesis following the station number.

Gastropoda Cuvier, 1797

Prosobranchia Milne Edwards, 1848

Vetigastropoda Salvini-Plawen & Haszprunar, 1987

Trochoidea Rafinesque, 1815

Skeneidae Clark, 1851

Akritogyra conspicua (Monterosato, 1880)

Distribution.- Mediterranean endemic. Central and Western Mediterranean (WARÉN, 1992).

Material examined.- 97HF1 (2 + 12 juvs.); living specimens.

Remarks.- First record in the Eastern Mediterranean.

Caenogastropoda Cox, 1959

Rissooidea Gray, 1847

Rissoidae Gray, 1847

Alvania electa (Monterosato, 1874)

Distribution.- Atlantic-Mediterranean. Sicily (CARUS, 1893).

Material examined.- 95700 (2); living specimens.

Remarks.- First record in the Eastern Mediterranean.

Benthonella tenella (Jeffereys, 1869)

Plate 2, Figure A.

Distribution.- Atlantic-Mediterranean. Crete (Janssen, 1989; Koutsoubas *et al.*, 1992); southwest of Cyprus, 34.08.27N, 31.52.52'E (Janssen, 1989); Israel (Janssen, 1989).

Material examined.- 94H15 (1); 95HF1 (11); 95HF2 (2); 95HF3 (2); 95HF8 (12); 95HF9 (1); 95H4 (2); 95H7 (1); 95H13 (4); 95H14 (1); 95H15 (1); 95H19 (1); 95H20 (1); 96A1 (1); 96A2 (4); 96A5 (1); 96A6 (1); 96HA (1); 96H4 (2); 96H6 (2); 96H14 (1); 96H20 (1); 97H7 (10); 97H14 (3); 97H19 (2); 97H21 (5); 97H52 (4); 97HF1 (3); 97HF2 (2); 97HF3 (3); 97HF4 (2); 97HF6 (3); 97HF7 (3); 97HF8 (1); 97HF9 (6); 98H4 (3); 98H15 (11); 98H19 (9); 98H20 (8); 98H21 (4); 98H23 (5); 98H24 (8); 98H25 (3); 98H26 (9); 98H27 (7); 99A1 (2); 99A3 (3); 99A4 (3); 99A6 (32); 99A7 (4); 99H7 (2); 99H15 (3); 99H19 (2); 99H20 (3); 99H21 (1); 99H23 (2); 99H25 (1); 99H27 (3); 99H28 (1). A total of 230 living specimens, about 80% juveniles.

Firoloidea Rafinesque, 1815 Atlantidae Rang, 1829 *Atlanta fusca* Souleyet, 1852

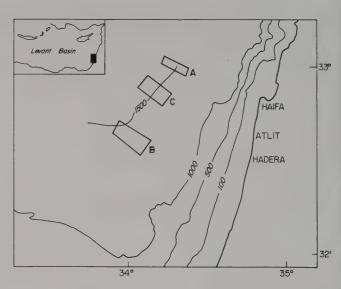


Plate 1. Map of the Mediterranean coast of Israel showing the locations of the Investigated areas (see also table 1). A, Haifa; B, Hadera; C, Atlit.

Tavola 1. Mappa delle coste Mediterranee di Israele con l'indicazione delle aree studiate (vedi anche tab.1). A, Haifa; B, Hadera; C, Atlit.

Distribution.- Cosmopolitan. South Aegean Sea (Kiortosis, 1969; Kotsoubas *et al.*, 1997); Levant sea (Vatova, 1974); Israel (Van Aartsen *et al.*, 1989).

Material examined.- 94H19 (1); 94A4 (1); 95A4 (1); 97H14 (1); 99A6 (13). A total of 17 adult shells, no living specimens.

Atlanta peronii Lesueur, 1817

Distribution.- Cosmopolitan. South Aegean Sea (Kotsoubas *et al.*, 1997); Crete, (Forbes, 1844; Jeffreys, 1883; Koutsoubas *et al.*, 1992); Cyclades (Forbes, 1844); Dodecanese (Forbes, 1844); Israel (Barash & Danin, 1992).

Material examined.- 94H6 (1); 96HA (11); 96H4 (3); 96H6 (1); 96H14 (1); 97H7 (1); 97H19 (1); 97H52 (3); 97HF1 (4); 97HF3 (3); 97HF7 (3); 97HF8 (3); 97HF9 (9); 98H15 (9); 98H19 (3); 98H20 (2); 98H22 (4); 98H23 (2); 98H25 (2); 98H28 (2); 99A1 (1); 99A3 (1); 99A5 (1); 99A6 (88); 99A7 (2); 99H4 (2); 99H19 (2); 99H21 (1); 99H23 (1); 99H27 (5). A total of 172 adult shells, no living specimens.

Oxygyrus keraudrenii (Lesueur, 1817)

Bellerophina minuta Forbes, 1844:186.

Distribution.- Cosmopolitan. South Aegean Sea (Forbes, 1844; Kotsoubas *et al.*, 1997); Levant sea (Vatova, 1974); Israel (Barash & Danin, 1992).

Material examined.- 94A1 (1); 94A4 (1); 94A7 (1); 95A1 (1); 95A4 (1); 95A7 (1); 97H21 (2). Juvenile shells only.

Carinariidae De Blainville, 1818

Carinaria lamarckii (Peron & Lesueur, 1810)

Distribution.- Cosmopolitan. Ionian sea (VATOVA, 1974).

Material examined.- 99A6 (1 juv.) shell.

Remarks.- First record in the Levant sea.

Firolidae Rafinesque, 1815

Firoloida desmarestia Lesueur, 1817



Plate 2, Figures B,C.

Cyclostrema minutum Jeffereys, 1883:395, pl. 16, fig. 1.

Cyclostrema solutum Di Geronimo, 1974:148.

Distribution.- Atlantic-Mediterranean. South Aegean Sea (Jeffreys, 1883; Kotsoubas *et al.*, 1992, 1997); Aegean (Moraitou-Apostolopoulou, 1985); Israel (Barash & Danin, 1992).

Material examined.- 99A6 (72) adult shells, no living specimens.

Conoidea Rafinesque, 1815

Conidae Rafinesque, 1815

Microdrillia loprestiana (Calcara, 1841)

Pleurotoma loprestianum Sturany, 1896:10.

Distribution.- Atlantic-Mediterranean. Crete (Jeffreys, 1883; Sturany, 1896; Koutsoubas *et al.*, 1992; 1997); Dodecanese (Sturany, 1896); Libya (Sturany, 1896); Rhodes (Zenetos & Van Aartsen, 1995); Israel (Haas, 1951; Barash & Danin, 1992).

Material examined.- 97H52 (1 juv.) living specimen.

Benthomangelia macra (Watson, 1881)

Plate 3, Figures A-C.

Pleurotoma (Mangelia) macra Sturany, 1896:11.

Distribution.- Atlantic-Mediterranean. Crete, (STURANY, 1896; JANSSEN, 1989; KOUTSOUBAS et al., 1992; 1997; 2000); Dodecanese (STURANY, 1896); Southwest of Cyprus, 34.08.27N, 31.52.52'E (JANSSEN, 1989); Israel (JANSSEN, 1989).

Material examined.- 94H5 (1); 94H15 (2); 94H16 (1); 94H19 (1); 94A1 (1); 94A5 (1); 94A7 (1); 95A5 (1); 95A7 (1); 95HF2 (2); 95HF3 (5); 95HF6 (2); 95HF8 (5); 95HF9 (1); 95H4 (5); 95H7 (4); 95H13 (3); 95H15 (1); 95H19 (4); 96A1 (1); 96A2 (1); 96A3 (1); 96HA (1); 96H6 (4); 96H20 (3); 97H7 (1); 97H21 (2); 97H52 (3); 97HF1 (2); 97HF6 (1); 98H20 (3); 98H21 (4); 98H23 (2); 98H24 (2); 98H25 (2); 98H26 (3); 98H27 (2); 98H28 (2); 99A4 (2); 99A6 (5); 99A7 (5); 99H7 (2); 99H20 (2); 99H21 (3); 99H23 (2); 99H25 (1); 99H27 (2); 99H28 (2); 99H29 (2). A total of 112 living specimens, about 90% juveniles.

Heterobranchia Gray, 1840

Heterostropha Fisher, P., 1885

Valvatoidea Gray, 1840

Hyalogyrinidae Warén & Bouchet, 1992

Xenoskenea pellucida (Monterosato, 1874)

Distribution. - Atlantic-Mediterranean. Central and Western Mediterranean (Warén *et al.*, 1993); South Aegean Sea (Tenekidis, 1989; Koutsoubas *et al.*, 1997).

Material examined.- 95HF2 (1, juv.); 95HF8 (1, juv.); shells.

Remarks.- First record in the Levantine sea.

Pyramidelloidea Gray, 1840

Pyramidellidae Gray, 1840

Chrysallida flexuosa (Monterosato, 1874)

Distribution.- Atlantic-Mediterranean. Crete (Karakassis, 1991;

Koutsoubas et al., 2000); Israel (Carrozza, 1984).

Material examined.- 95700 (1) living specimen.

Turbonilla micans (Monterosato, 1875)

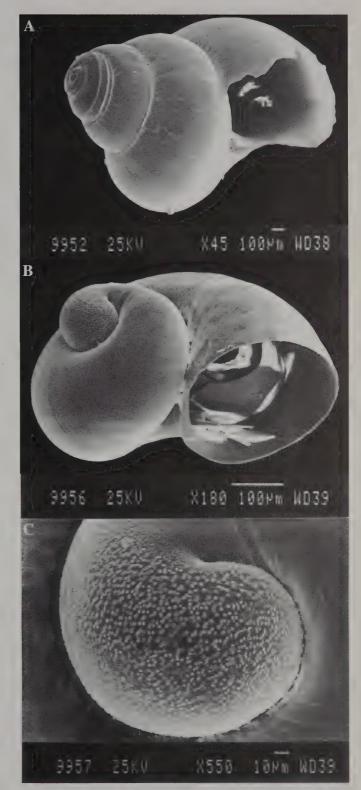


Plate 2. Benthonella tenella (Jeffreys, 1869), A. x 45; Firoloida desmarestia Lesueur, 1817, B. x 180; C. x 550 detail of protoconch.

Distribution.- Atlantic-Mediterranean.

Material examined.- 97HF9 (2); 99H23 (1); adult, living specimens.

Remarks.- First record in the Eastern Mediterranean.



Opistobranchia Milne Edwards, 1848

Cephalaspidea Fisher, P., 1883

Acteonidae D'Orbigny, 1842

Crenilabium exile (Jeffreys, 1870)

Distribution.- Atlantic-Mediterranean. Aegean Sea (Carus, 1893; Koutsoubas & Koukouras, 1993); Crete, (Janssen, 1989; Koutsoubas et al., 1992; 2000); Rhodes (Tenekidis, 1989; Zenetos & Van Aartsen, 1995); Israel (Janssen, 1989; s, 1992).

Material examined.- 98H23 (1); 98H27 (1); 99A6 (1); juvenile, living specimens.

Cylichnidae Adams, H. & A., 1854

Roxania utriculus (Brocchi, 1814)

Bulla utriculus Forbes, 1844:164.

Distribution.- Atlantic-Mediterranean. Aegean Sea (Koutsoubas & Koukouras, 1993); Crete, (Forbes, 1844; Koutsoubas et al., 1992); Cyclades (Forbes, 1844); Rhodes (Tenekidis, 1989; Zenetos & Van Aartsen, 1995); Israel (Haas, 1951; Barash & Danin, 1971, 1992).

Material examined.- 97HF9 (1 juv.) shell only.

Thecosomata De Blainville, 1824

Euthecosomata Meisenheimer, 1905

Cavoliniidae Gray, 1850

Cavolinia gibbosa gibbosa (D'Orbigny, 1835)

Hyalea gibbosa Forbes, 1844:132.

Distribution.- Endemic to the eastern Mediterranean. Aegean Sea (RAMPAL, 1970; KOUTSOUBAS & KOUKOURAS, 1993); Crete, (FORBES, 1844; JEFFREYS, 1883); Egypt (RAMPAL, 1970); Cyprus (RAMPAL, 1970); Israel (Almogi-Labin & Reiss, 1977; Barash & Danin, 1971, 1992).

Material examined.- 99A6 (2) adult specimens, shell only.

Remarks.- The morphological characters of our specimens correspond to the eastern Mediterranean subspecies, which is distinguished from *C. gibbosa flava* (rarely collected in the Mediterranean) in its more rounded ventral surface. Corselli and Grecchi (1987, 1990) considered *C. gibbosa gibbosa* a geographically-isolated entity of a recent (Holocene) penetration from the Atlantic Ocean.

Cavolinia inflexa (Lesueur, 1813)

Distribution.- Cosmopolitan. Aegean Sea (Forbes, 1844; Koutsoubas & Koukouras, 1993); Levantine basin (Pastouret, 1970; Rampal, 1975).

Material examined.- 99A6 (5) adult specimens, shells only.

Diacria trispinosa (Lesueur, 1821)

Distribution.- Cosmopolitan. Aegean Sea (PASTOURET, 1970; RAM-PAL, 1975; KOUTSOUBAS & KOUKOURAS, 1993).

Material examined.- 99A6 (1) single broken shell.

Remarks.- Though *D. trispinosa* is widely distributed in the Mediterranean, its presence in the sea was considered accidental by Corselli and Grecchi (1990). First record in the Levantine sea.

Clio pyramidata lanceolata (Lesueur, 1813)

Cleodora pyramidata Forbes, 1844:132.

Distribution.- Indo West Pacific Ocean, Eastern Atlantic, Mediter-

ranean. Crete (Forbes, 1844; Koutsoubas & Koukouras, 1993); Cyclades (Forbes, 1844); Asia Minor (Forbes, 1844); Cyprus (Tornaritis, 1987; Cecalupo & Quadri, 1996).

Material examined. 94A1 (2); 95A1 (1); 98H24 (2); 98H26 (1); 98H28 (2); 99A6(160). A total of 168 adult specimens, shells only. Many more fragments were collected, but not counted.

Remarks.- Most reports of this species refer to a subspecies known from the Mediterranean only from shells (Bedulli et al., 1995). The specimens collected off the Israeli coast present the morphological characters of the subsp. *C. pyramidata lanceolata* (Rampal, 1975).

Clio cuspidata (Bosc, 1802)

Cleodora cuspidata Forbes 1844:132.

Distribution.- Cosmopolitan. Aegean Sea (Koutsoubas & Koukouras, 1993); Crete (Forbes, 1844); Cyprus (Tornaritis, 1987); Israel (Almogi-Labin & Reiss, 1977; Barash & Danin, 1992).

Material examined. 99A6 (3) adult specimens, shells only.

Creseis acicula Rang, 1828

Distribution.- Cosmopolitan. Aegean (Moraitou-Apostolopoulou, 1985; Koutsoubas & Koukouras, 1993); Crete (Jeffreys, 1883; Koutsoubas et al., 1992); northern Levantine Sea (Berdugo & Kimor, 1967); Egypt (Vatova, 1974); Israel (Berdugo & Kimor, 1967; Almogi-Labin & Reiss, 1977; Barash & Danin, 1971, 1992). Material examined.- 99A6 (4) adult specimens, shells only.

Creseis virgula virgula Rang, 1828

Distribution.- Cosmopolitan. Aegean Sea (Koutsoubas & Koukouras, 1993); Crete (Koutsoubas *et al.*, 1992); Egypt (Vatova, 1974); Israel (Almogi-Labin & Reiss, 1977; Barash & Danin, 1971, 1992).

Material examined.- 94A4 (1); 95A4 (1); 99A6 (18), adult specimens, shells only.

Hyalocylis striata (Rang, 1828)

Criseis striata Forbes, 1844:132.

Distribution.- Atlantic-Mediterranean. Aegean Sea (RAMPAL, 1970; KOUTSOUBAS & KOUKOURAS, 1993); Crete (FORBES, 1844); Cyclades (FORBES, 1844); Asia Minor (FORBES, 1844); Cyprus (RAMPAL, 1970); Libya (RAMPAL, 1970); Egypt (RAMPAL, 1970); Israel (ALMOGI-LABIN & REISS, 1977; BARASH & DANIN, 1992).

Material examined.- 96H4 (1); 99A6 (3) juvenile specimens, shells only.

Styliola subula (Quoy & Gaimard, 1827)

Clio subula Jeffreys, 1883:401.

Distribution.- Cosmopolitan. Crete (Jeffreys, 1883); Cyprus (Cecalupo & Quadri, 1996); Israel (Almogi-Labin & Reiss, 1977; Barash & Danin, 1971, 1992).

Material examined.- 94H5 (3); 94H15 (3); 95A1 (1); 94A2 (3); 94A7 (3); 95A2 (3); 95A7 (3); 95HF1 (1); 95HF8 (1); 95HA (2); 95H15 (3); 95H19 (1); 96H4 (2); 96H14 (2); 97H14 (1); 97H21 (1); 97HF7 (1); 98H21 (1); 99A4 (1); 99A6 (277); 99A7 (1). A total of 314 adult specimens, shells only.

Limacinidae Gray, 1840



Limacina trochiformis (D'Orbigny, 1836)

Spirialis trochiformis Jeffreys, 1883:401.

Spiratella trochiformis Vatova, 1974:107.

Distribution.- Tropical and subtropical seas. Aegean Sea (Vatova, 1974; Koutsoubas & Koukouras, 1993); Crete (Jeffreys, 1883; Carus, 1893; Koutsoubas *et al.*, 1992); Rhodes (Berdugo & Kimor, 1967); Cyprus (Bogi *et al.*, 1989); Israel (Almogi-Labin & Reiss, 1977; Barash & Danin, 1992).

Material examined.- 94A1 (1); 94A4 (1); 95A1 (1); 95A4 (1); 95H4 (1); 95H13 (2); 95H19 (2); 95700 (1); 99A6(411). A total of 421 adult shells, no living specimens.

Limacina inflata (D'Orbigny, 1836)

Distribution.- Cosmopolitan. Aegean Sea (Koutsoubas & Koukouras, 1993); Crete (Jeffreys in Carus, 1893; Berdugo & Kimor, 1967; Koutsoubas *et al.*, 1992); northern Levantine Basin (Berdugo & Kimor, 1967); Cyprus (Bogi *et al.*, 1989); Israel (Berdugo & Kimor, 1967; Barash & Danin, 1971, 1992; Almogi-Labin & Reiss, 1977).

Material examined.- 94H16 (1); 94A2 (2); 94A4 (3); 94A7 (2); 95A2 (2); 95A4 (3); 95A7 (2); 95HF7 (1); 95HA (3); 95H7 (1); 95H13 (2); 95H15 (1); 95H19 (1); 96A2 (1); 96A3 (1); 96HA (5); 96H4 (10); 96H20 (1); 97HF1 (3); 97HF9 (7); 99A4 (1); 99A6 (445). A total of 498 adult shells, no living specimens.

Peraclidae Tesch, 1913

Peracle reticulata (D'Orbigny, 1836)

Distribution.- Cosmopolitan. Aegean Sea (Vatova, 1974; Koutsoubas & Koukouras, 1993); Israel (Almogi-Labin & Reiss, 1977; Barash & Danin, 1992).

Material examined.- 99A6 (8) adult specimens, shells only.

Cymbuliidae Gray,1840

Gleba cordata Forskål, 1776

Plate 4, Figure A.

Distribution.- Cosmopolitan. Between Sicily and Libya (RAMPAL, 1975).

Material examined.- 99A6 (31). Adult specimens, shells only. Remarks.- First record in the Eastern Mediterranean.

Bivalvia Linnaeus, 1758

Protobranchia Pelseneer, 1889

Nuculoida Dall, 1889

Yoldiidae Glibert & Van De Poel, 1965

Yoldia micrometrica (Seguenza, G., 1877)

Plate 4, Figure B.

Distribution.- Atlantic-Mediterranean. Crete (Janssen, 1989; Koutsoubas *et al.*, 1992); Eratosthenes seamount (Galil & Zibrowius, 1998); Israel (Janssen, 1989).

Material examined.- 94H3 (1); 94H4 (4); 94H5 (1); 94H6 (1); 94H7 (1); 94H9 (1); 94H19 (1); 94A1 (1); 94A4 (2); 94A5 (9); 94A1 (1); 95A1 (1); 95A4 (2); 95HF1 (20); 95HF2 (7); 95HF3 (16); 95HF4 (4); 95HF6 (3); 95HF7 (12); 95HF8 (37); 95HF9 (7); 95HA (19); 95H4 (27); 95H7 (4); 95H13 (8); 95H14 (7); 95H15 (11); 95H19 (6); 95H20 (6); 96A1 (18); 96A2 (6); 96A3 (2); 96A4 (7); 96A5 (4); 96A6 (5); 96HA (15); 96H4 (10); 96H6

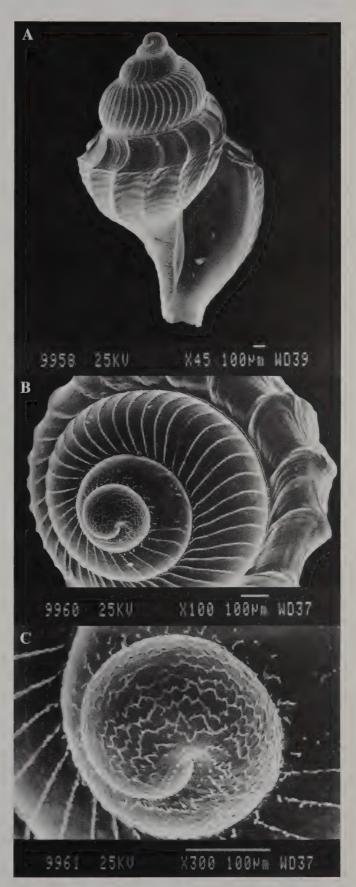


Plate 3. Benthomangelia macra (Watson, 1881). A. x 45; B. x 100 detail of protoconch; C. x 300 detail of protoconch.



(12); 96H14 (6); 96H20 (20); 97H7 (7); 97H14 (21); 97H19 (5); 97H21 (29); 97H51 (14); 97H52 (25); 97HF1 (19); 97HF3 (15); 97HF4 (2); 97HF6 (3); 97HF7 (7); 97HF8 (16); 97HF9 (37); 98H4 (19); 98H7 (2); 98H19 (8); 98H20 (22); 98H21 (18); 98H22 (6); 98H23 (13); 98H24 (8); 98H25 (21); 98H26 (9); 98H27 (26); 98H28 (9); 99A1 (4); 99A3 (3); 99A4 (7); 99A6 (2); 99A716 (2); 99H4 (24); 99H7 (3); 99H15 (11); 99H19 (19); 99H20 (9); 99H21 (21); 99H23 (11); 99H24 (4); 99H25 (5); 99H26 (3); 99H27 (12); 99H28 (1); 99H29 (1). A total of 827 living specimens, 80% juveniles.

Remarks.- *Y. micrometrica* is the most common and abundant species in our samples. As far as we can determine it is the first record of *Y. micrometrica* being collected in great numbers in the Mediterranean bathyal, conceivably, the species took advantage of the lack of competitors to occupy an empty ecological niche. Most of the specimens are juveniles.

Yoldiella philippiana (Nyst, 1845)

Arca (Bathyarca) koreni sensu (Kobelt, 1891) non (Danielseen, 1859).

Distribution.- Atlantic-Mediterranean. Lybia (Sturany, 1896); Crete (Sturany, 1896); Cyclades (Sturany, 1896).

Material examined.- 94H16 (1); 95HF7 (2); 96A1 (1); 96H4 (1); 96H6 (1); 96H14 (1); 97HF7 (1); living, juvenile specimens.

Remarks.- First record in the Levantine sea.

Pteromorphia Beurlen, 1944

Arcoida Stoliczka, 1871

Arcidae Lamark, 1809

Bathyarca pectunculoides (Sacchi, 1834)

Arca pectunculoides Jeffreys, 1883:394.

Arca (Bathyarca) pectunculoides Sturany, 1896:20.

Distribution.- Boreal. Aegean Sea (Jeffreys in Carus, 1893); Crete (Jeffreys, 1883; Sturany, 1896; Janssen, 1989; Koutsoubas *et al.*, 2000); Libya (Sturany, 1896); Dodecanese (Sturany, 1896); Rhodes (Zenetos & Van Aartsen, 1995); Israel (Janssen, 1989; Barash & Danin, 1992).

Material examined.- 94H15 (1); 95HF1 (2); 95HF8 (2); 95HF9 (1); 95HA (2); 95H4 (1); 95H7 (2); 95H13 (2); 95H14 (1); 95H15 (1); 96A4 (1); 96H4 (2); 96H14 (1); 97H7 (5); 97H14 (2); 97H19 (3); 97H21 (2); 97H51 (2); 97H52 (4); 97HF1 (2); 97HF3 (3); 97HF7 (1); 98H20 (2); 98H21 (1); 98H22 (4); 99A4 (1); 99A6 (1); 99H7 (2); 99H15 (4); 99H19 (1); 99H21 (2); 99H23 (2); 99H27 (1). A total of 64 living specimens, 50% juveniles.

Bathyarca philippiana (Nyst, 1848)

Distribution.- Atlantic-Mediterranean. Crete (Janssen, 1989; Koutsoubas *et al.*, 1992); Eratosthenes seamount (Galil & Zibrowius, 1998).

Material examined.- 94A5 (5 juvs.), living specimens.

Pterioida Newell, 1965 Limidae Rafinesque, 1815 *Limatula subauriculata* (Montagu, 1808) *Lima elongata* Forbes, 1844:192.

Lima (Limatula) subauriculata Sturany, 1896:21.

Distribution.- Boreal. Aegean Sea (Jeffreys, 1879); Crete (Forbes, 1844; Sturany, 1896; Koutsoubas *et al.*, 1992; 2000); Libya (Sturany, 1896); Cyclades (Forbes, 1844); Dodecanese (Forbes, 1844); Israel (Barash & Danin, 1992).

Material examined.- 97HF9-(53).

Pectinidae Rafinesque, 1815

Cyclopecten cf. boskynsi (Forbes, 1844)

Pecten hoskynsi Forbes, 1844:192.

Pecten (Amussium) hoskynsi Sturany, 1896:20.

Distribution.- Atlantic-Mediterranean. Aegean Sea (Jeffreys, 1879); Crete (Koutsoubas *et al.*, 2000); Asia minor (Forbes, 1844); Cyclades (Sturany, 1896); Dodecanese (Sturany, 1896); Libya (Sturany, 1896).

Material examined.- 99H27 (1), valve of adult specimen.

Remarks.- First record in the Levantine sea.

Heterodonta Neumayr, 1884

Veneroida Adams, H. & A., 1857

Thyasiridae Dall, 1900

Thyasira granulosa (Monterosato, 1874)

Distribution.- Atlantic-Mediterranean. Crete (Koutsoubas et al., 1992); Rhodes (Zenetos & Van Aartsen, 1995).

Material examined.- 97HF9 (1 juv.) living specimen.

Remarks.- First record in the Levantine sea.

Thyasira oblonga (Monterosato, 1878)

Plate 4, Figure C.

Distribution.- Atlantic-Mediterranean.

Material examined.- 95HF2 (4); 95HF7 (2) living adult specimens.

Remarks.- This species was often erroneously described and depicted as *Thyasira* (or *Leptaxinus*) *subovata* (Jeffreys, 1881), a species we consider distinct from *Thyasira oblonga* (GIRIBET & PEÑAS, 1997; CECALUPO & GIUSTI, 1989). First record in the Eastern Mediterranean.

Thyasira eumyaria (Sars, M., 1870)

Distribution.- Ionian Sea (Parenzan, 1976; DI Geronimo, 1974).

Material examined.- 95HA (1); 96A1 (1); 96H4 (2); 98H28 (1); 99H27 (1) living specimens, 2 adults, 4 juveniles.

Remarks.- First record in the Levantine Sea.

Semelidae Stoliczka, 1870

Abra longicallus (Scacchi, 1834)

Scrobicularia longicallis Jeffreys, 1883:395.

Syndosmya longicallis Sturany, 1896:15.

Distribution.- Atlantic-Mediterranean. Crete (Jeffreys, 1883; Jeffreys in Carus, 1893; Sturany, 1896; Koutsoubas et al., 1992; 2000). Libya (Sturany, 1896); Cyclades (Sturany, 1896); Dodecanese (Sturany, 1896); Rhodes (Jeffreys, 1881a; Zenetos & Van Aartsen, 1995); Cyprus (Hadjichristophorou et al., 1997); Israel (Janssen, 1989).

Material examined.- 97HF9 (1 juv.) living specimen.



Kelliellidae Fischer, P., 1887

Kelliella abyssicola (Forbes, 1844)

Kellia abyssicola Forbes, 1844:192.

Kelliella miliaris Sturany, 1896:17.

Distribution.- Boreal. Crete (Forbes, 1844; Sturany, 1896; Janssen, 1989; Koutsoubas *et al.*, 1992; 2000); Libya (Sturany, 1896); Cyclades (Forbes, 1844); Dodecanese (Sturany, 1896); Asia Minor (Forbes, 1844); Rhodes (Zenetos & Van Aartsen, 1995); Eratosthenes seamount (Galil & Zibrowius, 1998); Israel, (Janssen, 1989).

Material examined.- 94H4 (3); 94H5 (7); 94H15 (5); 94H16 (2); 94H19 (5); 94A1 (1); 94A2 (1); 94A5 (12); 94A6 (1); 95A1 (1); 95A2 (1); 95A6 (1); 95HF1 (10); 95HF2 (4); 95HF3 (6); 95HF4 (2); 95HF6 (3); 95HF7 (3); 95HF8 (11); 95HF9 (2); 95HA (8); 95H4 (10); 95H7 (5); 95H13 (7); 95H14 (4); 95H15 (7); 95H19 (11); 95H20 (1); 95700 (66); 96A1 (7); 96A2 (1); 96A3 (2); 96A4 (1); 96A5 (1); 96A6 (1); 96HA (13); 96H4 (14); 96H6 (8); 96H14 (3); 96H20 (8); 97H7 (9); 97H13 (1); 97H14 (6); 97H21 (7); 97HF1 (22); 97HF2 (3); 97HF3 (9); 97HF4 (3); 97HF6 (2); 97HF7 (3); 97HF8 (16); 98H4 (5); 98H7 (3); 98H15 (7); 98H19 (3); 98H20 (6); 98H21 (6); 98H22 (4); 98H23 (7); 98H24 (6); 98H25 (8); 98H26 (7); 98H27 (6); 98H28 (4); 99A3 (1); 99A6 (112); 99A7 (1); 99H4 (3); 99H7 (2); 99H15 (2); 99H19 (5); 99H20 (6); 99H21 (8); 99H23 (2); 99H26 (3); 99H27 (3); 99H29 (2). A total of 561 living specimens, 50% juveniles.

Anomalodesmata Dall, 1889

Pholadomyoida Newell, 1965

Lyonsiidae Fischer, P., 1887

Allogramma formosa (Jeffreys, 1882)

Lyonsia aegeensis Sturany 1896:15.

Lyonsia formosa aegeensis Parezan, 1976:390.

Distribution.- Atlantic-Mediterranean. Sicily (Jeffreys, 1881b).

Material examined.- 95HA (1 juv.); 97H52 (1 juv.) broken, living specimens.

Remarks.- First record in the Levantine Sea.

Cuspidariidae Dall, 1886

Cuspidaria rostrata (Spengler, 1793)

Neaera rostrata Sturany, 1896:16.

Distribution.- Atlantic-Mediterranean. Aegean Sea (Jeffrey in Carus, 1893); Crete (Janssen, 1989; Koutsoubas *et al.*, 1992; 2000); Libya (Sturany, 1896); Cyclades (Sturany, 1896); Rhodes (Zenetos & Van Aartsen, 1995); Cyprus (Demetropoulos, 1971; Hadjichristophorou *et al.*, 1997); Israel (Haas, 1951; Barash & Danin, 1982, 1992).

Material examined.- 94A7 (1); 95A7 (1); 96A1 (2); 99H20 (1); juvenile, living specimens.

Cardiomyia costellata (Deshayes, 1835)

Neaera costellata Sturany, 1896:16.

Distribution.- Atlantic-Mediterranean. Crete (Forbes, 1844; Jeffreys, 1883; Jeffreys in Carus, 1893; Janssen, 1989; Koutsoubas *et al.*, 1992; 2000); Cyclades (Forbes, 1844); Asia Minor (Forbes, 1844); Libya (Sturany, 1896); Rhodes (Zenetos & Van Aartsen); Cyprus (Demetropoulous, 1971); Eratosthenes

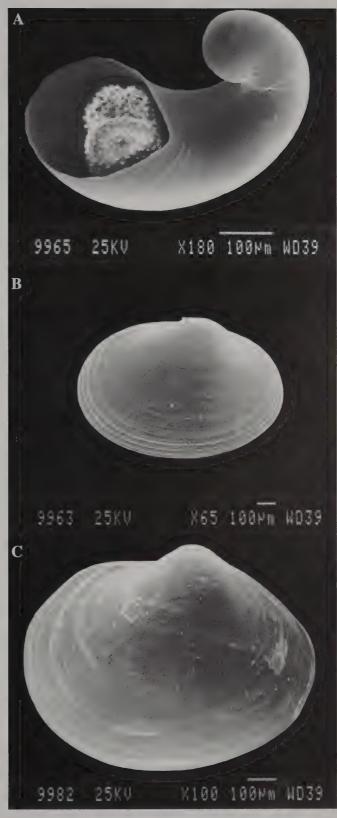


Plate 4. Gleba cordata Forskål, 1776. A. x 180; Yoldia micrometrica (Seguenza, G., 1877). B. x 65; Thyasira oblonga (Monterosato, 1878), C. x 100.

seamount (Galil & Zibrowius, 1998); Israel (Haas, 1951; Barash & Danin, 1982, 1992; Janssen, 1989).

Material examined.- 94H5 (7); 94H6 (3); 94H15 (6); 94H16 (15);



94H19 (3); 94A1 (3); 94A4 (6); 94A5 (3); 94A6 (5); 95A1 (3); 95A4 (6); 95A6 (6); 95HF1 (3); 95HF2 (3); 95HF3 (1); 95HF4 (2); 95HF6 (1); 95HF7 (1); 95HF8 (9); 95HF9 (3); 95HA (4); 95H4 (6); 95H13 (3); 95H15 (4); 95H19 (10); 95H20 (5); 96A1 (3); 96A2 (3); 96A3 (2); 96A4 (4); 96A5 (5); 96A6 (3); 96HA (1); 96H4 (9); 96H6 (12); 96H14 (5); 96H20 (7); 97H7 (10); 97H13 (3); 97H14 (1); 97H19 (3); 97H21 (4); 97H51 (3); 97H52 (6); 97HF2 (2); 97HF6 (1); 97HF7 (2); 97HF8 (3); 97HF9 (2); 98H4 (3); 98H15 (35); 98H19 (7); 98H20 (1); 98H22 (2); 98H23 (5); 98H24 (2); 98H26 (3); 98H27 (6); 98H28 (3); 99A1 (6); 99A3 (1); 99A4 (12); 99A5 (7); 99A6 (2); 99A7 (6); 99H4 (8); 99H7 (19); 99H15 (19); 99H19 (17); 99H20 (22); 99H21 (2); 99H23 (12); 99H24 (6); 99H25 (6); 99H27 (8); 99H29 (2). A total of 437 living specimens, all juveniles.

Scaphopoda Bronn, 1862 Gadilida Starbogatov, 1982 Entalinidae Chistikov, 1979 *Entalina tetragona* (Brocchi, 1814)

Dentalium quinquaulae Forbes, 1844:188.

Siphonodentalium quinquangulare Sturany, 1896:34.

Distribution.- Atlantic-Mediterranean. Aegean Sea (VAMVAKAS, 1971); Crete, (Forbes, 1844; Jeffreys, 1883; Sturany, 1896; Janssen, 1989); Cyclades (Forbes, 1844); Asia Minor (Forbes, 1844); Israel (Haas, 1951; Janssen, 1989; Barash & Danin, 1992).

Material examined.— 94H5 (2); 94H19 (7); 95HF2 (6); 95HF8 (9); 95HA (11); 95H4 (5); 95H7 (1); 95H13 (3); 95H14 (2); 95H15 (8); 95H19 (9); 95H20 (3); 96A1 (15); 96A4 (1); 96A5 (4); 96A6 (1); 96HA (48); 96H4 (13); 96H6 (13); 96H14 (4); 96H20 (14); 97H7 (3); 97H13 (1); 97H14 (20); 97H19 (3); 97H21 (31); 97H51 (2); 97H52 (28); 97HF1 (18); 97HF2 (4); 97HF3 (7); 97HF4 (1); 97HF7 (3); 97HF8 (8); 97HF9 (32); 98H4 (4); 98H7 (1); 98H15 (4); 98H19 (5); 98H20 (11); 98H21 (4); 98H22 (7); 98H23 (3); 98H24 (4); 98H25 (19); 98H26 (5); 98H27 (14); 99A1 (2); 99A4 (4); 99A5 (1); 99A6 (11); 99A7 (2); 99H4 (6); 99H7 (1); 99H15 (1); 99H19 (8); 99H20 (11); 99H21 (3); 99H23 (3); 99H25 (3); 99H27 (6); 99H28 (1); 99H29 (1). A total of 485 living specimens, 60% juveniles.

DISCUSSION

Much of the material examined consists of juvenile specimens and empty shells — only 23 of the 42 species collected were represented by living specimens, 11 of these were represented solely by juvenile specimens. Though an established method of collecting suprabenthic crustaceans, mainly percarids, from the deep sea (Cartes & Sorbe, 1993, 1997), the use of a plankton net attached to a deep-water trawl may affect the components and size of the biota sampled, so that full-sized live molluscan specimens may be undercollected. However, Di Geronimo (1974) found that in a sample collected by dredge at 1500 m "non sono stati raccolti esemplari viventi", and that other samples "è constituta da individui giovani o addirittura solo dalla protoconca" (Di Geronimo, 1974). Janssen (1989), whose material was collected both by beam-trawl and box-core, stated that "all true deepwater species are represented by shells only".

The deep waters of the eastern Mediterranean are separated from the Atlantic Ocean by the Gibraltar and Sicilian straits, yet the most common benthic molluscs in depths greater than 1000 m off the Israeli coast are the Atlanto-Mediterranean and Boreal Yoldia micrometrica, Kelliella abyssicola, Cardyomia costellata, Entalina tetragona, Benthomangelia macra, Benthonella tenella, and Bathyarca pectunculoides present respectively in 83, 77, 76, 63, 59, 50 and 33 out of the 90 samples collected during our study. The same species were identified by Janssen (1989) in material sampled by boxcore at a station off the Israeli coast at 1217 m. K. abyssicola, C. costellata, E. tetragona and B. pectunculoides are eurybathic species with upper bathymetric range well within the circalittoral (>150 m), whereas both the more stenobathic B. macra and B. tenella have epipelagic larvae (BOUCHET & Warén, 1979), enabling them to overcome the barrier posed by the shallow Gibraltar and Sicilian sills.

BOUCHET & TAVIANI (1992) have suggested that much of the Mediterranean deep-sea fauna is made-up of non-reproducing pseudopopulations, whereas DI GERONIMO et al. (2001), who studied the bathyal thanatocoenoses molluscs in the Southern Tyrrhenian Sea, propose that the deep Mediterranean benthos is composed of biogeographically autochthonous species. Since the populations of the most common benthic molluscs in depths greater than 1000 m off the Israeli coast are composed of both adult and juvenile specimens, and gravid benthic decapod crustaceans and fish were collected from the depths of the Levantine sea, our results support the "autochthonous" model (Galil & Goren, 1994; Goren & Galil, 1997; Fishelson & Galil, 2001).

"The floro-faunistic impoverishment of the eastern Mediterranean compared with the western Mediterranean richness in species is well documented" (SARA, 1985), but on occasion inaccurate. Thus, SARA's (1985) assertion "Not one of the eight species of Pteropoda of the western Mediterranean reaches the eastern zone" is erroneous – already Forbes (1844) described nine the costomate species from the Aegean sea. In fact, 12 of the 13 the cosomate species known from the Mediterranean (Corsell & Grecchi, 1990) were collected in the present study off the coast of Israel. SARA's error notwithstanding, the progressive faunistic decrease from west to east in the Mediterranean Sea has been more pronounced for the deep benthos than for the whole fauna (FREDJ & LAUBIER, 1985). Indeed, the low-diversity, low-density Levantine deep water fauna is considered the poorest in the Mediterranean. Comparing extant benthic deep-water molluscs of the western with those of the eastern Mediterranean we find that species that are common in the western basin had not been found in the eastern.

DI GERONIMO et al. (2001) believe the impoverishment of the deep fauna stems from the onset of the warm homothermy that led to the "disappearance of most cold-stenothermic species..., and a drop in diversity and richness in general".

The deep homothermy, extreme oligotrophy, and high salinity prevent settlement by members of the stenothermic and stenohalinic Atlantic bathyal, as far as they were able to cross the Gibraltar and the Sicilian sills (Pérès, 1985). In addition, during the past 450 kyr eleven layers of organic-rich sediments (sapropels) have been deposited in the eastern Mediterranean, the most recent layer deposited between 9000 and 6000 years BP (Troelstra et al., 1991; Jorissen et al., 1993; Cheddadi & Rossignol-Strick,



Table 1. Data of sampling stations A- Atlit; H- Hadera; HF-Haifa

Station	coordinates	depth (m)	date	Station	coordinates	depth (m)	date
94H3	32°31'N 34°14'E	1336	December 1994	97HF3	32°57'N 34°34'E	1349	September 1997
94H4	32°28'N 34°16'E	1290	December 1994	97HF4	32°59'N 34°33'E	1389	September 1997
94H5	32°32'N 34°07'E	1374	December 1994	97HF6	32°00'N 34°30'E	1434	September 1997
94H6	32°36'N 34°02'E	1456	December 1994	97HF7	32°57'N 34°40'E	1227	September 1997
94H7	32°41'N 34°04'E	1482	December 1994	97HF8	32°86'N 34°36'E	1313	September 1997
94H15	32°42'N 34°11'E	1439	December 1994	97HF9	32°1'N 34°28'E	1439	September 1997
94H16	32°39'N 34°16'E	1403	December 1994	97H7	32°41'N 34°3'E	1493	September 1997
94H19	32°31'N 34°06'E	1390	December 1994	97H1	32°41'N 34°10'E	1467	September 1997
95A1	32°47'N 34°20'E	1437	January 1995	97H14	32°40'N 34°13'E	1415	September 1997
95A2	32°51'N 34°20'E	1454	January 1995	97H19	32°33'N 34°4'E	1421	September 1997
95A3	32°52'N 34°18'E	1487	January 1995	97H21	32°29'N 34°17'E	1272	September 1997
95A4	32°53'N 34°17'E	1503	January 1995	97H51	32°27'N 34°4'E	1362	September 1997
95A5	32°52'N 34°14'E	1533	January 1995	97H52	32°30'N 34°19'E	1269	September 1997
95A6	32°53'N 34°10'E	1558	January 1995	98H4	32°29'N 34°15'E	1296	November 1998
95A7	32°45'N 34°36'E	1012	January 1995	98H7	32°40'N 34°2'E	1500	November 1998
95HF1	32°59'N 34°34'E	1362	September 1995	98H15	32°41'N 34°13'E	1243	November 1998
95HF2	32°59'N 34°35'E	1337	November 1995	98H19	32°33'N 34°3'E	1462	November 1998
95HF3	32°57'N 34°37'E	1311	September 1995	98H20	32°31'N 34°3'E	1344	November 1998
95HF4	33°00'N 34°33'E	1384	September 1995	98H21	32°27'N 34°15'E	1276	November 1998
95HF5	33°00'N 34°28'E	1456	November 1995	98H22	32°30'N 34°2'E	1450	November 1998
95HF6	32°59'N 34°30'E	1433	September 1995	98H23	32°28'N 34°2'E	1389	November 1998
95HF7	33°00'N 34°41'E	1175	September 1995	98H24	32°29'N 34°3'E	1391	November 1998
95HF8	33°00'N 34°35'E	1345	November 1995	98H25	32°25'N 34°6'E	1351	November 1998
95HF9	33°01'N 34°28'E	1471	November 1995	98H26	32°27'N 34°16'E	1277	November 1998
95HA	32°30'N 34°11'E	1350	November 1995	98H27	32°29'N 34°19'E	1251	November 1998
95H4	32°29'N 34°15'E	1300	November 1995	98H28	32°25'N 34°5'E	1319	November 1998
95H7	32°40'N 34°11'E	1480	November 1995	99A1	32°47'N 34°21'E	1413	November 1999
95H13	32°41'N 34°11'E	1485	November 1995	99A3	32°51'N 34°17'E	1484	November 1999
95H14	32°38'N 34°15'E	1400	November 1995	99A4	32°51'N 34°16'E	1486	November 1999
95H15	32°40'N 34°14'E	1450	November 1995	99A5	32°52'N 34°15'E	1509	November 1999
95H19	32°31'N 34°7'E	1400	November 1995	99A6	32°53'N 34°13'E	1522	November 1999
95H20	32°32'N 34°18'E	1279	November 1995	99A7	32°53'N 34°10'E	1557	November 1999
95700	32°41'N 34.40'E	734	November 1995	99H4	32°31'N 34°17'E	1296	November 1999
96A1	32°46'N 34°20'E	1427	September 1996	99H7	32°41'N 34°3'E	1485	November 1999
96A2	32°54'N 34°22'E	1453	September 1996	99H15	32°41'N 34°13'E	.1448	November 1999
96A3	32°50'N 34°17'E	1471	September 1996	99H19	32°33'N 34°4'E	1421	November 1999
96A4	32°52'N 34°17'E	1501	September 1996	99H20	32°31'N 34°5'E	1394	November 1999
96A5	32°52'N 34°14'E	1518	September 1996	99H21	32°29'N 34°16'E	1284	November 1999
96A6	32°54'N 34°15'E	1528	September 1996	99H23	32°31'N 34°58'E	1412	November 1999
96HA	32°31'N 34°12'E	1365	October 1996	99H24	32°28'N 34°4'E	1398	November 1999
96H4	32°30'N 34°17'E	1292	October 1996	99H25	32°26'N 34°4'E	1344	November 1999
96H6	32°39'N 34°02'E	1505	October 1996	99H26	32°30'N 34°18'E	1272	November 1999
96H14	32°40'N 34°11'E	1397	October 1996	99H27	32°27'N 34°18'E	1260	November 1999
96H20	32°30'N 34°06'E	1374	October 1996	99H28	32°25'N 34°5'E	1330	November 1999
97HF1	32°58'N 34°37'E	1299	September 1997	99H29	32°25'N 34°2'E	1353	November 1999
97HF2	32°59'N 34°33'E	1374	September 1997				



1995), with upper depth limits quoted at 700 m (Thunell et al., 1984), 300 m (RHOLING & GIESKES, 1989), and 150 m in the north Aegean Sea (Perissoratis & Piper, 1992). Sapropels contain microfossils of planktonic origin but benthic fossils are mostly absent (Castradori, 1993; Rholing, 1994), indicative of anoxic bottom waters. The recurring stagnant (dysoxic and anoxic) episodes resulted in a reduction, or even demise of deep bottom-living fauna unable to avoid extinction by adapting to shallower depth: "Several species of deep-water ostracodes that are still common in the Western Mediterranean became extinct in the Eastern Mediterranean basin at the onset of early Holocene S1 sapropel deposition" (VAN HARTEN, 1987). The epipelagic fauna was unhurt: 9 of the 12 thecosomate species collected in this work were recorded from cores dating to the upper and middle Pleistocene taken in the Ionian sea (GRECCHI, 1984). George & Menzies (1968) and Menzies (1973) believe that the bathyal bottoms of the Levant are still inhospitable, or even azoic, after the last sapropelic event. The Quaternary anoxic events, combined with the barrier posed by the shallow Siculo-Tunisian sill, and the basin's oligotrophy and homothermy provide a plausible explanation for the sparse and impoverished mollusc fauna in the Levantine bathyal.

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A new species of *Ersilia* (Caenogastropoda, Eulimidae) from the Plio-Pleistocene of the central Mediterranean area

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KEY WORDS: Mollusca, Gastropoda, Eulimidae, Taxonomy, Pliocene, Pleistocene, Italy, Greece.

ABSTRACT

A new species of the genus Ersilia Monterosato, 1872 (Caenogastropoda, Eulimidae), E. aliceae n. sp., is described from the Pliocene (Lazio, Central Italy) and Pleistocene (W Sicily, Italy and NW Peloponnesus, Greece) of the central Mediterranean area. This is the first finding of the genus Ersilia for the Plio-Pleistocene. It is compared with the living E. mediterranea (Monterosato, 1869) and E. stancyki Warén, 1980, and with the fossil E. oligocaenica Lozouet, 1999. E. aliceae n. sp. closely resembles E. mediterranea, from which it mainly differs by having less numerous spiral lines and rounded teeth on the outer lip. Furthermore, E. aliceae n. sp. has a multispiral protoconch suggesting a planktotrophic larval development, while E. mediterranea has a paucispiral protoconch suggesting a non-planktotrophic development.

RIASSUNTO

Viene descritta una nuova specie fossile del genere Ersilia Monterosato, 1872 (Caenogastropoda, Eulimidae), E. aliceae n. sp., ritrovata nel Pliocene dell'Italia centrale (Magliano Sabina, Roma) e nel Pleistocene siciliano (Monte Serro, Sicilia occidentale) e greco (Killini, Peloponneso nord-occidentale). Si tratta della seconda specie fossile nota per il genere Ersilia, conosciuto dall'Oligocene superiore con la specie E. oligocaenica Lozouet, 1999, e della prima segnalazione nel Plio-Pleistocene. La specie è morfologicamente affine all'unico rappresentante vivente segnalato in Mediterraneo, E. mediterranea (Monterosato, 1869), dal quale differisce per avere linee spirali più incise e meno numerose (5-7 contro le 10-15 della specie vivente) e per la presenza di piccoli denti tubercoliformi sul labbro esterno. Inoltre, le protoconche delle due specie, multispiralata nella specie fossile (2.2-2.5 giri, altezza 260-315 µm, larghezza 220-250 µm, diametro del nucleo 40-50 µm) e paucispiralata in quella vivente (1.3-1.5 giri, altezza 210-220 µm, larghezza 250-280 µm, diametro del nucleo 70-75 µm), indicherebbero uno sviluppo larvale differente: planctotrofico in E. aliceae n. sp. e non-planctotrofico in E. nediterranea. E. aliceae n. sp. viene confrontata anche con la specie atlantica E. stancyki Warén, 1980 e con quella fossile E. oligocaenica.

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INTRODUCTION

Eulimid gastropods are a common component in Tertiary and Quaternary mollusc associations. Nevertheless, the only fossil record (LOZOUET, 1999) for the eulimid genus *Ersilia* is *Ersilia* oligocaenica Lozouet, 1999, from the French Upper Oligocene. This genus, described by MONTEROSATO (1872: 33) as a Lacunidae, was definitively attributed to Eulimidae by WARÉN (1980). Two living species belonging to *Ersilia* are known: *Ersilia mediterranea*, described by MONTEROSATO (1869: 15, fig. 8) as *Lacuna mediterranea* from the Mediterranean Sea, and *E. stancyki* Warén, 1980 from the Western Atlantic.

A study on the malacofaunas from the Lower Pleistocene of Monte Serro (Dattilo, Western Sicily) allowed the finding of a specimen belonging to the genus *Ersilia*. This specimen was erroneously identified as *E*. cfr. *mediterranea* (Monterosato) by Garilli (1998, see this reference also for the associated molluscan fauna). Later, two specimens of *Ersilia* sp. from the Pleistocene of Killini (North Western Peloponnesus, Greece) and from the Pliocene of Magliano Sabina (Roma, Central Italy) (see Tuccimei, 1880, for age and associated molluscan fauna) were sent to me by Stefano Palazzi. These specimens showed some original morphological features which are not present in other congeneric or similar recent and fossil eulimids, and thus a new species is described on this material in the present work.

SYSTEMATICS

Classis Gastropoda Cuvier, 1797

Ordo Neotaenioglossa Haller, 1882 Familia Eulimidae H. & A. Adams, 1853 Genus *Ersilia* Monterosato, 1872 *Ersilia aliceae* n.sp. Pl. 1, Figs 1-5, 8-10.

TYPE MATERIAL

Holotype (H 1.5 mm, D 0.8 mm, Pl. 1, Figs 1, 2, 7, 8), from the Lower Pleistocene of Monte Serro (Dattilo, Trapani, Western Sicily), Museo Geologico G.G. Gemmellaro, Palermo, Italy (DATP 001/2971).

Paratype 1 (H 1.2 mm, D 0.7 mm, Figs 3, 4, 10), from the Pleistocene of Killini (Elea, North-Western Peloponnesus, Greece), Goulandris Natural History Museum, Kifissia, Greece (17/5).

Paratype 2 (H 2.0 mm, D 1.0 mm, Figs 5, 9), from the Pliocene of Magliano Sabina (Roma, Central Italy), Museo Zoologico, Bologna, Italy (MZB 40001).

DERIVATIO NOMINIS

The species is named after my daughter Alice.

DESCRIPTION

Small, rather sturdy, ovate, moderately inflated shell reaching 2 mm in height (paratype 2). Protoconch conical, multispiral consisting of 2.2-2.5 whorls (according to VERDUIN's, 1977 method), sculptured by curved, opisthocyrt axial lines reaching from suture to suture. Protoconch height 315, 310 and 260 µm



respectively in paratype 2, holotype and paratype 1; width 250 µm in holotype and paratype 2 and 220 µm in paratype 1; nucleus diameter 40-50 µm. Protoconch/teleoconch boundary well marked by a slightly sinuated scar. Teleoconch consisting of 2, 2.2 and 2.7 (respectively in paratype 1, holotype and paratype 2) convex whorls rapidly growing in height, sculptured by narrow, incised spiral lines numbering 5 (paratype 1) to 7 (holotype and paratype 2) in the penultimate whorl and by moderately curved, prosocyrt growth lines. Spiral lines, under magnification higher than 40-50x, looking like a fine chain being run across by very small, irregular and narrow axial ribs, lacking on the remaining surface, and having strongly indented borders. Sutures inclined and incised. Body whorl well developed, making up about 3/4 of shell height. Aperture semicircular and broad, making up about 3/4 (2/3 in paratype 2) of body whorl height and about 1/2 of total height. Outer lip rounded, slightly retracting in its lower portion, with small rounded tubercles, mainly occurring on the central part of lip (tubercles have been observed in paratype 1 which is the only specimen with a well preserved outer lip). Columella broader and flattened towards the lowermost part, with a small umbilical chink which can be partly covered by a callus (paratype 2).

LOCUS TYPICUS

Monte Serro, Dattilo, Trapani, Western Sicily (Fig. 1), Foglio 257 IV NE (Dattilo) of Carta d'Italia (I.G.M., 1971), UTM: 33S TC 899 068.

STRATUM TYPICUM

The holotype is from a bulk sample collected at about 70 cm from the base of a yellowish silt-sandy bed Lower Pleistocene (Emilian) in age, about 1 m thick, cropping out in the south western side of Monte Serro, 170 m above sea-level (see GARILLI, 1998 for the age and the position in a geological log of the outcrop).

STRATIGRAPHICAL DISTRIBUTION

This species is known only from the Pliocene and Pleistocene of the central Mediterranean area.

PALEOECOLOGICAL OBSERVATIONS

According to the available data, all the living eulimids are parasitic in or on echinoderms (BOUCHET & WARÉN, 1986; WARÉN 1980, WARÉN & LEWIS, 1994). In particular, the species *E. mediterranea* (Monterosato) and *E. stancyki* Warén, have respectively been found on the ophiuroids *Ophioderma longicauda* (Retzius, 1805) (MIFSUD, 1995) and *Ophiolepis elegans* (Lütken, 1859) (WARÉN, 1980).

The mollusc assemblages from Monte Serro (GARILLI, 1998) and Killini (under study) are characterised by the abundance of *Bittium latreillii* (Payraudeau, 1826), *B. reticulatum* (Da Costa, 1778), *Tricolia* spp., trochids (especially *Jujubinus* spp.), rissoids, mainly *Alvania cimex* (Linneo, 1758) and *A. geryonia* (Nardo, 1847 ex Chiereghini ms.), indicative of a shallow-water depositional setting. In particular, the Monte Serro malacofauna was referred by GARILLI (1998) to an ecotone between the "Sables Vaseaux superficiels en Mode Calme" (SVMC) and the "Herbie-

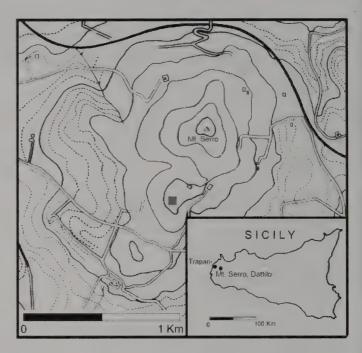


Fig.1 - Location of type locality. The shaded square indicates the fossiliferous bed of Mt. Serro.

Fig.1 - Localizzazione della località tipo. Il quadratino pieno indica l'affioramento fossilifero di Monte Serro.

re de Posidonies" (HP) biocoenoses (sensu Pérès & PICARD, 1964). Paratype 2 (Pliocene of Magliano Sabina) is from a Cladocora Ehrenberg, 1834 yellow sandy level (S. PALAZZI, pers. comm., 2000).

Paleoclimatically, it is worth remarking the occurrence of the termophilic gastropod *Jujubinus* (?) *bullula* (P. Fisher, 1877) (see RUGGIERI & UNTI, 1988, for interesting consideration about this species) in the Monte Serro and Killini assemblages, suggesting warm-temperate water conditions.

REMARKS

E. aliceae n. sp. closely resembles E. mediterranea (Monterosato) (Pl. 1, Fig. 6) from which it differs in having less numerous and more incised spiral lines (5-7 vs. 10-15 in the penultimate whorl), a less globose shell shape and well defined and rounded teeth on the outer lip (Pl. 1, Fig. 4). Furthermore, E. aliceae n. sp. has a multispiral, conical protoconch (Pl. 1, Figs 8-10), with a small nucleus, suggesting a planktotrophic larval development, while E. mediterranea has a blunt apex (pl. 1 Fig. 11) with a paucispiral protoconch (width 250-280 μm, height 210-220 μm, nucleus 70-75 μm) consisting of 1.3 to 1.5 whorls suggesting a non-planktotrophic development.

E. aliceae n. sp. is also similar to the Western Atlantic species E. stancyki Warén (WARÉN 1980, figs 12, 13) which substantially has a more slender shell, a completely smooth protoconch, a proportionally higher aperture (WARÉN, pers. comm., 1999), a different teleoconch sculptural pattern and no teeth on the outer lip. E. aliceae n.sp. is also different from E. oligocaenica Lozouet, which has a smooth outer lip, a closer-set sculpture of small



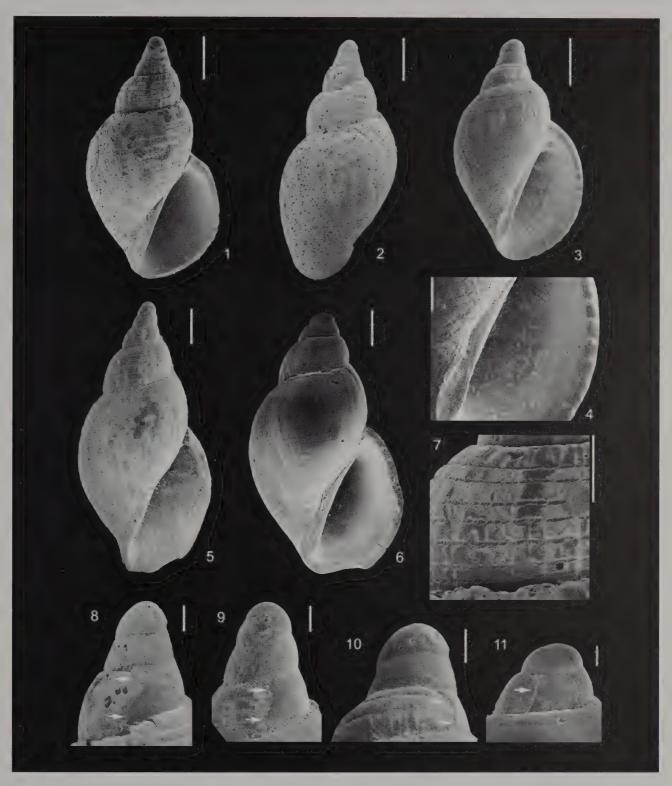


Plate 1: Figs 1, 2, 7, 8-Ersilia aliceae n. sp., holotype, Monte Serro, Dattilo, W Sicily, Lower Pleistocene; apertural view (1), dorsal view (2), sculpture of the first teleoconch whorl (7) and protoconch (8). Figs 3, 4, 10-Ersilia aliceae n. sp., paratype 1, Killini, NW Peloponnesus, Greece, Pleistocene; apertural view (3), particular of aperture (note the umbilical chink and the small teeth on the outer lip) (4) and protoconch (10). Figs 5, 9-Ersilia aliceae n. sp., paratype 2, Magliano Sabina, Roma, Central Italy, Pliocene; ventral view (5) and protoconch (9). Figs 6, 11-Ersilia mediterranea, Marina di Cinisi, Palermo, NW Sicily; apertural view (6) and protoconch (11). White arrows indicate protoconch/teleoconch boundary. Scale bars: 250 µm in Figs. 1, 2, 3, 5, 6; 100 µm in Figs. 4, 7; 80 µm in Fig. 10 and 60 µm in Figs 8, 9, 11.

Tavola 1: Figg. 1, 2, 7, 8 Ersilia aliceae n. sp., olotipo, Monte Serro, Dattilo, Sicilia occidentale, Pleistocene inferiore; vista frontale (1), vista dorsale (2), scultura del primo giro di teleoconca (7) e protoconca (8). Figg. 3, 4, 10 Ersilia aliceae n. sp., paratipo 1, Killini, Peloponneso nord occidentale, Grecia, Pleistocene; vista frontale (3), particolare dell'apertura (notare la fessura dell'ombelico e i piccoli denti sul labbro esterno) (4) e protoconca (10). Figg. 5, 9 Ersilia aliceae n. sp., paratipo 2, Magliano Sabina, Roma, Italia centrale, Pliocene; vista frontale (5) e protoconca (9). Figg. 6, 11 Ersilia mediterranea, Marina di Cinisi, Palermo, Sicilia nord occidentale; vista frontale (6) e protoconca (11). Le frecce bianche indicano il limite protoconca/teleoconca. Scala di riferimento: 250 µm in Figg. 1, 2, 3, 5, 6; 100 µm in Figg. 4, 7; 80 µm in Fig. 10 e 60 µm in Figg. 8, 9, 11.



dimples arranged in spiral lines, a more slender shell shape and a narrower aperture (LOZOUET, 1999, pl. 9, figs 8, 9; pl. 20, fig. 12).

Too many gaps in the paleontological record preclude any phylogenetic inference about $E.\ aliceae$ n. sp., $E.\ mediterranea$ (1) and the oldest European representative $E.\ oligocaenica$.

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(1) While printing, E. mediterranea has been recorded for the Pliocene of Tuscany by CHIRLI & BOGI (2002). This record points out the Pliocene coexistence of E. aliceae n.sp. and E. mediterranea.



Nomenclature of the smaller Mediterranean Cerithium species

Serge Gofas, Vittorio Garilli & Marie-Catherine Boisselier-Dubayle

KEY WORDS: Cerithiidae, Mediterranean, taxonomy, lectotype designations.

ABSTRACT

The Mediterranean morphotypes generally confused under the name Cerithium rupestre have been shown to belong to two different species. The correct name for one of the species, living in very shallow rocky areas including splashpools, is Cerithium lividulum Risso,1826, and for the other, usually living in the shallow infralittoral community of photophilous algae, C. renovatum Monterosato, 1884. Type material is figured for all nominal species in this species group introduced prior to 1900, and for some of the species proposed later. Orientations for future work in the taxonomy of this species group are suggested.

RIASSUNTO

E' stato mostrato che le entità morfologiche generalmente confuse sotto il nome Cerithium rupestre appartengono a due specie distinte. I nomi corretti per questi taxa sono Cerithium lividulum Risso, 1826, per la specie vivente su fondali rocciosi in acque più superficiali dell'infralitorale superiore e nelle pozze di marea, e Cerithium renovatum Monterosato, 1884, per la specie solitamente vivente su fondali infralitorali ricoperti da alghe fotofile. Cerithium lividulum ha una conchiglia robusta con giri appena convessi, coste assiali talora tuberculate e cordoni spirali piani. La colorazione è data da screziature bruno-verdastre decorrenti spiralmente su uno sfondo bianco. La conchiglia di Cerithium renovatum è più piccola (inferiore ai 20 mm nel materiale esaminato), più delicata e snella, e presenta una serie regolare di tubercoli, costantemente di colore bianco, che iniziano nei primi giri e raggiungono il loro massimo sviluppo in corrispondenza del penultimo. Cerithium renovatum non mostra coste assiali evidenti. Le ovature delle due specie, deposte in acquario, sono differenti. Relativamente a questi taxa, viene illustrato il materiale tipo di tutte le specie nominali introdotte prima del 1900 e di alcune proposte successivamente. Sulla base del materiale studiato e dei dati desunti dalla letteratura, viene fornito un quadro schematico della distribuzione dei due taxa. Viene inoltre suggerita una guida per futuri lavori tassonomici su questo gruppo di specie.

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INTRODUCTION

The genus *Cerithium* is represented in the Mediterranean Sea and Ibero-Moroccan area by several species, which usually occupy nearshore environments and may be locally very common. There are discrepant views on their taxonomy at the species and genus level. One extreme view is that every recognizable morph should be named as a species (e.g. LOCARD, 1902; KOBELT, 1908; MONTEROSATO, 1910; NORDSIECK, 1974). BUCQUOY, DAUTZENBERG & DOLLFUS (1884) adopted the other extreme, accepting only two valid species, one of large size, *Cerithium vulgatum* Bruguière, 1792, and a smaller one for which they used the inappropriate name *C. rupestre* Risso, 1826. This has set the keynote for over a century of Mediterranean malacology.

The correct picture nevertheless lies somewhere in between. A quite realistic view is given in the "Atlante" of GIANNUZZI-SAVELLI *et al.* (1996), but without a discussion or justification. Recent work using allozyme electrophoresis (BOISSELIER-DUBAYLE & GOFAS, 1999) has demonstrated that two different species can be distinguished in what has been known hitherto as "Cerithium rupestre" sensu B.D.D.

The purpose of this paper is to provide a basis for naming correctly the two species of "smaller" *Cerithium* present in the Western Mediterranean. It is not within the scope to revise all available names, for which more taxonomic work is needed. This revision will be restricted to the species named before 1900, and to some of the later described forms for which we have examined type material. It is based mainly on material examined in Muséum National d'Histoire Naturelle, Paris (hereafter MNHN).

TAXONOMY

The arguments for separating Cerithium species have been investigated by Boisselier-Dubayle & Gofas (1999), where we used the names C. lividulum for one species, and conservatively maintained the name "C. rupestre" for the other, which we show here to be correctly named C. renovatum Monterosato, 1884. Data from 11 informative allozyme loci vielded fixed alternative alleles in a sympatric population at les Embiez, France, for Aspartate-Aminotransferase (Aat), Glucose-6-phosphate isomerase (Gpi), Isocitrate dehydrogenase (Idh), Lactate dehydrogenase (Ldh), Mannose-6-phosphate isomerase (Mpi), D-octopine dehydrogenase (Odh), one locus of Phosphoglucomutase (Pgm-2) and one of Superoxide dismutase (Sod-2), which is overwhelming evidence for a reproductive isolation. Considering genetic distances calculated from allele frequencies, the population of *C*. renovatum clusters with the local C. vulgatum and not with C. lividulum.

There are morphological features which allow recognition of the two species (Fig. 1). *Cerithium lividulum* has a sturdy shell, with hardly convex whorls, normally no varixes along the early whorls. The sculpture consists of axial folds and of flat, poorly separated spiral cords. The axial folds sometimes bear some blunt tubercles along their midline. The external surface bears a pattern of greenish brown mottles organized along the spirals on a white background, and which may be darker on the abapical part of the whorls. The shell of *C. renovatum* is smaller (less than 20 mm in our material), more delicate and slender, with a regular series of knobs starting early on the spire and best developed on the penultimate whorl, but otherwise the whorls do not



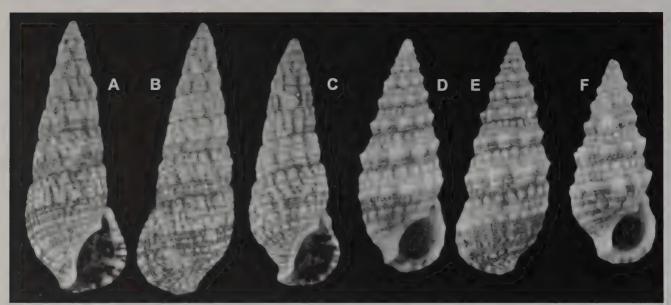


Fig.1. A, B, C: Cerithium lividulum Risso, 1826, actual sizes 24.1 and 22.1 mm; D, E, F: C. renovatum Monterosato, 1884, actual sizes 20.9 and 18.2 mm. Specimens from sympatric populations studied with allozyme electrophoresis, from Ile des Embiez, France. The specimen on Fig. D-E was maintained in aquarium for spawning (as in Fig. 2B).

Fig. 1. A, B, C: Cerithium lividulum Risso, 1826, dimensioni degli esemplari: 24,1 e 22,1 mm; D, E, F: C. renovatum Monterosato, 1884, dimensioni degli esemplari: 20,9 e 18,2 mm. Esemplari provenienti da popolazioni simpatriche (Ile des Embiez, Francia), studiati mediante elettroforesi degli alloenzimi. L'esemplare raffigurato in D-E è stato mantenuto in acquario fino alla deposizione dell'ovatura (riportata in Fig. 2B).

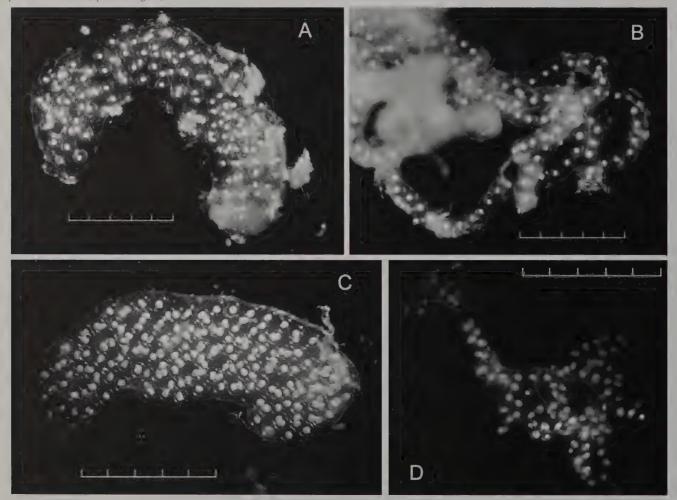


Fig.2. Egg masses of Cerithium lividulum (left) and C. renovatum (right) obtained in aquaria. A, B: from Les Embiez, France, collected in April 1999; C, D: from Palermo, NW Sicily, Italy, collected in June 2000. Scale bars are 5 mm.

Fig. 2. Ovatura di Cerithium lividulum (a sinistra) e di C. renovatum (a destra) deposte in acquario. A, B: da Les Embiez (Francia), raccolti nell'aprile 1999; C, D: da Palermo, raccolti nel giugno 2000. Scala: 5 mm.



show distinct axial folds. Compared to spinose forms attributed to *C. lividulum*, the tubercles are more aligned along a keel and are made more conspicuous by a whitish area running along them; the profile of the whorls shows a keel since very early teleoconch whorls. This paler area continues on the body whorl and interrupts the mottled pattern thereon.

The spawn of the two species is also different (Fig. 2). *Cerithium lividulum* lays a massive gelatinous mass across which 4-5 eggs may fit, whereas *C. renovatum* produces a single strand of eggs.

NOMENCLATURE

Cerithium rupestre Risso, 1826.

The specimen in Risso collection (Fig. 3) is small, slender and nodose, and belongs to the complex of *C. vulgatum*. The most similar morphology is found among morphs identified as *C. protractum* (Bivona, 1838) by GIANNUZZI SAVELLI *et al.* (1996: 26-27), which belongs to the *C. vulgatum* species group. Thus, it cannot be used for any of the two species considered here. Anyway, it is not desirable to use the name *C. rupestre* in the sense of BUCQUOY, DAUTZENBERG & DOLLFUS, 1884, since this would not be a clear statement of which species is intended. The possibility that the specimen has been substituted since Risso's publication must also be considered, although it does not essentially contradict Risso's laconic description. The specimen illustrated in Philippe Gény's plate produced ca. 1840 and published by ARNAUD (1978) is a totally different shell, probably exotic, with

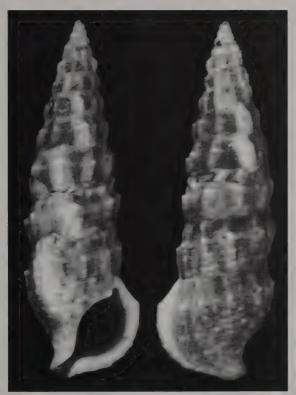


Fig. 3. The lectotype, designated by ARNAUD (1978) of Cerithium rupestre Risso, 1826. Actual size: 27 mm (photo MNHN).

Fig. 3. Lectotipo di $Gerithium\ rupestre\ Risso,\ 1826\ designato\ da\ Annaud\ (1978).$ Dimensione dell'esemplare: 27 mm (foto MNHN).

a definitely distorted siphonal canal like in *Cerithium aluco* (Linné, 1758).

Cerithium lividulum Risso, 1826 (Fig. 4)

Cerithium lividulum Risso, 1826: 154

Type locality: Alpes Maritimes, Mediterranean France

This species is represented in Risso's material, and a lectotype was designated by Arnaud (1978: 122, pl. 8, fig. 25). The name has remained largely unused, mainly because the other name *C. rupestre* was being used in its place, but there were some clues to its identity since Monterosato (1884: 120) used it as a valid name. This is the name to be applied to one of the species formerly known as *C. rupestre sensu* Bucquoy, Dautzenberg & Dollfus, 1884.

The following synonyms are considered:

Cerithium pictum Anton, 1839

This name has remained obscure but has been clarified since SCHNIEBS (2000: 81-82) designated and figured a lectotype, undoubtedly conspecific with *C. lividulum*.

Cerithium mediterraneum Deshayes, 1843: 313-314

This name was proposed in the Deshayes edition of "Animaux sans vertèbres" and the original reference cites several authors who misused the names *Cerithium tuberculatum* Lamarck (*sensu* BLAINVILLE, 1826) and *C. fuscatum* (*sensu* COSTA, 1829). It should be noted that, although PHILIPPI (1836: 193, pl. 11, fig. 7) and KIENER (1841-42: 30-31) credited the name *C. fuscatum* to O.G. Costa, this name is not separately available and is merely a misuse of *C. fuscatum* Gmelin, 1791.

Deshayes indicates for *C. mediterraneum* a habitat in small, quiet pools with abundant vegetation, which is consistent with what we know for *C. lividulum*. There is in MNHN a lot of 5 syntypes transferred in 1978 from the Deshayes collection in Ecole des Mines de Paris (Fig. 4 C-F). The specimens bear conspicuous spines on the whorls, and their genuine morphology matches that of populations collected in Northeastern Algeria, around Djidjelli (collection Etienne Nelva, MNHN).

Cerithium rupestre var. plicata Bucquoy, Dautzenberg & Dollfus, 1884: 203, pl. 23 figs 5-6.

Cerithium rupestre var. minor Bucquoy, Dautzenberg & Dollfus, 1884

The specific epithet *plicata* is preoccupied by *Cerithium vulgatum* var. *plicata* Philippi, 1836: 193, and the name *minor*, probably intended as infrasubspecific, is preoccupied many times by other varietal names. The material figured by BUCQUOY, DAUTZENBERG & DOLLFUS (1884) is anyway the basis for the widespread usage of the name *C. rupestre*, and one specimen is figured herein (Fig. 4 G-H).

Cerithium strumaticum Locard, 1886: 181-182, 565-566.

Cerithium strumaticum Locard, 1886 is based on a description, on a large series of syntypes from Mediterranean France, and on an



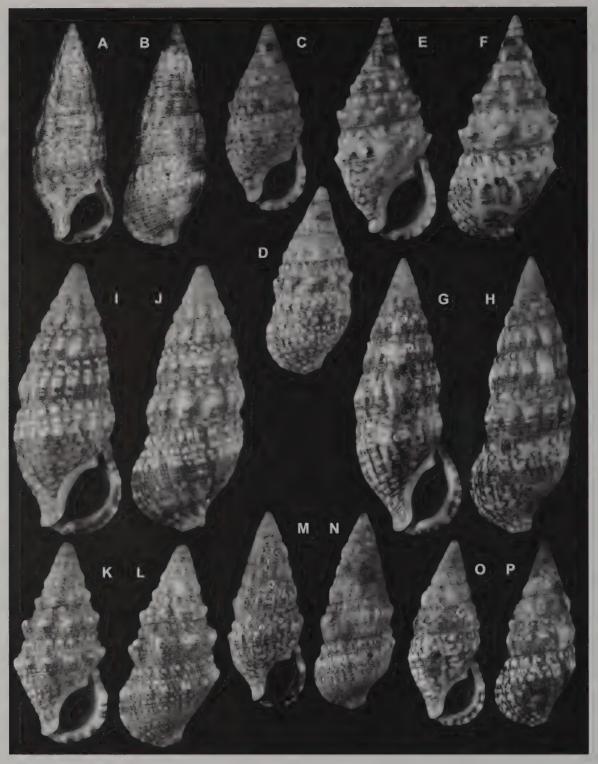


Fig. 4. Type specimens of nominal taxa assigned to Cerithium lividulum, all to scale. A, B: Lectotype of Cerithium lividulum Risso, 1826 designated by ARNAUD, 1978 (actual size 22 mm; photo MNHN). C, D, E, F: Syntypes of Cerithium mediterraneum Deshayes, 1843, no locality given (actual sizes 18.5 and 22.1 mm). G, H: Syntype of Cerithium rupestre var. plicata Bucquoy, Dautzenberg and Dollfus, 1884, from Roussillon, France (actual size 27.3 mm). I, J: Syntype of Cerithium strumaticum Locard, 1886 from Marseille, France (actual size 26.7 mm). K, L: Syntype of Cerithium massiliense from Marseille, France (actual size 20 mm). M, N: Lectotype, here designated, of Cerithium requieni Locard & Caziot, 1900 from Bonifacio, Corsica (actual size 19.2 mm). O, P: Syntype of Cerithium palustre Locard & Caziot, 1900 from Brando, near Bastia, Corsica (actual size 18.2 mm).

Fig. 4. Esemplari tipo di taxa nominali attribuiti a Cerithium lividulum; tutti alla stessa scala. A, B: lectotipo di Cerithium lividulum Risso, 1826 designato da ARNAUD, 1978 (dimensione: 22 mm; foto MNHN). C, D, E, F: sintipi di Cerithium mediterraneum Deshayes, 1843, senza località di provenienza (dimensioni: 18,5 e 22,1 mm). G, H: sintipo di Cerithium rupestre var. plicata Bucquoy, Dautzenberg e Dollfus, 1884, da Roussillon, Francia (dimensione: 27,3 mm). I, J: sintipo di Cerithium strumaticum Locard, 1886 da Marsiglia, Francia (dimensione: 26,7 mm). K, L: sintipo di Cerithium massiliense da Marsiglia, Francia (dimensione: 20 mm). M, N: lectotipo, qui designato, di Cerithium requieni Locard & Caziot, 1900 da Bonifacio, Corsica (dimensione: 19,2 mm). O, P: sintipo di Cerithium palustre Locard & Caziot, 1900 da Brando, nei pressi di Bastia, Corsica (dimensione: 18,2 mm).



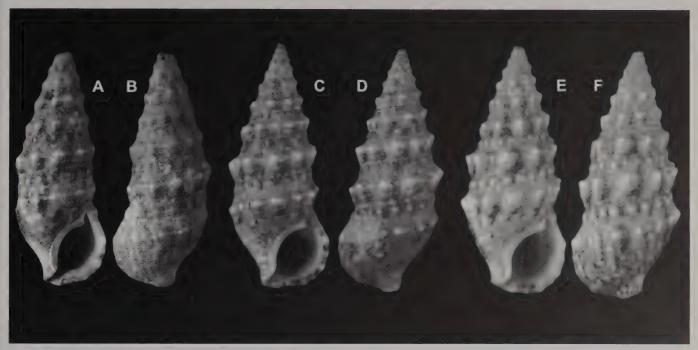


Fig. 5. Specimens of nominal taxa assigned to Cerithium renovatum, all to scale. A, B: Lectotype, here designated, of Cerithium vulgatum var. pulchella Philippi, 1836 (actual size 15.5 mm; ZMHU, Berlin). C, D: Specimen from Palermo, labeled as C. renovatum by Monterosato (actual size 16.2 mm; MNHN). E, F: Syntype of C. payraudeaui Locard & Caziot, 1900 from Brando, Corsica (actual size 16.5 mm).

Fig. 5. Esemplari di taxa nominali attribuiti a *Cerithium renovatum*; tutti alla stessa scala. A, B: lectotipo, qui designato, di *Cerithium vulgatum* var. *pulchella* Philippi, 1836 (dimensione: 15,5 mm; ZMHU, Berlino). C, D: esemplare proveniente da Palermo, classificato da Monterosato come *C. renovatum* (dimensione: 16,2 mm; MNHN). E, F: sintipo di *C. payraudeaui* Locard & Caziot, 1900 da Brando, nei pressi di Bastia, Corsica (dimensione: 16,5 mm).

explicit reference to the description and figure of *C. rupestre* var. *plicata* in BUCQUOY, DAUTZENBERG & DOLLFUS (1884). LOCARD (1886) further named two varieties *major* and *ventricosa* after B.D.D.'s figures 5 and 6. Localities given are Sète, Martigues, Marseille, Toulon, St. Nazaire (now Sanary s/mer, not to be confused with the St. Nazaire on the French Atlantic coast), La Seyne, Cannes, Menton, "etc". There are syntypes in Coll. Locard from Sète, Marseille, Sanary, Toulon, La Seyne, all conspecific, as well as non-type material from other Mediterranean localities. We illustrate here a syntype from Marseille (Fig. 4 I-J).

Cerithium massiliense Locard, 1886: 182, 566.

This is a morph which we consider conspecific with *C. lividulum*, but is characterized by a spinose ornamentation of the shell. Locard cites material from Marseille, Toulon, Nice, "etc...". There are syntypes in Coll. Locard from Marseille (Fig. 4 K-L) and Toulon, and non-type material from several other Mediterranean localities.

Cerithium palustre Locard & Caziot, 1900: 110-111.

The localities mentioned in the original description are in Corsica: Ajaccio, and Brando (a small place near Bastia, to the North; Fig. 4 O-P). These are smaller than the specimens from the same localities on which the authors recognize *C. strumaticum*, and have a quite blunt sculpture. We consider them

conspecific as well with *C. lividulum*. LOCARD & CAZIOT (1900) describe a *C. palustre* var. *elongata*, which they state being found on the coast of mainland France; this latter name is preoccupied several times (*Cerithium elongatum* Anton, 1839, *Cerithium elongatum* Sowerby G.B. II, 1855).

Cerithium requieni Locard & Caziot, 1900; p. 110.

The syntypes present in MNHN consist of three lots from Brando, Bonifacio and Pietranera, and may contain a mixture of *C. lividulum* and *C. renovatum*. One specimen from Bonifacio, which we identify as *C. lividulum*, has been selected here as lectotype (Fig. 4 M-N), in good agreement with the diagnosis which states that it is recognized by its blunt sculpture. The specimens from Brando are rather spinose and belong to *C. renovatum*.

Lithocerithium bellicosum Monterosato, 1917: 19

This species is based on Monterosato's material from Libya, but also on Figs 44-46 of Pallary (1913). It is similar to specimens collected in Djerba, Tunisia ascertained as *G. lividulum*. We do not know the whereabouts of Pallary's Egyptian material.

Cerithium archipelagicum Gaglini, 1992

This was a nomen nudum, alluded to only by MONTEROSATO (1917) in connection with *L. bellicosum*, and is based on specimens from the Greek Aegean islands. It was described by GAGLINI (1992: 126-127, 147) but introduced as a synonym,



thus not available.

Cerithium renovatum Monterosato, 1884 (Fig. 5)

Cerithium renovatum is a replacement name for Cerithium vulgatum var. pulchella Philippi, 1836, preoccupied by Cerithium pulchellum J. de C. Sowerby, 1832 and senior homonym of Cerithium pulchellum Dujardin, 1837. Thus the type material and type locality are of Philippi (1836).

Cerithium vulgatum var. pulchella Philippi, 1836: 193, pl. 11, fig. 9.

Two syntypes were found in Berlin Museum, with labels that allow to trace them to Philippi's material. We here select as lectotype (Fig. 5 A-B) the smallest specimen (15.5 x 6.1 mm), which is quite worn but agrees with the concept that MON-TEROSATO (1884) had of the species when he introduced the replacement name. It is also this specimen which best agrees with the "natural size" of 18 mm which can be measured on Philippi's plate 11, fig. 9. The other specimen is larger (24.0 x 9.4 mm) and probably does not belong to the same species. We also figure (Fig. 5 C-D) a specimen selected in a lot of 4 specimens from Palermo sent by Monterosato (with manuscript label "C. vulgatum var. pulchella Ph.") to MNHN. This and several other lots from Palermo in MNHN, (H. Fischer collection: 4 specimens; Vignal collection 5 + 3 specimens, Locard collection, 18 specimens, probably from the same source), represent an abundant material which we consider conspecific.

Cerithium payraudeaui Locard & Caziot, 1900: 109-110 (ex Monterosato ms.).

This is based on specimens from Corsica which are somewhat stouter than the Sicilian *C. renovatum*. There are five lots of syntypes in MNHN, all conspecific: Pietranera (3 specimens), Bastia (5 specimens), Brando (4 specimens; Fig. 5 E-F), St Florent (1 specimen) and Bonifacio (2 specimens). It may be attributed to *C. renovatum*, but also resembles some corsican populations which go to *C. lividulum* based on spawn and allozymes. *Thericium locardi* Nordsieck, 1974; p. 11 is an unnecessary replacement name for *Cerithium payraudeaui* Locard & Caziot 1900, which is a senior homonym of *Thericium payraudeaui* Nordsieck, 1974 ex Monterosato (nomen nudum, 1910), a representative of the *C. vulgatum* group.

Discussion of the name Cerithium aluchensis "Chiereghini"

BRUSINA (1898) suggested that *Cerithium aluchensis* "Chiereghini" should have priority over *C. renovatum*. We disagree with his view and hold *C. aluchensis* as a synonym of *C. vulgatum*. In the same paper, BRUSINA (1898) introduces in synonymy (thus, not making it available) another name *C. monterosatoi*, which he had intended at one time for what Monterosato renamed *C. renovatum*.

The name Murex aluchensis was first published by NARDO (1847: 61-62), in synonymy of Cerithium vulgatum var. tuberculata Philippi, 1836. Nardo's text is organized in two columns (pagi-

nated separately), the left one listing the names as in Chiereghini's manuscript, the right one giving the "modern synonymy" (which we understand as the names that Nardo deemed valid); when no "modern" name was stated, the Chiereghini name was formally made available with a short description. Thus, we interpret all other names as introduced in synonymy, and not immediately available according to the terms of the code of zoological nomenclature. The name *Murex aluchensis* was made available (with author and date Nardo, 1847) when BRUSINA (1898) used it as a valid name and cited Nardo.

PHILIPPI (1836), in turn, had based his *C. vulgatum* var. *tuberculata* on figure 82 of BUONANNI (1684). This figure represents a tall, nodose form of the *C. vulgatum* group. Furthermore, Nardo states that Chiereghini's material originates from the Golfo Veneto (the Northern Adriatic), where such forms are found, whereas neither *C. lividulum* nor *C. renovatum* are represented. BRUSINA (1866) stated that *C. renovatum* (which he then called *C. minutum* Serres) is found only in Southern Dalmatia, not north of Lesina (now Hvar Island, Croatia).

However, Brusina (1870) had used the name *C. aluchensis* with a different meaning. There, he did not refer to Nardo's publication, and made the name available by giving two references:

- Cerithium vulgatum var. tuberculata Philippi, 1836
- "Cerithium minutum" as figured in Sowerby, 1855: 865, pl. 181, fig. 122.

Thus the name *C. aluchensis* Brusina, 1870 ex Chiereghini ms. is separately available and is a junior homonym of *C. aluchensis* (Nardo, 1847 ex Chiereghini ms.). Also, the specimen figured in Sowerby's Thesaurus conchyliorum is not conspecific with *C. aluchensis* (Nardo, 1847). *Cerithium minutum* in SOWERBY (1855: 865) is given with a reference to Philippi and can be traced to "*C. vulgatum* var. *minuta* M. de Serres" as misapplied in PHILIPPI (1836). Thus, it is not a separately available taxon. It is a stout, spinose form resembling the types of *C. mediterraneum* (our Fig. 4 E-F), and probably belongs to *C. lividulum*.

MONTEROSATO (1899) ironically pointed out how Brusina's views on *Cerithium aluchensis* changed from 1870 to 1898, and how both interpretations disagree with Nardo's (1847) which has priority.



Fig. 6. Reproduction of fig. 82 in BUONANNI (1684), as mirror image so as to restore the aspect of the original woodcut. The figured specimen is hereby designated as lectotype of *Cerithium vulgatum var. tuberculata* Philippi, 1836, of *Murex aluchensis* Nardo, 1847 ex Chiereghini ms. and of *Cerithium aluchensis* Brusina, 1870, ex Chiereghini ms.

Fig. 6. Riproduzione della fig. 82 di BUONANNI (1684), riflessa in modo da ripristinare l'aspetto originale della xilografia. L'esemplare figurato viene qui designato come lectotipo di Cerithium vulgatum var. tuberculata Philippi, 1836, di Murex aluchensis Nardo, 1847 ex Chiereghini ms. e di Cerithium aluchensis Brusina, 1870, ex Chiereghini ms.



We hereby designate the specimen figured in BUONANNI (1684: fig. 82, here reproduced Fig. 6) as lectotype of *C. vulgatum* var. *tuberculata* Philippi, 1836, of *C. aluchensis* (Nardo, 1847 ex Chiereghini ms.) and of *C. aluchensis* Brusina, 1870, ex Chereghini ms. in order to make these taxa objective synonyms.

Other names which require further investigation.

There are many more names, all posterior to 1900, available in this species group but most of them cannot be soundly evaluated in the light of the evidence at hand. First, there is a need to settle the taxonomic issues and to make clear statements of which morphological entities represent valid species or subspecies, and this requires genetic investigation. There is at this time no evidence for the status of such forms as found in the eastern Mediterranean or the Atlantic archipelagos, which may, or may not, have genetic continuity with the two species we presently recognize in the Western Mediterranean. The following notes, not exhaustive, could be made in the course of this study of type material in MNHN.

Cerithium tingitanum Pallary, 1920 ex Monterosato ms.: 45, pl. 1, figs 3, 4.

Pallary introduced validly this name with a figure, but stated *C. renovatum* as a synonym. There are several possible syntypes of

this species in MNHN (Fig. 7 A-B), but the figured specimen was not found. We also collected specimens with a similar morphology in several localities of the Strait of Gibraltar (Barbate, Getares, Tangiers, Ceuta, M'diq), but not on the northern shore of the Alboran sea which one of us (S.G.) has intensely prospected. Their morphology is confusing, with many traits of *C. renovatum* allied to a size more usual for *C. lividulum*. One possibility is that this represents the westernmost occurrence of *C. renovatum*, in continuity with the main range of the species along the Moroccan Mediterranean coast, but these populations need to be investigated for enzyme polymorphism and spawn.

Hirtocerithium fartulum Monterosato, 1923: 9.

There are three possible syntypes (Fig. 7 C-D) from the type locality Benghazi, Libya, in Vignal collection from Monterosato. These might represent spinose morphs of *C. lividulum*, but a genetic characterization of these North African populations is needed before anything more can be said.

Cerithium syriacum Pallary, 1938: 34, pl. 1, fig. 22.

The Pallary collection in MNHN contains several syntypes (Fig. 7 G-H) from Beyrouth, Lebanon, which have a genuine mosaic-like sculpture and uniform whitish colour. There is also a need for investigation of their relationship with the main

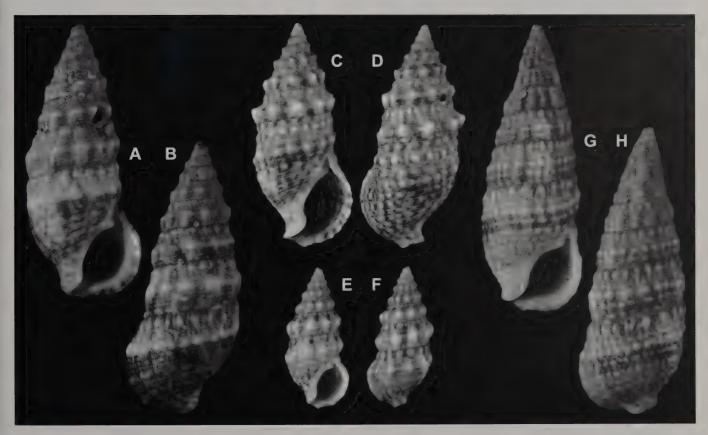


Fig. 7. Type specimens of some nominal taxa introduced after 1900, all to scale. A, B: Syntype of Cerithium tingitanum Pallary, 1920 ex Monterosato ms., from Tangiers, Morocco (actual size 22.7 mm). C, D: Possible syntype of C. fartulum Monterosato, 1923 from Benghazi, Libya (actual size 18.6 mm; specimen in Vignal collection, MNHN, obtained from Monterosato). E, F: Syntype of C. phaeniciacum Pallary, 1938 from Tartous, Syria (actual size 11.9 mm). G, H: Syntype of C. syriacum Pallary, 1938 from Beyrouth, Lebanon (actual size 24 mm).

Fig. 7. Esemplari tipo di alcuni taxa nominali introdotti dopo il 1900; tutti alla stessa scala. A, B: sintipo di Cerithium tingitanum Pallary, 1900 ex Monterosato ms., da Tangeri, Marocco (dimensione: 22,7 mm). C, D: possibile sintipo di C. fartulum Monterosato, 1923 da Bengasi, Libia (dimensione: 18,6 mm; collezione Vignal, MNHN, esemplare ottenuto da Monterosato).

E, F: sintipo di C. phaeniciacum Pallary, 1938 da Tartous, Siria (dimensione: 11,9 mm). G, H: sintipo di C. syriacum Pallary, 1938 da Beirut, Libano (dimensione: 24 mm).



range of *C. lividulum*. PALLARY (1938) also described a *Cerithium syriacum* var. *strigosa* Pallary, 1938: 34, pl. 1, fig. 13, of which we did not find any type material.

Cerithium phaeniciacum Pallary, 1938; p. 33, pl. 1 figs 7-8. This small form was described from Tartous, Syria and is morphologically very distinct from the sympatric *C. syriacum*. There is a possibility that these are the eastern Mediterranean representatives of, respectively, *C. renovatum* and *C. lividulum*, but this has to be demonstrated. There is a lot of three possible syntypes (Fig. 7 E-F) of *C. phaeniciacum* from Tartous, obtained from Pallary, in coll. Vignal, MNHN.

The following names are available for Atlantic insular populations, in which future research may find support for separating different subspecies or species.

- Thericium strumaticum canariense Nordsieck, 1974 (type locality Puerto la Cruz, Tenerife).
- Thericium medrickyi Nordsieck, 1974 (type locality Lanzarote).
- Thericium stomum Nordsieck, 1974 (type locality Lanzarote).

Distribution of the species

The maps of Figs 8-9 have been established on material examined in MNHN collection, and allow a rough representation of the distribution of these species. The distribution of *C.*

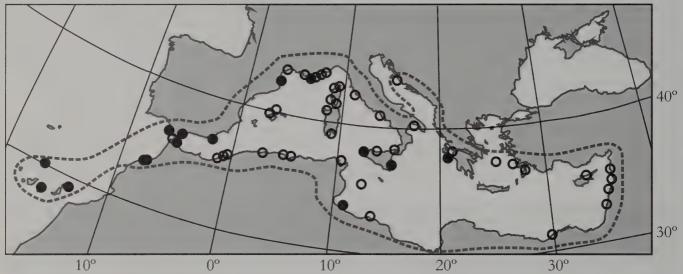


Fig. 8. Distribution of *Cerithium lividulum* according to material examined in MNHN. Open circles: material collected prior to 1950 (mostly, prior to 1900); solid circles: material collected later than 1950 (mostly, later than 1980). Broken line: generalized track deduced from this material and literature data.

Fig. 8. Distribuzione di Cerithium lividulum in base al materiale esaminato al MNHN. Cerchi vuoti: materiale raccolto antecedentemente al 1950 (ed in gran parte prima del 1900); cerchi pieni: materiale raccolto dopo il 1950 (principalmente dopo il 1980). Linea tratteggiata: distribuzione della specie in base al materiale studiato ed a dati presenti in letteratura.

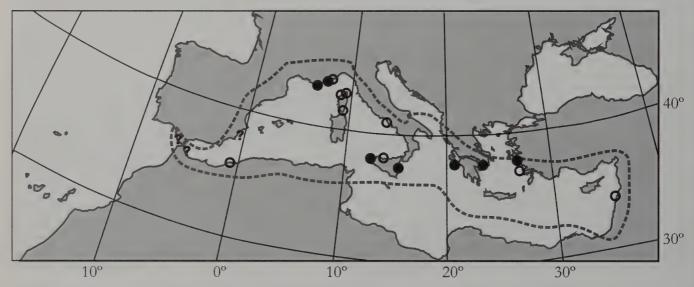


Fig. 9. Distribution of Cerithium renovatum according to material examined in MNHN. Open circles: material collected prior to 1950 (mostly, prior to 1900); solid circles: material collected later than 1950 (mostly, later than 1980). Broken line: generalized track deduced from this material and literature data. The (?) refer to localities with C. tingitanum morphs.

Fig. 9. Distribuzione di Cerithium renovatum in base al materiale esaminato al MNHN. Cerchi vuoti: materiale raccolto antecedentemente al 1950 (ed in gran parte prima del 1900); cerchi pieni: materiale raccolto dopo il 1950 (principalmente dopo il 1980). Linea tratteggiata: distribuzione della specie in base al materiale studiato ed a dati presenti in letteratura. I punti interrogativi si riferiscono a località in cui sono stati rinvenuti esemplari atribuibili al morfotipo C. tingitanum.



lividulum is the broadest, with a range extending from the Eastern Mediterranean to the Canary Islands, Selvagens and Madeira. The conspecificity of the macaronesian populations on one side, of Eastern mediterranean populations on the other needs confirmations, but populations on the Atlantic coast of Morocco agree in morphology and habitat with those of the Western Mediterranean, and the population from Malaga province has been checked with allozyme electrophoresis. Some gaps in the distribution may reflect a lack of data but others are real. The species has been searched for, and not found, on the rocky shore of Granada province, where C. vulgatum is common. It seems absent from the northernmost Adriatic (Brusina, 1866; Cossignani et al., 1992); the only Adriatic sample in our material originates from Veli Rat, near Zadar, Croatia (Vignal collection, MNHN).

The distribution of *C. renovatum* is more restricted. It includes the Mediterranean coast of France, Corsica, and Sicily. Smaller morphs which may be this species are found in the Eastern Mediterranean, in Greece and (as *C. pheniciacum*) on the Levantine coast. If *C. tingitanum* were to be proved conspecific, the range would include also the Straits of Gibraltar; the continuity with other parts of the range would be through North African populations, given that the species is absent on the shores of Málaga and Granada provinces.

ACKNOWLEDGEMENTS

We are extremely grateful to Matthias Glaubrecht, curator of Mollusca at the Museum fur Naturkunde, Berlin, for making available to us the syntypes of *C. vulgatum* var. *pulchella*, to Luca Galletti (Monreale, Palermo) for providing the photos of the egg masses coming from Sicily, to Sarah Samadi (Institut de Recherche pour le Développement) for the egg masses coming from France, and to Stefano Palazzi (Modena) for help in the bibliographical search. The contribution of one of us (V.G.) was carried out in the Muséum National d'Histoire Naturelle thanks to funds of the European Commission (Environment and Climate Program-Marie Curie, contract n° ENV4-CT98-5125).

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Lavoro accettato il 15 marzo 2002



Prima segnalazione di Mathilda barbadensis Dall, 1889, in Mediterraneo (Heterostropha, Mathildidae)

Romualdo Rocchini

KEYWORDS

Mathildidae, Mar Tirreno, new findings, thanatocoenose

ABSTRACT

Mathilda barbadensis Dall, 1889, is reported for the first time in the Mediterranean Sea on the base of two empty shells collected in the Tuscan Archipelago (Gorgona Is.) and Sicily (Cape S. Alessio). The specimen from the Tuscan Archipelago was collected at a depth of 300-450 m, together with four specimens of Tuba jeffreysi (Dall, 1889), all inside an amphora fragment. The specimen from Sicily was collected on a silty sand bottom at 210 m. With this new record, the number of species of the family Mathildidae known in the Mediterranean Sea can be fixed at seven. However, since both specimens were dead collected, it is still not possible to undoubtely ascertain that M. barbadensis could be a member of the present living fauna

RIASSUNTO

Si riportano qui due ritrovamenti dall'Arcipelago Toscano (isola di Gorgona) e dalla Sicilia (Capo S. Alessio) di conchiglie prive di parti molli di Mathilda barbadensis Dall, 1889.

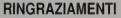
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Le specie della famiglia Mathildidae finora note per il Mediterraneo (OLIVERIO & NOFRONI, 1986, 1988; GIUSTI & NOFRONI, 1995) sono sei, cinque attribuite al genere Mathilda Semper, 1865, ed una al genere Tuba Lea, 1833 (su un totale di nove specie finore note per il NE Atlantico: CLEMAM., http://www.mnhn.fr/base/malaco.html).

In un campione di sedimento detritico-fangoso rinvenuto all'interno di un'anfora pescata ad una profondità di 300-450 m sui fondali circostanti l'isola della Gorgona (Arcipelago Toscano: Fig. 1), sono stati rinvenuti quattro esemplari di Tuba jeffreysi (Dall, 1889) (Fig. 9) ed un esemplare di una specie del genere Mathilda, simile per alcuni versi a M. coronata Monterosato, 1875, ma distinta per il differente rapporto altezza/diametro e per la differente disposizione

della scultura spirale.

Questo esemplare è decisamente ricollegabile a Mathilda barbadensis Dall) (1889 (Fig. 2-3-6-7-8). Questo taxon era già stato riportato per il Mediterraneo da PIANI (1981), che citò barbadensis proprio tra i sinonimi di M. coronata. Successivamente OLIVERIO & NOFRO-NI (1986) assimilarono barbadensis a M. gemmulata Semper, 1865 (Fig. 4). In seguito (OLIVERIO & NOFRONI, 1988) tale sinonimia fu rimossa, risultando gemmulata (= canariensis Dautzenberg, 1889, = haasi Mienis, 1978, = vanaartseni De Jong & Coomans, 1988) una specie ad ampia distribuzione (dai Caraibi al Mediterraneo orientale) ma di acque più superficiali, e decisamente distinta morfologicamente. Un secondo esemplare, anch'esso privo di parti molli è stato rinvenuto (Dr. S. Giacobbe, Messina: com. personale) presso Capo S. Alessio, in un ricco campione fango-sabbioso (biocenosi del detritico del largo) da 210 m di profondità. Si illustra qui l'esemplare rinvenuto alla Gorgona. La vera M. barbadensis non era perciò stata rinvenuta effettivamente in Mediterraneo finora. I due esemplari oggetto di questa nota sono stati raccolti privi di parti molli, quindi l'appartenenza di M. barbadensis alla fauna attuale mediterranea non può essere confermata.



Si ringraziano Antonio Bonfitto (Bologna) e Bruno Dell'Angelo

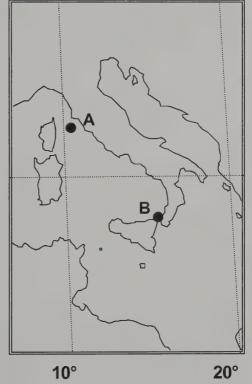


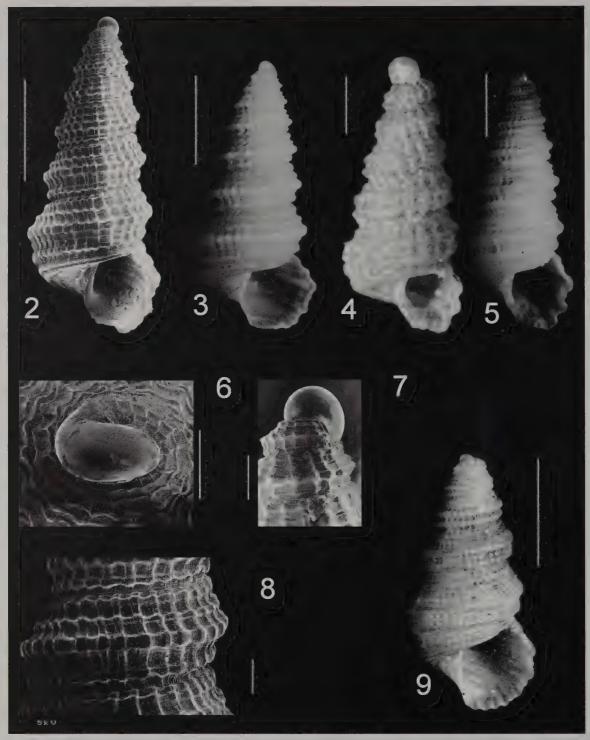
Fig. 1. Località di rinvenimento di Mathilda barbadensis in Mediterraneo. A: Is. della Gorgona (Arcipelago Toscano). B: Capo S. Alessio.

Fig. 1. localities where Mathilda barbadensis specimens were recorded in the Mediterranean Sea. A: Gorgona Island (Tuscan Archipelago). B: S. Alessio Cape.

(Prato) per l'assistenza nella realizzazione delle foto al SEM, Italo Nofroni e Marco Oliverio (Roma) per le informazioni su M. barbadensis, Salvatore Giacobbe (Messina) per la segnalazione del secondo esemplare, ed infine i fratelli Roberto e Sandro Mancini per la pazienza nel fornirmi il materiale di ricerca.

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Figg. 2-9. Mathildidae da Is. della Gorgona. 2-3: Mathilda barbadensis; 4: M. gemmulata; 5: M. cochlaeiformis; 6-7: due viste della protoconca di M. barbadensis; 8: dettaglio della scultura di un giro mediano di M. barbadensis; 9: Tuba jeffreysi. Scale: 3 mm (2-5, 9), 500 μm (6-8).

Figs 2-9. Mathildidae from Gorgona Island. 2,3: Mathilda barbadensis; 4: M. gemmulata; 5: M. cochlaeformis; 6,7 two views of the protoconch of M. barbadensis; 8: detail of the sculpture of a middle whorl of M. barbadensis; 9: Tuba jeffreysi. Scale: 3 mm (2-5, 9), 500μ (6-8).

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A revision of Marginella reeveana Petit, 1851

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KEY WORDS: Marginellidae, Marginella, endemism, Ghana.

ABSTRACT

Marginella reeveana Petit, 1851 is revised on the basis of its type material. A lectotype is appointed and a new type locality is established. The variability of the shell is displayed. The distribution and the relationships of the species are discussed. M. reeveana is considered as an isolated southern representant of a widely distributed tethyan group, ranging in the recent from the Arabian sea to West Africa.

RIASSUNTO

La specie Marginella reeveana Petit, 1851 viene ridescritta sulla base dell'esame del materiale tipico e di nuovi ritrovamenti. Tra i tre sintipi conservati presso il Natural History Museum di Londra (NHM), viene designato un lectotipo. La località tipo originale "Guinea", viene qui corretta con "Ghana", un tempo Guinea Britannica, considerando l'apparente assenza della specie nell'attuale Guinea. La distribuzione della specie, la variabilità della conchiglia e le relazioni con specie simili vengono discusse. In particolare, M. reeveana sembra essere endemica delle coste del Ghana e potrebbe rappresentare il relitto tetideo più meridionale di un gruppo di specie ad ampia diffusione, il "M. denticulata complex", i cui rappresentanti attuali sono ora distribuiti dal Mar Arabico fino all'Africa Occidentale. M. reeveana può essere facilmente distinta da queste specie affini, grazie alla presenza di alcune peculiarità morfologiche: i) coste assiali più numerose che si estendono lungo tutta la lunghezza della conchiglia, ii) profilo più arrotondato e piriforme, iii) protoconca più grande, iv) labbro più inspessito.

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INTRODUCTION

The species *Marginella splendens* Reeve, 1842 was based on 3 shells belonging to the renowned collection of Hugh Cuming (1791-1865) and said to originate from 'Guinea'. *M. reeveana* was given by PETIT (1851) as replacement name for *M. splendens* Reeve (non Grateloup, 1834).

Despite the many important samplings of Marginellidae made off the coasts of West Africa (including the Guinean coast and the Gulf of Guinea) during the 150 years following its description, the species has only ever been known from the three syntypes in the Natural History Museum, London. It is hardly surprising therefore that the Guinean origin of this species has been doubted, with some workers tending to believe in a varietal status of the taxon, whose shell morphology and pattern of decoration strongly resembles the species *M. denticulata* (Link, 1807). The latter is widespread off central Senegal (Dakar and Petite Cote) and displays a very high variability of both shell morphology and decoration rarely seen in the family Marginellidae. Some similar forms are known from Casamance (southern Senegal) to Sierra Leone. It can be assumed that several sibling species may well be separated in the future from this *M. denticulata* complex.

During the years 1985-86, the second author obtained a lot of 9 live collected specimens, trawled up by commercial fishing operations in 40-60 metres off Elmina, central coast of Ghana. Due to the rapid exhaustion of fish stocks, the trawlers soon left the area and the supply of *M. reeveana* samples came to an end. This restricted lot of shells does, however, enable us to have a more detailed look at the variability of the species and its likely distribution whilst allowing us to review its taxonomic status.

ABBREVIATIONS

NHM Natural History Museum, London.
MC Museum of Hugh Cuming
AWC A. Wakefield collection
PRC P. Ryall collection

spm live collected specimen

SYSTEMATICS

Genus: Marginella Lamarck, 1799 [Type species, by monotypy: *Voluta glabella* Linné, 1758.]

Marginella reeveana Petit, 1851 (Figs 1-9)

Marginella splendens REEVE, 1842 b, p.249, pl. 277, figs 2-3. Marginella reeveana PETIT, 1851, p.51, replacement name for Marginella splendens Reeve, 1842, non-Marginella splendens Grateloup, 1834.

Type material. 3 syntypes in NHM (Figs 1-3), with several accompanying labels. Original label: "Marginella splendens Reeve Type Guinea M.C." Recent label by K. Way, present curator of Mollusca: "BM(NH) reg. no. 1975008 Syntypes Conch. Icon. 15 Marginella Sp. 30 1865 Guinea. Hugh Cuming Colln. 3 spcs Acc. N. 1829".

As noted by TOMLIN (1917), the largest syntype of *M. splendens* is evidently the figured type. This largest specimen (Figs 1,2) is hereby appointed as lectotype of *Marginella reeveana* Petit, 1851, replacement name of *M. splendens* Reeve, 1842.



Other material examined. Ghana, off Elmina, 40-60 m: 8 spm, PRC; 1spm, AWC.

Type locality. "Guinea". According to the apparent absence of the species off the present state of Guinea and its occurrence off the present state of Ghana (formerly British Guinea), the type locality is here confirmed as "Guinea, present state of Ghana, West Africa".

Original description. In his Conchologica Systematica, Reeve (1842 b: 249) quotes his new species as 'Marginella splendens, (Humphrey, MSS). Nobis, Proceedings Zool. Soc., 1842' and illustrates it with two figures (pl. 277, figs 2,3: dorsal and ventral views) corresponding to our lectotype (Figs 1,2). No further description is given.

The reference made by REEVE to the Proceedings constitutes a mistake, as the corresponding article (1842 a) does not deal with *Marginella splendens*, nor indeed with any other Marginellidae. Tomlin (1917) has confirmed the quotation and drawings in Conchologica Systematica (REEVE, 1842 b) as constituting the original description.

Complementary description. SOWERBY (1846: 375) gave the first consistent definition of the species on the basis of the type material, and illustrated both the largest syntype (1846: pl. 74, fig. 23) in ventral view (Fig. 1) and the medium sized syntype (pl. 74, fig. 24) in dorsal view (Fig. 3):

« 7. Marginella Splendens (pl. LXXIV. f.23, 24), Reeve, Zool. Proc. 1842, and Conch. Syst.

M. ovali, creberrimè plicata, pallidè rosea, griseo vel rubro nebulata, maculis griseis vel roseis curvatis per seriebus tribus dispositis; spira producta, anfractibus rotundatis; columella plicis quatuor sub-quadratis ultimus duabus qbliquis; labio externo anticè emarginato, intùs crenulato, extùs punctato.

Like *M. glabella* in general form, but finely ridged, and having three rows of curved spots on the back, besides the minute dots which cover the shell. In Mr. Cumings Collection; from Guinea"

PETIT (1851: 51) implicitly proposed that *M. reeveana* as a replacement name for *M. splendens*, without giving any further description or figure.

Redescription. (on the basis of the new material examined, Figs 1-9): The shell has 3.5 whorls and presents a pyriform outline. Protoconch large and pronounced. Spire small, triangular and stepped, the whorls are inflated, the shoulder is strongly rounded, the surface of the entire body whorl is sculptured by narrow, tightly packed axial ribs. Outer lip thick and arched, 16-20 strong labial teeth, 4 strong and sinuous columellar plications. The shell is covered by minute black dots, regularly distributed along rows in spiral alignment. The decoration of the body whorl is completed by 2 spiral rows of chevrons oriented towards the left and separated by a large spiral colour band situated at the level of the upper part of the aperture. The spire whorls show a row of chevrons which may be partially hid-

den under the suture above. The inside of the siphonal canal bears a dark colour patch along its border. The colour decoration ranges from pinkish to orange-brown or greyish-blue, on a whitish to light yellowish background.

Dimensions. Shell measurements of the syntypes: 23.8 x 13.5mm (Figs 1,2), 19.4 x 11mm (Fig. 3), 14.9 x 9.7 mm. Other material studied: L=19 to 25.25 mm.

Distribution. The species seems to be endemic from the central coast of Ghana (not known from off Ivory Coast nor from the eastern part of the Gulf of Guinea).

DISCUSSION

The new material studied shows itself to be very similar to the type material, and *M. reeveana* therefore seems to be a very constant species. The morphology of the shell exhibits several original characters which allow it to be distinguished from *M. denticulata 'sensu auctorum'*: the axial ribs are more numerous and extend the length of the shell, the outline is more pyriform and rounded, the protoconch is larger and the labrum is more thickened. These features are not found in the numerous intra & interpopulational variations observed within the *M. denticulata* complex ranging in the Senegal/Sierra Leone area, and *M. reeveava* can be strictly separated on the basis of these shell morphology particularities. The decoration however is the same as that observed throughout the full range of variation observed in *M. denticulata* from central Senegal.

Any form of *M. denticulata* and any other close relatives are known off Ivory Coast or from other places in the Gulf of Guinea, where *M. reeveana* appears as being the single representant of the *M. denticulata* complex.

The endemic status of *M. reeveana* in Ghana is not entirely a surprise as the region has special biogeographic significance. The coast of Ghana belongs to the "atypical tropical region" ranging from Cape Palmas to Benin, and is characterized by a rather high productivity of demersal species (1.2 mt/km²) compared to the surrounding regions (LE LOEUFF & VON CONSEL, 1998: 318). This higher productivity seems to be correlated to a higher diversity.

Due to the seasonal alternance of cold upwelling currents, the region has maintained a noticeable incidence of species showing strong affinities with the recent temperate European fauna, together with representatives of intertropical groups resulting from the Pliocene Euro-West African stock. Within this region, the coast of Ghana can be considered (LE LOEUFF & VON CONSEL, 1998: 319) as a "relict pocket" for the old tethyan fauna.

M. reeveana must be considered as belonging to this case, being the most southern ranging representative of the M. denticulata complex, and showing strong affinities with M. obtusa Sowerby, 1846 from the Arabian Sea, which is apparently associated itself with several sibling species in the same area. It can be assumed that M. reeveana is a representant of a widely distributed tethyan group of Marginella, which is being conserved off West Africa through a fragmented set of species. The M. denticulata complex





Figs 1-9: Marginella reeveana Petit, 1851. Figs 1,2: lectotype NHM, 23.8 x 13.5 mm. Fig.3: paralectotype NHM, 19.4 x 11.0 mm. Fig. 4: PRC, 21.85 x 13 mm. Fig.5: AWC, 19.75 x 11.6 mm. Fig.6: PRC, 25.25 x 15.30 mm. Fig.7: PRC, 23.95 x 13.70 mm. Fig.8: PRC, 23.7 x 14.30 mm. Fig.9: PRC, 24.2 x 14.10 mm.

Figg. 1-9: Marginella reeveana Petit, 1851. Figure 1-2: lectotipo NHM, 23.8 x 13.5 mm. Fig.3: paralectotipo NHM, 19.4 x 11.0 mm. Fig. 4: PRC, 21.85 x 13 mm. Fig.5: AWC, 19.75 x 11.6 mm. Fig.6: PRC, 25.25 x 15.30 mm. Fig.7: PRC, 23.95 x 13.70 mm. Fig.8: PRC, 23.7 x 14.30 mm. Fig.9: PRC, 24.2 x 14.10 mm.



and the M. obtusa complex are under study by the first author.

ACKNOWLEDGEMENTS

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Djeddilia and Escoffieria, abnormal specimens of Potamididae (Gastropoda: Caenogastropoda: *Potamides, Tympanotonos*)

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KEY WORDS: Mollusca, Djeddilia, Escoffieria, Potamides, Tympanotonos, taxonomy.

ABSTRACT The relationships of the enigmatic modern Red Sea genus *Djeddilia* with the Pliocene genus *Escoffieria* are discussed. Both *Djeddilia* and *Escoffieria* (a junior synonym of *Potamides*) were based on abnormal, partially uncoiled specimens. *Djeddilia djeddilia* is probably a synonym of *Potamides conicus* and in consequence *Djeddilia* would be another junior synonym of *Potamides*.

RIASSUNTO Jousseaume, nel corso della sua lunga carriera di malacologo, ha descritto molti taxa problematici di difficile interpretazione. Nel corso del presente

lavoro vengono discusse le relazioni fra un enigmatico genere del Mar Rosso, *Ojeddilia*, e il genere pliocenico *Escoffieria*. L'olotipo di *Ojeddilia djeddilia* Jousseaume, 1894, fu rinvenuto a Djedda o Jiddah (Coste del Mar Rosso, Arabia), ed è caratterizzato da giri disgiunti e da una conchiglia parzialmente svolta. La stessa morfologia caratterizza anche il genere *Escoffieria* Fontannes, 1880 (specie tipo: *Escoffieria fischeri* Fontannes, 1880), descritto per i depositi pliocenici della Francia sud orientale. Sia *Djeddilia* che *Escoffieria* (che è sinonimo più giovane di *Potamides*) furono istituiti su esemplari anormali, a conchiglia parzialmente svolta. *Djeddilia* die *diddilia* è probabilmente un sinonimo di *Potamides conicus* e, di conseguenza, *Djeddilia* sarebbe un altro sinonimo di *Potamides*. Altre specie di *Potamididae*, a partire dal genere miocenico *Tympanotonos* fino ai recenti *Potamides* e *Batilla-ria* possono presentare spettacolari deformazioni della conchiglia probabilmente causate, durante la crescita della stessa, da fenomeni di stress ambientale (ad esempio crisi anossiche) molto frequenti in un ambiente variabile quale quello occupato da questa famiglia di gasteropodi.

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INTRODUCTION

Various problematic genera or species have been described by Jousseaume. However Kase & Valdés (1997) have shown that Bertinia bertinia Jousseaume, 1883 is based on the spatula shell layer of the patellogastropod Cellana nigrolineata (Reeve, 1854). Van Aartsen (2001) focused our attention on an other enigmatic gastropod described by Jousseaume (1894) from Djedda or Jiddah (Red Sea coast, Arabia). The specimen is the holotype of Djeddilia djeddilia Jousseaume, 1894, which is characterised by disjointed whorls and a partially uncoiled shell. In the original description the systematic position of Djeddilia djeddilia was not indicated. However, Jousseaume (1894) compared it with the Eocene genus Velainella (Trochidae) and with Columna Perry, 1811 (Achatinidae). LAMY (1932) evoked a distant similarity with Seguenzia laxa Jeffreys, 1885 (which has the shape of species of Skeneidae or Haloceratidae) or with Pyrgulina (?) (Pyramidellidae). VAN AARTSEN (2001) excluded a position in the Pyramidellidae and suggested a relationship with the family Vermetidae. In fact, that author did not realizes that, as already indicated by Lamy, this species was probably based on an abnormal specimen. In this paper, the systematic position of Djeddilia is revaluated after comparison with fossil Potamididae.

RESULTS

Recently, in our monograph of the fauna of the Aquitanian Stage (Lozouet *et al.* 2001), we discussed the position of various species of Potamididae. We noted that the especially large collections of Miocene *Tympanotonos* specimens, generally include a few very

deformed specimens, among which some specimens have partially uncoiled whorls. Cossmann & Peyrot (1922) assigned these specimens to the genus Escoffieria (as Escoffieria degrangei Cossmann & Peyrot, 1922). We collected specimens very similar to E. degrangei (Fig. 8) that belong without doubt to the species Tympanotonos tournoueri (Mayer, 1878) (see Lozouet et al. 2001, pl. 9, figs 7-9). The genus Escoffieria Fontannes, 1880 (type species: Escoffieria fischeri Fontannes, 1880) has been described from Pliocene deposits of southeastern France. As indicated by Fontannes the unique(?) specimen of Escofferia fischeri (Figs 1-2, 7) was collected from a bed that contains principally Potamides species [supposed to be P. granosus (Borson, 1821) and P. graecus (Deshayes, 1832)]. In the opinion of Fontannes (1879-1882), Escoffieria is a "section" of Potamides characterised by an uncoiled shell. Escoffieria degrangei is very similar to E. fischeri (see Figs 7-8) and both are based on abnormal specimens of Potamididae (Potamides and Tympanotonos). In turn, Djeddilia djeddilia closely resembles a broken specimen of Escoffieria (Figs 5-6, 7, 8). As indicated by LAMY (1932), the columellar part is "divisé en deux lèvres par un sillon longitudinal" (divided in two lips by a longitudinal groove), it is clearly not at all a columellar fold but the consequence of the shell defomation.

One Recent species of Potamididae, generally identified as *Pirenella conica* (Blainville, 1826), is extremely abundant on the Red Sea shores. LOZOUET (1986) showed that *Pirenella* Gray, 1847 (type species: *Cerithium conicus* Blainville, 1826) is a junior synonym of *Potamides* Brongniart, 1810, and that the Mediterranean



and Red Sea species *Potamides conicus* (= *Cerithium cailliaudi* Potiez & Michaud, 1833) is very similar to the Pliocene species *P. graecus* (Deshayes, 1832). A close relationship with the type species of *Potamides* (*P. lamarcki* Brongniart, 1810 from the Oligocene of France) was also pointed. *Potamides conicus* is well known for this remarkable tolerance of temperature and salinity fluctuations and exhibits a high variability of the shell sculpture (Plaziat 1989). *Potamides conicus* occurs at Djedda, and in the Jousseaume Collection (in MNHN) there is at least one sample of shells with a small size, similar to that of the holotype of *Djeddilia djeddilia*. Another numerous sample of *Potamides conicus* from Djibouti, also similar in size to the Djedda specimens, is present in the Jousseaume Collection. Some specimens from Djibouti (Fig. 3)



Figs 1-8, Escoffieria, Potamides, Tympanotonos and Djeddilia species.

Figg. 1-8, specie di Escoffieria, Potamides, Tympanotonos e Djeddilia.

Figs 1-2, 7, Escoffieria fischeri Fontannes [= ?Potamides granosus (Borson)], Pliocene (France); by Fontannes (1880, pl. 10, fig. 19), height: 16 mm.

Figg. 1-2, 7, Escoffieria fischeri Fontannes [= ?Potamides granosus (Borson)], Pliocene (Francia); da Fontannes (1880, tav. 10, fig. 19), altezza: 16 mm.

Fig. 3, Potamides conicus (Blainville), Red Sea (Djibouti), coll. Jousseaume (MNHN-Paris), specimen with a slightly distorted early spire, height: 3.2 mm.

Fig.3, Potamides conicus (Blainville), Mar Rosso (Gibuti), coll. Jousseaume (MNHN-Parigi), esemplare con le spire iniziali leggermente distorte, altezza: 3,2 mm.

Fig. 4, Tympanotonos tournoueri (Mayer), Aquitanian Stage, Saucats (Lariey) (France), coll. MNHN-Paris, height: 11 mm.

Fig. 4, Tympanotonos tournoueri (Mayer), Aquitaniano, Saucats (Lariey) (Francia), coll. MNHN-Parigi, altezza: 11 mm.

Figs 5-6, *Djeddilia djeddilia* Jousseaume, 1894, holotype (6 by Lamy, 1932) [= ?*Potamides conicus* (Blainville)], Red Sea (Djedda, Arabia), coll. Jousseaume (MNHN-Paris), height: 3.3

Figg. 5-6, Djeddilia djeddilia Jousseaume, 1894, olotipo (6 in Lamy, 1932) [= ?Potamides conicus (Blainville)], Mar Rosso (Gidda, Arabia Saudita), coll. Jousseaume (MNHN-Parigi), altezza: 3,3 mm.

Fig. 8, Escoffieria degrangei Cossmann & Peyrot, 1922 [= Tympanotonos tournoueri (Mayer)], Lower Miocene (Aquitanian), Saucats (Lariey), France, coll. MNHN-Paris, height: 6.5 mm. Fig. 8, Escoffieria degrangei Cossmann & Peyrot, 1922 [= Tympanotonos tournoueri (Mayer)], Miocene Inferiore (Aquitaniano), Saucats (Lariey), Francia, coll. MNHN-Parigi, altezza: 6,5 mm.

show a slight deformation leading to a more slender and distorted apex.

CONCLUSION

These examples reveal that different species of the genera *Tympanotonos* and *Potamides* may have spectacular, recurrent deformations in their shell growth that are probably a consequence of stress in a fluctuating environment. Thus, Furota *et al.* (2002) have shown similar deformations of the shells of potamids snail *Batillaria multi-formis* (Lischke, 1869) in severe hypoxia habitats.

Morphs with disjointed whorls and partially uncoiled shells are certainly the most fascinating monsters, and it is not surprising that they received different generic names. The unique specimen of *Djeddilia djeddilia* has been collected on a beach and is very abraded. We may therefore infer that *Djeddilia djeddilia* is likely to be founded on an abnormal, incomplete, partially uncoiled specimen of *Potamides conicus* (Blainville). We may accordingly conclude that *Djeddilia* Jousseaume, 1894 and *Escoffieria* Fontannes, 1880 are simply synonyms of *Potamides* Brongniart, 1810.

ACKNOWLEDGEMENTS

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Lavoro accettato il 2 Gennaio 2003



Fifteen new species and ten new records of *Turbonilla* Risso, 1826 (Gastropoda, Heterobranchia, Pyramidellidae) from Brazil

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KEY WORDS: Pyramidellidae, Turbonilla, taxonomy, Brazil, South America, new species.

ABSTRACT

Fifteen new species of Turbonilla from Brazilian coast are described: Turbonilla farinatiae n. sp., T. goytacazi n. sp., T. farroupilla n. sp., T. lepta n. sp., T. parviscymna n. sp., T. fluminensis n. sp., T. aracruzensis n. sp., T. midas n. sp., T. maestratii n. sp., T. kaapor n. sp., T. capixaba n. sp., T. scapulata n. sp., T. paulinoi n. sp., T. rhachialis n. sp., T. uaca n. sp. The following species are for the first time recorded from the Brazilian coast: T. rhabdota (Watson, 1886), T. aff. zulmae Pimenta & Absalão, 1998, T. aff. enna Bartsch, 1927, T. aff. unilirata Bush, 1899, T. krebsii (Mörch, 1875), T. pupoides (d'Orbigny, 1842), T. portoricana Dall & Simpson, 1901, T. stimpsoni Bush, 1899, T. aff. riisei (Mörch, 1875), T. aff. anira Bartsch, 1927.

RIASSUNTO

Nonostante siano note ben 324 specie di "Turbonilla" per le coste dell'Atlantico occidentale, solo venti di queste sono state finora riportate per le coste brasiliane. Nel presente contributo, quindici nuove specie di Turbonilla vengono descritte per il Brasile: Turbonilla farinatiae n. sp., T. goytacazi n. sp., T. farroupilha n. sp., T. lepta n. sp., T. parviscymna n. sp., T. fluminensis n. sp., T. aracruzensis n. sp., T. midas n. sp., T. maestratii n. sp., T. kaapor n. sp., T. capixaba n. sp., T. scapulata n. sp., T. paulinoi n. sp., T. rhachialis n. sp., T. uaca n. sp.. Vengono inoltre riportate alcune nuove segnalazioni di specie appartenenti al genere Turbonilla, originariamente descritte per la Florida, i Caraibi e le coste dell'Argentina: Turbonilla rhabdota (Watson, 1886), T. aff. zulmae Pimenta & Absalão, 1998, T. aff. enna Bartsch, 1927, T. aff. unilirata Bush, 1899, T. krebsii (Mörch, 1875), T. pupoides (d'Orbigny, 1842), T. portoricana Dall & Simpson, 1901, T. stimpsoni Bush, 1899, T. aff. riisei (Mörch, 1875), T. aff. anira Bartsch, 1927.

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INTRODUCTION

The taxonomy of the genus *Turbonilla* from the Brazilian coast is poorly understood, in contrast to the coasts of Africa and Europe, where recent syntheses on *Turbonilla* have been carried out (e.g., Aartsen, 1981; Schander, 1994; Peñas *et al.*, 1996; Peñas & Rolán, 1997). This lack can be addressed to the absence of a major taxonomic review on the family Pyramidellidae, added to the fact that the family is considered one of the most taxonomically confused and poorly known families of marine gastropods. Indeed, taxonomic studies on the Pyramidellidae in a region where it is poorly studied often acknowledge the existence of several undescribed species (e.g. Schander, 1994; Peñas & Rolán, 1997).

In spite of the very large number of "turbonillid" species described from the western Atlantic (324 species listed in Odé, 1996), only 20 species have been reported from the Brazilian coast (Rios, 1994; Absalão *et al.*, 1996; Pimenta & Absalão, 2001; Pimenta & Absalão, 2002).

Recent collections-based studies on pyramidellids from Brazil led us to recognize a large number of undescribed species of the genus *Turbonilla*, and some new records of species of the same genus, originally described from Florida, the Caribbean, and the coast of Argentina.

MATERIALS AND METHODS

Optical photographs were taken through a Zeiss SV-11 microscope. Scanning Electronic photographs were taken by: Zeiss LEO 1450 VP, at "Laboratório de Microscopia Eletrônica" from Universidade do Estado do Rio de Janeiro; Stereoscan 200, Cambridge Instruments, Inc. at ANSP; Zeiss LEO 440 at Labo

ratório de Microscopia Eletrônica of MZSP; Zeiss LEO 940A at Laboratório de Tribologia e Materiais of the Universidade Federal de Uberlândia; and Zeiss DSM 960 at Departamento de Ciências dos Materiais e Metalurgia of the Pontifícia Universidade Católica do Rio de Janeiro.

In the "Material examined" lists, numbers betweens square brackets refer to the number of shells.

Abbreviations used: --Collections: ANSP - Academy of Natural Sciences of Philadelphia, Philadelphia; BMNH - British Museum Natural History, London; DF - Private collection Daniel Forcelli; IBUFRJ - Instituto de Biologia / Universidade Federal do Rio de Janeiro, Rio de Janeiro; MACN - Museu Argentino de Ciencias Naturales, Buenos Aires; MNHN - Muséum National d'Histoire Naturelle, Paris; MNRJ – Museu Nacional / Universidade Federal do Rio de Janeiro, Rio de Janeiro; MORG - Museu Oceanográfico "Eliézer de Carvalho Rios", Rio Grande; MZSP - Museu de Zoologia da Universidade de São Paulo, São Paulo; MMUFRPE - Museu de Malacologia da Universidade Federal Rural de Pernambuco, Recife; USNM -National Museu of Natural History, Washington, DC; ZMA -Zoölogish Museum Amsterdam, Amsterdam; -- Expeditions: AMASSEDS - A Multidisciplinary Amazon Shelf Sedimentary Study; CFVII - Comissão Oceanográfica Cabo Frio VII; GEO-MAR XII - Comissão Oceanográfica Geologia Marinha XII; PADCT Programa de Apoio ao Desenvolvimento Científico e Tecnológico; REVIZEE - Recursos Vivos da Zona Econômica Exclusiva; --Collectors: NOAG - Navio Oceanográfgico Astro Garoupa / Petrobras S.A.; AS - Navio Oceanográfico Atlântico Sul / Brazilian Navy; Eq.Zoo. - Equipe do Departamento de Zoologia / IBUFRJ; NOAN - Navio Oceanográfico Antares /



Brazilian Navy; NOAS - Navio Oceanográfico Almirante Saldanha / Brazilian Navy; NOWB - Navio Oceanográfico Professor W. Besnard / Universidade de São Paulo; RVCI - Research Vessel Columbus Iseling, University of Miami.

Systematic

Genus Turbonilla Risso, 1826

Turbonilla Risso, 1826: 224. Type species: Turbonilla costulata Risso, 1826: 224, fossil, Pliocene, France. Subsequent designation by Hermannsen (1852).

Remarks

The genus *Turbonilla* is taken herein in an wide concept, as used by Aartsen (1981), Schander (1994), Peñas *et al.* (1996), and Peñas & Rolán (1997). In view of the confused and poorly established subgeneric classification, with many artificial and

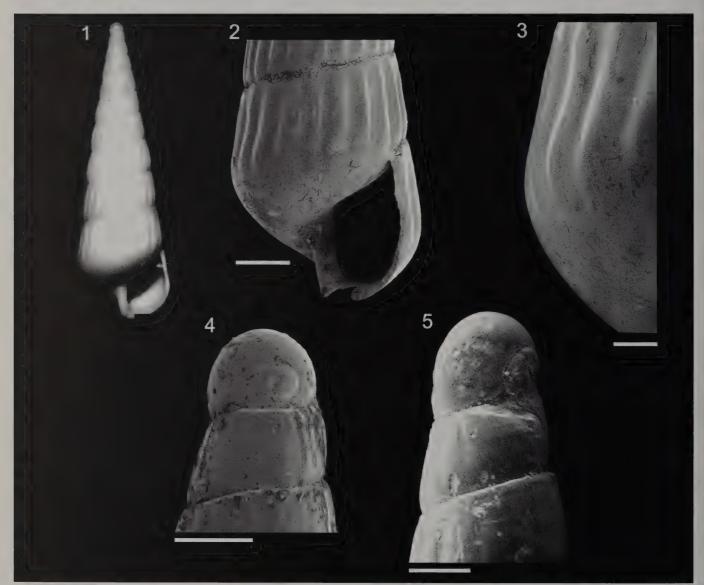
poorly defined subgenera in need of major revision (SCHANDER, 1994), we adopted no subgeneric classification.

Turbonilla farinatiae n. sp. (figs 1-5)

Turbonilla (Turbonilla) atypha Bush, 1899: FARINATI (1993: 299, fig. 2).

Description

Shell tall, strictly conical; color milk-white. Teleoconch whorls almost flat-sided, slightly concave on middle of latter whorls. Suture shallow and straight. Protoconch heterostrophic planispiral; diameter about 320 µm. Axial ribs low, broad, straight and orthocline or slightly opisthocline, fairly distinct on initial 2-4 whorls; 22 ribs on body whorl of holotype; interspaces narrow, about half as wide as the ribs, bearing microscop-



Figs 1-5. Turbonilla farinatiae n. sp. 1-4: holotype (MNRJ 8931); 5: paratype (ZMA 4.02.019). Fig. 1: whole shell (length 6.4 mm); fig. 2: last whorl; fig. 3: detail of periphery of base; figs 4, 5: protoconch. Scale bars: 200 µm.

Figg. 1-5. Turbonilla farinatiae n. sp. 1-4: olotipo (MNRJ 8931); 5: paratipo (ZMA 4.02.019). Fig. 1: conchiglia intera (lunghezza 6,4 mm); fig. 2: ultimo giro; fig. 3: dettaglio della periferia della base; figure 4, 5: protoconca. Scala di riferimento: 200 μm.



ic axial growth lines, especially visible below the sutures. Spiral sculpture absent, except for microscopic striations. Base rounded with evanescent axial ribs that project a little over it, but do not reach the umbilical region. Aperture rhomboid. Columella straight, without fold. Outer lip thin. No umbilical fissure.

Dimensions

Holotype with 9.25 teleoconch whorls; height 6.4 mm; width 2.0 mm

Type material

Holotype: MNRJ 8931, off Cabo Frio, Rio de Janeiro State, CFVII # 6147 (22°53.7'S, 041°50.5'W, 50 m), 24/iii/1983,

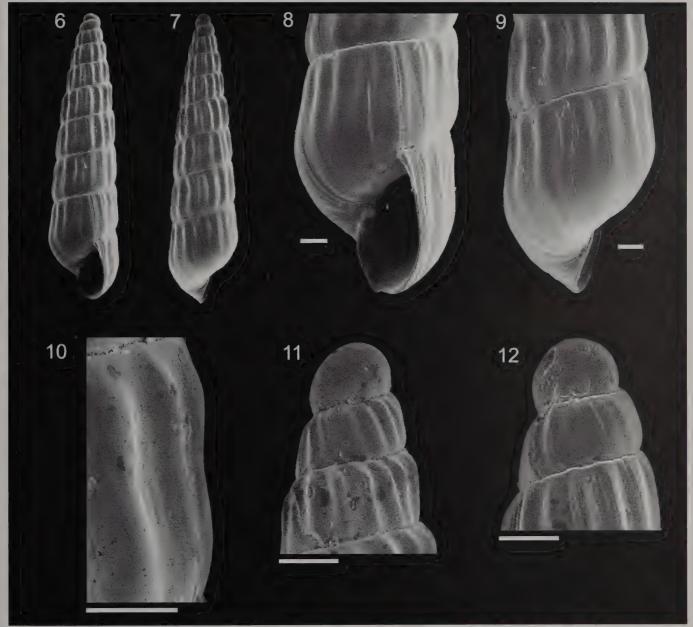
NOAS coll. Paratypes: MORG 41037, type locality; IBUFRJ 11898; ANSP 410339, Monte Alto, Espírito Santo State, 28/i/1986; MNRJ 8930; MNHN; ZMA 4.02.019, off Rio de Janeiro State CFVII # 6165 (23°02.8'S, 042°46'W, 56 m), 23/iii/1983, NOAS coll.; MZSP 35856, Arquipélago de Santana, Macaé, Rio de Janeiro State, v/1993, NOAG coll.

Type locality

Off Cabo Frio, north coast of Rio de Janeiro State (22°53.7'S, 041°50.5'W, 50 m), southeast coast of Brazil.

Additional material

--Espírito Santo State: IBUFRJ 8859, off Camburi (45 m),



Figs 6-12. Turbonilla goytacazi n. sp. holotype (MNRJ 8935). Figs 6, 7: whole shell (length 6.6 mm); figs 8, 9: last whorl; fig. 10: detail of periphery of whorl; figs 11, 12:protoconch. Scale bars: 200 μm.

Figg. 6-12. Turbonilla goytacazi n. sp. olotipo (MNRJ 8935). Figg. 6, 7: conchiglia intera (lunghezza 6,6 mm); figure 8, 9: ultimo giro; fig. 10: dettaglio della periferia di un giro; figg. 11, 12: protoconca. Scala di riferimento: 200 µm.



10/vi/1986, Eq.Zoo coll. [1]; IBUFRJ 8858, off Camburi (60 m), 15/i/1987, Eq.Zoo coll. [1]; --Rio de Janeiro State: IBUFRJ 6488, Arquipélago de Santana, Macaé, v/1993, NOAG coll. [3]; IBUFRJ 8853, off Cabo Frio, CFVII # 6147 (22°53.7'S, 041°50.5'W, 50 m), 24/iii/1983, NOAS coll. [7]; IBUFRJ 8854, off Cabo Frio, CFVII # 6199 (23°17'S, 044°15'W, 48 m), 02/iv/1983, NOAS coll. [1]; MORG 13576, off Rio de Janeiro State (40-50 m), "barco de pesca" coll. [1]; IBUFRJ 8855, CFVII # 6165 (23°02.8'S, 042°46'W, 56 m), 23/iii/1983, NOAS coll. [9].

Distribution

Southeast of Brazil (Espírito Santo and Rio de Janeiro States); Holocene of Bahia Blanca, Argentina.

Etymology

This species is named after Dr. Ester Farinati, who contributed a lot to the knowledge of pyramidellids from South America.

Remarks

FARINATI (1993) reported *T. atypha* Bush, 1899 from the Holocene of Bahia Blanca, Argentina. However, the illustration provided does not correspond to *T. atypha*, which is more slender and has more convex whorls and somewhat sinuous axial ribs. The figure in Farinatia (1993) indicates, rather, an exemplar of *Turbonilla farinatiae* with a slightly wider than normal shell.

The most similar species from the southwest Atlantic is *Turbonilla uruguayensis* Pilsbry, 1897, which has a very similar shell shape, but is larger and bears stronger axial ribs which end abruptly on the periphery of the last whorl. In *T. farinatiae* the ribs project slightly over the base (figs 2, 3). Moreover, both *T. uruguayensis* and *T. atypha* have helicoid protoconchs, while *T. farinatiae* has a planispiral one (figs 4, 5). *Turbonilla paucistriata* (Jeffreys, 1884: 361, pl. 27, fig. 6), described from the Mediterranean, has a very similar sculpture pattern and a planispiral protoconch, but it is less conical than *T. farinatiae* (fig. 1), having a somewhat pupoid shell shape. Moreover, the ribs of *T. paucistriata* are less marked and the inner lip is much more developed than in *T. farinatiae*.

Turbonilla goytacazi n. sp. (figs 6-12)

Description

Shell tall, moderately conical; color milk-white with yellow spiral band above the sutures. Teleoconch whorls almost straight in profile, but slightly concave on middle, slightly shouldered below the sutures, specially on initial whorls. Suture somewhat deep, straight. Protoconch heterostrophic planispiral; diameter about 330 µm. Axial ribs broad, low, straight and orthocline or prosocline with a spiral line of constriction on the middle; 17 on last whorl of holotype; interspaces narrow, about half as wide as the ribs, bearing microscopic axial growth lines. Spiral sculpture absent, except for very thin spiral striae. Base rounded, with evanescent axial ribs that project a little over the it, but do not reach the umbilical region. Aperture rhomboid, tending to pyriform. Columella obliquely straight, with obsolete fold. Outer lip thin. No umbilical fissure.

Dimensions

Holotype with 8 teleoconch whorls; height 6.6 mm; width 1.7 mm.

Type material

Holotype: MNRJ 8935, Bacia de Campos, Rio de Janeiro State (22°15'25"S, 040°19'39"W, 100 m), NOAG coll.; Paratypes: MNHN; ANSP 410340, off Rio de Janeiro State, REVIZEE # 38C (22°00'24"S, 040°05'13"W, 100 m), 08/ii/1997, NOAG coll.; MORG 41038, Bacia de Campos, Rio de Janeiro State (22°15'20"S, 040°19'45"W, 100 m), NOAG coll.; MNRJ 8936, Bacia de Campos, Rio de Janeiro State (22°15'07"S, 040°19'56"W, 95 m), NOAG coll.; IBUFRJ 11899, off Bacia de Campos, Rio de Janeiro State, NOAG coll.; MZSP 28877, 23°57.5'S, 044°53'W, 75 m, 27/vii/1986, NOWB coll.; ZMA 4.02.020, Bacia de Campos, Rio de Janeiro State (22°15'26"S, 040°19'41"W, 105 m), NOAG coll.

Type locality

Bacia de Campos, Rio de Janeiro State (22°15'25"S, 040°19'39"W, 100 m), southeast coast of Brazil.

Additional material

IBUFRJ 11003, off Espírito Santo State, REVIZEE # EO 532 (19°43.857'S, 039°26.653'W, 359 m), Navio Thalassa coll. [1]; MORG 38641, off Santa Catarina (28°44'S, 047°38'W, 200 m), ii/1987, AS coll. [1].

Distribution

Coast of Espírito Santo to north coast of Rio de Janeiro State and Santa Catarina State.

Etymology

This species is named after the Brazilian Indian tribe Goytacaz (Portuguese term to the Brazilian Indian word Waitacá, great runners) that inhabited the land area in Campos Bay.

Remarks

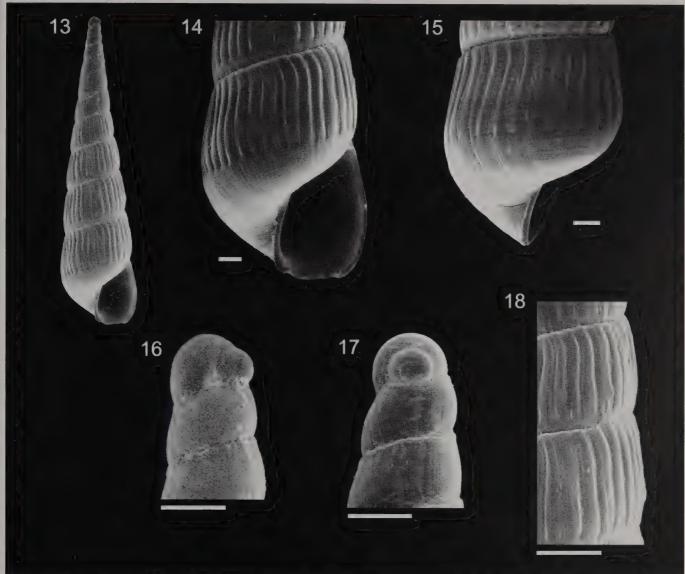
Turbonilla goytacazi (figs 6-12) resembles T. farinatiae (figs 1-5) in the shape of the axial ribs, the degree of their projection over the base (figs 8, 9), and protoconch type (figs 11, 12). The species can be distinguished by the general shape of the shell, which is less conical and narrower in T. goytacazi (figs 6, 7) and by the yellow spiral band, absent in T. farinatiae.

Turbonilla farroupilha n. sp. (figs 13-18)

Description

Shell tall, strictly conical, with acuminate apex; color milk-white. Teleoconch whorls slightly pyriform in profile, with accentuated convexity in the anterior half, above the suture. Suture somewhat deep, delicately sinuous by ribs projection. Protoconch heterostrophic helicoid; diameter about 260 µm. Axial ribs orthocline or opisthocline, sinuous, slender and very close each other; not distinct on initial 3-4 whorls; summits of the ribs projected over anterior suture; 40 ribs on body whorl of holotype; interspaces very narrow, about half as wide as the ribs, ending abrupt-





Figs 13-18. Turbonilla farroupilha n. sp. 13, 14, 17, 18: holotype (MORG 41049); 15: paratype (IBUFRJ 11900); 16: paratype (MNHN). Fig. 13: whole shell (length 7.8 mm); figs 14, 15: last whorl; figs 16, 17: protoconch; fig. 18: detail of sculpture. Scale bars: 250 µm.

Figg. 13-18. Turbonilla farroupilha n. sp. 13, 14, 17, 18: olotipo (MORG 41049); 15: paratipo (IBUFRJ 11900); 16: paratipo (MNHN). Fig. 13: conchiglia intera (lunghezza 7,8 mm); figg. 14, 15: ultimo giro; figure 16, 17: protoconca; fig. 18: dettaglio della scultura. Scala di riferimento: 250 µm.

ly at periphery of last whorl and some distance posterior to the summit of the succeeding whorl, thus leaving a rather broad, plain band above the suture. Spiral sculpture absent, except for microscopic spiral striations. Base rounded, smooth. Aperture rhomboid, well rounded anteriorly. Columella obliquely arcuate, without fold. Outer lip thin. No umbilical fissure.

Dimensions

Holotype with 10.5 teleoconch whorls; height 7.8 mm; width 1.8 mm

Type material

Holotype: MORG 41049 off Rio Grande, Rio Grande do Sul State (42 m), iii/1972, AS coll.; Paratypes: MNRJ 8919; ANSP 410341; MORG 41049; MNHN, type locality; IBUFRJ 11900; MZSP 35859 Armação, Santa Catarina State, Daniel Forcelli leg.

Type locality

Off Rio Grande (42 m), Rio Grande do Sul State, south coast of Brazil.

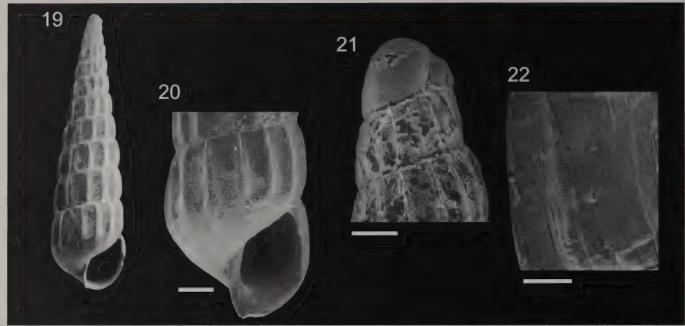
Additional material

--Rio de Janeiro State: IBUFRJ 9423, Arquipélago de Santana, Macaé, v/1993, NOAG coll. [3]; --Santa Catarina State: IBUFRJ 9399, Armação, D.Forcelli leg. [1]; IBUFRJ 9410, CFVII # 6165 (23°02.8'S, 042°46'W, 56 m), 23/iii/1983, NOAS coll. [2]; --Rio Grande do Sul State: MORG 21914, off Rio Grande (42 m), iii/1972, AS coll. [5]; MORG 27031, Parcel do Carpinteiro, 25/xi/1988, AS coll. [6].

Distribution

Southeast-souht coast of Brazil (Rio de Janeiro, Santa Catarina and Rio Grande do Sul States).





Figs 19-22. Turbonilla rhabdota (Watson, 1886). 19: (IBUFRJ 8899); 20-22 (IBUFRJ 5229); Fig. 19: whole shell (length: 5.6 mm); fig. 20: last whorl; fig. 21: protoconch; fig. 22: detail of whorl. Scale bars: fig. 20: 300 μm; figs 21, 22: 100 μm.

Figg. 19-22. Turbonilla rhabdota (Watson, 1886). 19: (IBUFRJ 8899); 20-22 (IBUFRJ 5229); Fig. 19: conchiglia intera (lunghezza: 5,6 mm); fig. 20: ultimo giro; fig. 21: protoconca; fig. 22: dettaglio di un giro. Scala di riferimento: fig. 20: 300 μm; figg. 21, 22: 100 μm.

Etymology

This species is named after the "Guerra dos Farrapos", a civil war that took place in Rio Grande do Sul State (1835-1845).

Remarks

Turbonilla farroupilha is well characterized by its acuminate apex (fig. 13), and the sigmoid ribs which are very closely packed (figs 14, 15) and faint in the earlier whorls (figs 13, 16, 17). The most similar species from the southwest Atlantic are Turbonilla uruguayensis Pilsbry, 1897 and Turbonilla atypha Bush, 1899. Both have a conical shell shape and lack spiral sculpture, and T. uruguayensis also bears faint ribs on the earlier whorls. However, in T. farroupilha, the earlier whorls narrow abruptly, giving rise to an acuminate apex (fig. 13), and the ribs are more sinuous, slender, and closely packed (figs 14, 15). Turbonilla turris (d'Orbigny, 1840), the holotype of which was illustrated by PIMENTA & ABSALÃO (2001), has the same kind of microscopic spiral striae and also a somewhat acuminate apex, but the ribs and the profile of the whorls are straight.

Turbonilla rhabdota (Watson, 1886) (figs 19-22)

Odostomia (Turbonilla) rhabdota Watson, 1886. Challenger Reports, Zoology 15: 491-492, pl. 32, fig. 4. Turbonilla rhabdota (Watson, 1886): Odé (1996: 56).

Types

Holotype: presumably at BMNH, not examined.

Type locality

Off Cuebra Island, West Indies (Chalenger St. 24: 18°38'30"N, 065°05'30"W, 700 m).

Material examined

--Pará State: IBUFRJ 8898, AMASSEDS # 4134 (off Pará), x/1991, RVCI coll. [1]; --Espírito Santo State: IBUFRJ 9009, off Camburi, 15/x/1997, Eq.Zoo. coll. [1]; IBUFRJ 5929, off Camburi #11A, 18/iii/1991, Eq.Zoo. coll. [2]; --Rio de Janeiro State: IBUFRJ 8899, Arquipélago de Santana, Macaé, v/1993, NOAG coll. [4]; --São Paulo State: MZSP 30908, Praia do Goes, Ilha de Santo Amaro, 04/i/21970, J.Vaz coll. [2]; MORG 38613, off São Vicente, vii/1974, J. Cololle coll. [1]; --Paraná State: MORG 15927, off Camboriu, viii/1971, E.Vokes & H.Vokes coll. [2]; MZSP28862, Praia de Caiobá, 15/xii/1948, J.P.Carvalho coll. [2].

Distribution

West Indies; north, southeast and south coasts of Brazil (Pará, Espírito Santo, Rio de Janeiro, São Paulo and Paraná States).

Remarks

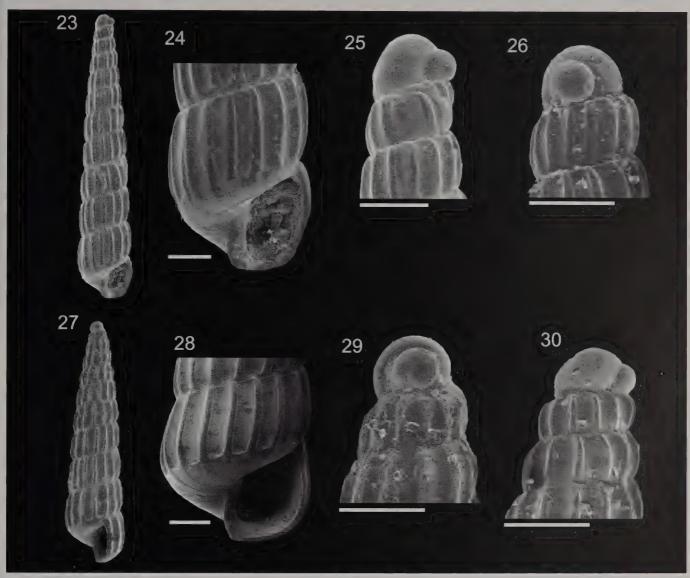
ODE (1996) listed *T. valida* Verrill & Bush, 1900 and *Turbonilla kymatoessa* (Watson, 1886) in synonymy with *T. rhabdota*. We do not agree with this statement. *Turbonilla valida* has more convex whorls, and its ribs are not straight and do not have the same characteristics of sharpness and spacing as *T. rhabdota*. *Turbonilla kymatoessa* has spiral striae in the interspaces and a planispiral protoconch (Watson, 1886).

Turbonilla lepta n sp. (figs 23-26)

Description

Shell small, moderately tall, very slender, elongate-conic to almost cylindrical in shape; fresh shells transparent, older ones white. Teleoconch whorls somewhat straight in profile. Suture





Figs. 23-26. Turbonilla lepta n. sp. 23-25: holotype (MNRJ 8939); 26: paratype (MNHN). Fig. 23: whole shell (length 4.6 mm); fig. 24: last whorl; figs 25, 26:protoconch. Scale bars: 200 µm.

Figs 27-30. Turbonilla parviscymna n. sp. 27-29: holotype (MNRJ 8922); 30: paratype (IBUFRJ 11902). Fig. 27: whole shell (length 3.8 mm); fig. 28: last whorl; figs 29, 30:protoconch. Scale bars: 200 µm.

Figg. 23-26. Turbonilla lepta n. sp. 23-25: olotipo (MNRJ 8939); 26: paratipo (MNHN). Fig. 23: conchiglia intera (lunghezza: 4,6 mm); fig. 24: ultimo giro; figg. 25, 26: protoconca. Scala di riferimento: 200 µm.

Figg. 27-30. Turbonilla parviscymna n. sp. 27-29: olotipo (MNRJ 8922); 30: paratipo (IBUFRJ 11902). Fig. 27: conchiglia intera (lunghezza 3,8 mm); fig. 28: ultimo giro; figg. 29, 30: protoconca. Scala di riferimento: 200 µm.

shallow, straight. Protoconch heterostrophic heliocoid; diameter about 230 µm. Axial ribs straight, slender orthocline; 16 on body whorl of holotype; interspaces about as wide as the ribs, ending abruptly at periphery of last whorl. Spiral sculpture absent. Base rounded, smooth. Aperture rhomboid. Columella straight, without fold. Outer lip thin. No umbilical fissure.

Dimensions

Holotype with 10.5 teleoconch whorls; height 4.6 mm; width 0.8 mm.

Type material

Holotype: MNRJ 8939, off Cabo Frio, Rio de Janeiro State, CFVII # 6147 (22°53.7'S, 041°50.5'W, 50 m), 24/iii/1983,

NOAS coll.; Paratypes: IBUFRJ 11901; MORG 41039; MZSP 35851, type locality; MNHN off Espírito Santo State; ANSP 410342, off north coast of Rio de Janeiro State, REVIZEE # D3 (22°52'S, 041°09'W, 80 m), 23/ii/1996, NOAN coll.; MNRJ 8940, Bacia de Campos, Rio de Janeiro State (22°15'36"S, 040°20'16"S, 100 m).

Type locality

Off Cabo Frio, north coast of Rio de Janeiro State (22°53.7'S, 041°50.5'W, 50 m), southeast coast of Brazil.

Additional material

--Espírito Santo State: IBUFRJ 8907, off Espírito Santo State [2]; -- Rio de Janeiro State: IBUFRJ 9440, off Cabo Frio, CFVII



6174 (23°16.8'S, 043°02.7'W, 92 m), 29/iii/1983, NOAS coll. [1]; IBUFRJ 9441, off Cabo Frio, CFVII # 6147 (22°53.7'S, 041°50.5'W, 50 m), 24/iii/1983, NOAS coll. [5]; IBUFRJ 10350, REVIZEE # D3 (22°52'S, 041°09'W, 80 m), 23/ii/1996, NOAN coll. [2].

Distribution

Southeast coast of Brazil (Espírito Santo and Rio de Janeiro States).

Etymology

This species is named after its slender, thin shell (*leptos*, Gr. = fine, small, thin, delicate).

Remarks

Turbonilla lepta (figs 23-26) is similar in dimensions and sculpture to *Turbonilla coomansi* Aartsen, 1993, with the same protoconch type and number of whorls (figs 25, 26). *Turbonilla lepta*, however, is more slender, tall, and has an almost cylindrical shape (fig. 23), while *T. coomansi* is more conical. Moreover, *T. coomansi* has somewhat curved and sharper axial ribs and convex whorls profile, while in *T. lepta*, the ribs and whorls profile are straight (fig. 23).

Turbonilla parviscymna n. sp. (figs 27-30)

Description

Shell small, conical; color white. Teleoconch whorls straight. Suture shallow, slightly sinuous by ribs projection. Protoconch heterostrophic helicoid; diameter about 200 µm. Axial ribs slender, straight, orthocline or slightly opisthocline; 17 on body whorl of holotype; interspaces about twice as wide as the ribs, ending abruptly at periphery of last whorl. Spiral sculpture absent. Base rounded, smooth. Aperture rhomboid. Columella straight, without fold. Outer lip thin. No umbilical fissure.

Dimensions

Holotype with 8.25 teleoconch whorls; height 3.8 mm; width 0.7 mm.

Type material

Holotype: MNRJ 8922, off Pará State, AMASSEDS # 3209 (01°20.93'N, 048°00.20'W, 53 m), RVCI coll.; Paratypes: ANSP 410343; MNHN; IBUFRJ 11902, type locality.

Type locality

Off Pará State, north coast of Brazil, AMASSEDS # 3209 (01°20.93'N, 048°00.20'W, 53 m).

Distribution

Known from type locality only.

Etymology

This species is named after its small protoconch in relation to shell width (parvus, L. = little; scymnus, L. = a young animal).

Remarks

Turbonilla parviscymna (figs 27-30) has the same sculpture and protoconch type (figs 29, 30) as *T. lepta* (figs 25, 26), but can be easily distinguished by its strictly conical shape (fig. 27), with a larger shell width in relation to the shell length, while *T. lepta* has an almost cylindrical shell shape, being very slender and tall (fig. 23).

Turbonilla fluminensis n. sp. (figs 31-34)

Description

Shell tall, slender, slightly conical; color white. Teleoconch whorls convex in profile. Suture somewhat deep, straight. Protoconch heterostrophic helicoid; diameter about 300 µm. Axial ribs slender, slightly sinuous, opisthocline; ending abruptly at periphery of last whorl; 18 on body whorl of holotype; interspaces as wide as the ribs. Spiral sculpture absent. Base rounded, smooth. Aperture rhomboid. Columella obliquely straight, without fold. Outer lip thin. No umbilical fissure.

Dimensions

Holotype with 10.25 teleoconch whorls; height 4.7 mm; width 0.9 mm.

Type material

Holotype: MNRJ 8943, off Macaé, Rio de Janeiro State, REVIZEE # D1 (22°48'S, 041°09'W, 69 m), 23/ii/1996, NOAN coll.; Paratypes: ZMA 4.02.021; MNRJ 8923; MZSP 35861, off north coast of Rio de Janeiro State, REVIZEE # D3 (22°52'S, 041°09'W, 80 m), 23/ii/1996, NOAN coll.; ANSP 410344, type locality; IBUFRJ 11903, Bacia de Campos, Rio de Janeiro State (22°47'47"S, 40°45'32"W), NOAG coll.; MNRJ 8924, Bacia de Campos, Rio de Janeiro State (22°15'20"S, 040°19'54"W, 100 m); MORG 41040, Bacia de Campos, Rio de Janeiro State (22°15'07"S, 040°19'56"W, 95 m); MNHN Espírito Santo I # 6446, 22/viii/1984, NOAS coll.

Type locality

Off Macaé (22°48'S, 041°09'W), Rio de Janeiro State, southeast coast of Brazil.

Additional material

-- Rio de Janeiro State: IBUFRJ 9443, off Cabo Frio, CFVII # 6147 (22°53,7'S, 041°50,5'W, 50 m), 24/iii/1983, NOAS coll. [1]; IBUFRJ 10349, REVIZEE # D3 (22°52'S, 041°09'W, 80 m), 23/ii/1996, NOAN coll. [1]; IBUFRJ 8906, off Bacia de Campos (22°15'25"S, 040°19'39" W, 100 m), NOAG coll. [1].

Distribution

Southeast coast of Brazil (off Espírito Santo and Rio de Janeiro States).

Etymology

"Fluminense" is the general determination to those people that was born in Rio de Janeiro State, Brazil.



Remarks

Turbonilla fluminensis (figs 31-34) is very similar in shell shape and sculpture to Turbonilla coomansi Aartsen, 1993, Turbonilla penistoni Bush, 1899, and Turbonilla enna Bartsch, 1927. However, T. fluminensis can be distinguished by its larger protoconch (mean diameter 300 μ m) which has about three whorls well projected in a helicoid type (figs 33, 34). The helicoid protoconchs of T. coomansi and T. penistoni bear about two whorls of medium size (mean diameters 210 and 200 μ m, respectively), and T. enna has a planispiral protoconch (figs 39, 40).

The whorls profile of *T. fluminensis* (figs 31, 32) is more regularly convex than *T. penistoni* (the holotype was illustrated by ABSALÃO & PIMENTA, 1999). The latter has a somewhat sinuous whorl profile, with the ribs projecting over the anterior suture.

Turbonilla aff. enna Bartsch, 1927 (figs.35-40)

Turbonilla enna Bartsch in Dall, 1927: 81-82; Odé (1996: 40).

Type locality

Off Fernandina, Florida (530 m).

Types

Holotype: USNM 360175; paratype: USNM 360176.

Material examined

The holotype and: --Rio de Janeiro State: IBUFRJ 9465, off Cabo Frio, CFVII # 6147 (22°53.7'S, 041°50.5'W, 50 m), 24/iii/1983, NOAS coll. [2]; IBUFRJ 9466, off Prainha, Arraila do Cabo, 1989, T.Almeida coll. [1]; IBUFRJ 10351, REVIZEE # D3 (22°52'S, 041°09'W, 80 m), 23/ii/1996, NOAN coll. [10]; IBUFRJ 7202, CFVII # 6194 (24°03.6'S, 044°07.6'W, 134 m), 01/iv/1983, NOAS coll. [2]; IBUFRJ 9463, off Bacia de Campos (22°15'20"S, 040°19'45"W, 100 m), NOAG coll. [4]; IBUFRJ 9464, Bacia de Campos (22°15'26"S, 040°19'41"W, 105 m), NOAG coll. [1].

Distribution

Fernandina (Florida); southeast coast of Brazil (off Rio de Janeiro State).

Remarks

The ribs of the Brazilian specimens determined herein as *Turbonilla* aff. *enna* Bartsch, 1927 (figs 36-40) are stronger and less straight than in the holotype (fig. 35). Our determination of these specimens as *T.* aff. *enna* is based on the strong similarity in the protoconch type and general shell shape. Both the holotype and the Brazilian specimens have very similar dimensions and convex whorls. The most striking similarity, however, lies in the protoconch, which is planispiral (figs 35, 39, 40) in contrast to all other species without spiral sculpture discussed herein (*T. lepta, T. parviscymna, T. penistoni, T. fluminensis, T. coomansi, and T. unilirata* Bush, 1899).

Turbonilla zulmae Pimenta & Absalão, 1998 (figs 41-44)

Turbonilla zulmae Pimenta & Absalão, 1998: 63 nom. nov. for

Turbonilla elongata Castellanos, 1982 non Pease, 1967. Turbonilla elongata Castellanos, 1982: 66-67, fig. 8; FARINATI (1993: 301, fig. 5).

Type locality

Pto. Quequén (40°25'S, 61°34'W, 30 m), Argentina.

Types

Holotype not located (probably lost). Paratypes: MACN 30807, Pto. Quequén, Est. Hidrobiologica, vii/1937, Carcelles, Parodiz coll. [13]; MACN 34015, Est. 16 del Shinkai Marú (38°31'S, 057°25'W, 50 m)[1].

Material examined

--Rio de Janeiro State: IBUFRJ 11896, off Cabiúnas, 21/iv/1993, NOAG coll. [1]; --Rio Grande do Sul State: MORG 17858, off Chuí, # 405 (65 m), 29/x/1968, NOWB coll. [4]; MZSP19301, off Chuí (34°32'S, 052°27'W, 65 m), 19/x/1968, NOWB coll. [5];

Remarks

Turbonilla zulmae Pimenta & Absalão, 1998 was proposed as a new name for Turbonilla elongata Castellanos, 1982 non Pease, 1967. Although the holotype of this species is probably lost, we were able to examine its paratypes, on which we base our discussion (fig. 41). This species is very similar to Turbonilla atypha Bush, 1899, but is more slender, with more marked and numerous axial ribs.

The specimens from Brazil (figs 42-44) have very similar general shell shape and axial ribs, but are less slender and have a more convex whorls outline than the illustrated paratype of *T. zulmae* (fig. 41).

Turbonilla aff. unilirata Bush, 1899 (figs 45-48)

Turbonilla unilirata Bush, 1899: 165, pl. 8, fig. 6; Warmke & Abbott (1962: 149); Jong & Coomans (1988: 127, pl. 20, figs. 660); Johnson (1989: 72); Odé (1996: 60); Absalão & Pimenta (1999: 83, fig. 11).

Turbonilla (Pyrgiscus) unilirata Bush, 1899: VOKES & VOKES (1983: 33, pl. 31, fig. 8).

Turbonilla cf. unilirata Bush, 1889: DIAZ & PUYANA (1994: 239, fig. 957).

Type locality

Saint Thomas, West Indies.

Types

Holotype by original designation: ANSP 79010; paratypes: ANSP 79010 (three specimens, missing).

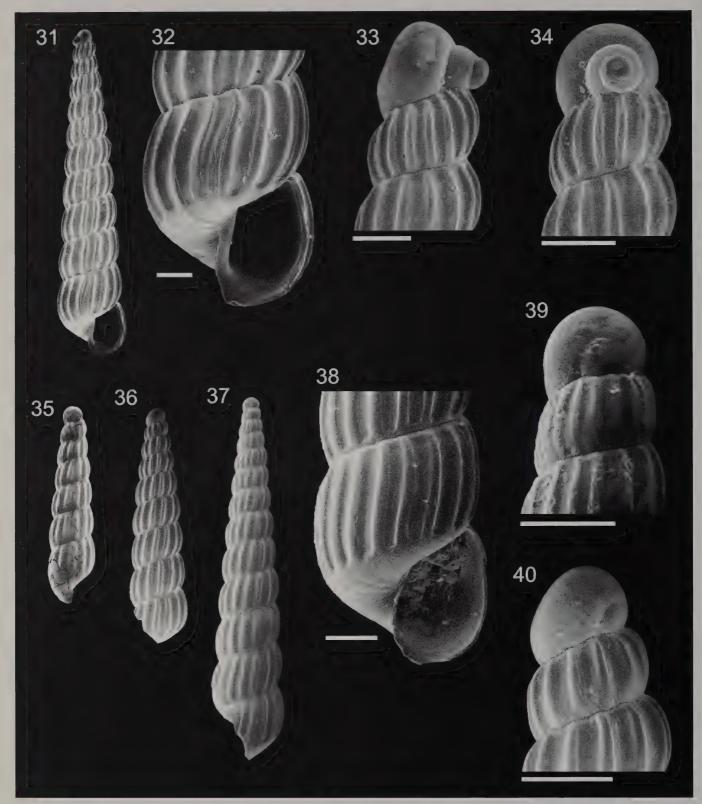
Material examined

The holotype and: MZSP 30907, off Rio de Janeiro State, PAD-CT 6627 (23°57.990'S, 043°52.560'W, 133 m) [5].

Distribution

West Indies; southeast coast of Brazil (off Rio de Janeiro State).





Figs 31-34. Turbonilla fluminensis n. sp. 31, 32: holotype (MNRJ 8943); 33, 34: paratype (MORG 41040). Fig. 31: whole shell (length 4.7 mm); fig. 32: last whorl; figs 33, 34: protoconch. Scale bars: 200 µm.

Figs 35-40. *Turbonilla* aff. *enna* Bartsch, 1927. 35: holotype (USNM 360175); 36-40 (IBUFRJ 10351). Figs 35-37: whole shells (lengths: 35: 4.2 mm, 36: 2.8 mm, 37: 5.0 mm); fig. 38: last whorl; figs 39, 40: protoconch. Scale bars: 200 µm.

Figs. 31-34. Turbonilla fluminensis n. sp. 31, 32: olotipo (MNRJ 8943); 33, 34: paratipo (MORG 41040). Fig. 31: conchiglia intera (lunghezza 4,7 mm); fig. 32: ultimo giro; figure 33, 34: protoconca. Scala di riferimento: 200 µm.

Figg. 35-40. Turbonilla aff. enna Bartsch, 1927. 35: olotipo (USNM 360175); 36-40 (IBUFRJ 10351). Figg. 35-37: conchiglie intere (lunghezza: 35: 4,2 mm, 36: 2,8 mm, 37: 5,0 mm); fig. 38: ultimo giro; figure 39, 40: protoconca. Scala di riferimento: 200 μm.



Remarks

For description, see BUSH (1899) and ABSALÃO & PIMENTA (1999). Turbonilla unilirata lacks spiral sculpturing, but can be clearly distinguished by a single spiral cord just below the suture, as seen in the holotype (fig. 45). As discussed by ABSALÃO & PIMENTA (1999: 83, fig. 12), a possible intraspecific variation of T. unilirata is characterized by shells with a more tapered outline and fewer axial ribs, with larger interspaces. The Brazilian specimens studied herein (figs 46-48) resemble this variation more closely, with the same tapered whorls and ribs pattern (fig. 46). However, the spiral cord (figs 46, 47) is not as marked as in the holotype of T. unilirata (fig. 45) and in its variation (ABSALÃO & PIMENTA, 1999: fig, 12), which led us to determine these specimens as T. aff. unilirata.

Turbonilla krebsii (Mörch, 1875) (figs 49-54)

Chemnitzia (Elusa) krebsii Mörch, 1875: 159-160.

Turbonilla krebsii (Mörch, 1875): Jong & Coomans (1988: 128, pl. 20, fig. 668); Odé (1996: 47); Redfern (2001: 150, pl. 67, figs 626A, 626B, 626C).

Turbonilla palmerae Aguayo & Jaume, 1936. synonymized by Jong & Coomans (1988)

Type locality

Saint Thomas.

Types

Two syntypes: ANSP 19983.

Material examined

The types above and: Grand Bahama Islands: ANSP 369280, Southwest Corner, Sweetings Cay, (26°36'45"N, 077°54'30"W), J.Worsfold coll. [2]; ANSP 370037, Fleming Road, Mosquito Point, (26°37'30"N, 078°54'W), J.Worsfold coll. [2]; ANSP 371818, 4 miles of Burmah Oil, (26°40'N, 078°09'W), J.Worsfold coll. [47]; --Bahia State: IBUFRJ 8860, off Bahia State, L.Trinchão coll. [1]; MNHN, Rio Vermelho, São Salvador, 1989, P.Maestrati coll. [2].

Distribution

Saint Thomas; Bahama Islands; northeast coast of Brazil (off Bahia State).

Remarks

Turbonilla krebsii (Mörch, 1875) is characterized by its pupoid shell (figs 49, 50), golden brown shell color, and spiral sculpture of about six rectangular pits in the interspaces (figs 51, 52). In contrast to these characters, which show very little variation, the axial ribs may be very faint in the last whorl, as in the holotype (fig. 49).

Turbonilla pupoides (d'Orbigny, 1842) (figs 55-60)

Chemnitzia pupoides d'Orbigny, 1842: 224, 225. Atlas (1853): pl. 16, fig 32, 36.

Turbonilla pupoides var. ischna Bush, 1899: 153-154, pl. 8, fig. 5; ABSALÃO & PIMENTA (1989: 82, fig. 21).

Pyrgostelis (Mormula) pupoides var. ischna (Bush, 1899): VERRILL & BUSH (1990: 351, pl. 65, figs. 21, 22).

Turbonilla (Pyrgiscus) pupoides (d'Orbigny, 1842): Vokes & Vokes (1983: 33, pl. 31, fig. 13).

Turbonilla pupoides ischna Bush, 1899: Jonhson (1898: 44).

Turbonilla pupoides (d'Orbigny, 1842): WARMKE & ABBOTT (1962: 148, pl. 26, fig. d); JONG & COOMANS (1988: 130, pl. 20, fig. 676); DIAZ & PUYANA (1994: 238, fig. 950); ODÉ (1996: 54); REDFERN (2001: 148, pl. 66, figs 616A, 616B).

Chemnitzia flavocincta C.B.Adams (1850). synonymyzed by Jong & Coomans (1988).

Type Locality

Cuba.

Types

Syntypes: BMNH 1854.10.4.148.

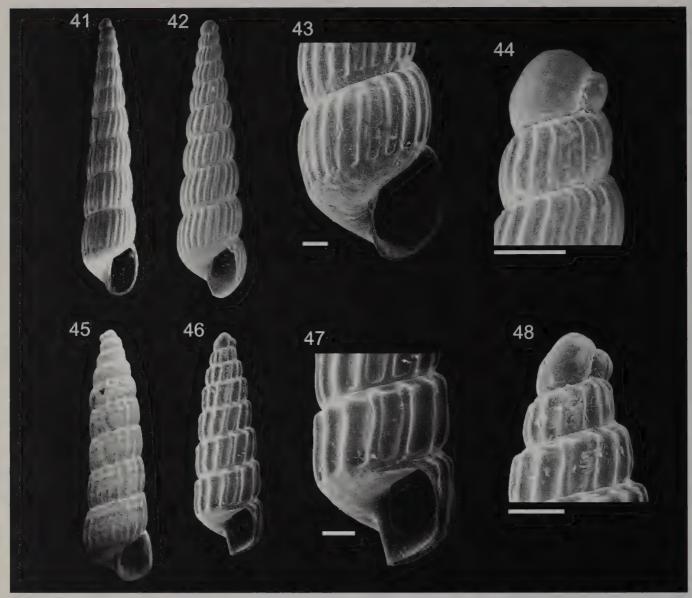
Material Examined

The types and: holotype of Turbonilla pupoides var. ischna Bush, 1899: ANSP 79014, no locality data; three paratypes of Turbonilla pupoides var. ischna Bush, 1899: ANSP 372507, no locality data; -- Grand Bahama Island: ANSP 371277, Dead Man's Reef [20]; --Bermuda: MORG 17506, off Bermuda [3]; --Bahia State: MORG 23909, Abrolhos bank, i/1985, Eq.MORG coll. [1]; MZSP28864, Abrolhos bank, v/1958, L.Pini Neto coll. [2]; MORG 23908, Abrolhos bank, i/1985, Eq. MORG coll. [90]; MORG 16401, off Itapuã, 17/vii/1967, E. Rios coll. [14]; IBUFRJ 11068, off Guarajuba, Itapuã, 1989, L.Trinchão coll. [4]; -- Espírito Santo State: IBUFRJ 8864, REVIZEE # VV38 (19°28'S, 038°22'W), 29/ii/1996, NOAN coll. [5]; IBUFRJ 7542, GEOMAR XII # 21 (20°49'80"S, 040°16'20"W, 37 m), 26/viii/1979, NOAC coll. [2]; IBUFRJ 7541, GEOMAR XII # 34 (21°15'30"S, 040°20'40"W, 46 m), 27/viii/1979, NOAC coll. [1]; IBUFRJ 8627, off Piúma, 1993 [5]; IBUFRJ 8865, off Espírito Santo State, 23/vi/1993 [2]; IBUFRJ 9779, REVIZEE # VV16 (21°10'S, 040°27'W, 27.65 m), 26/ii/1996, NOAN coll. [1]; IBUFRJ 10219, REVIZEE # C62 (20°30'02"S, 037°28'51"W, 96 m), 25/iv/1996, NOAN coll. [1]; IBUFRJ 10316, off Aracruz, 01/viii/1987 [1]; IBUFRJ 8862, off Espírito Santo State [3]; -- Rio de Janeiro State: IBUFRJ 7548, GEOMAR XII # 41 (21°21'S, 040°53'W, 12 m), viii/1979, NOAC coll. [2]; IBUFRJ 7543, GEOMAR XII # 76 (21°57'60"S, 040°51'W, 15 m), 28/viii/1979, NOAC coll. [1]; IBUFRJ 8863, REVIZEE # D1 (22°48'S, 041°09'W), 23/ii/1996, NOAN coll. [1]; IBUFRJ 7641, GEOMAR XII # 97 (22°07'50"S, 040°20'50"W, 67 m), 29/viii/1979, NOAC coll. [3].

Distribution

Cuba; Bahama Island; Yucatan Peninsula, Mexico; Colombian Caribbean; northeast and southeast coasts of Brazil (Bahia, Espírito Santo and Rio de Janeiro States).





Figs 41-44. Turbonilla zulmae Pimenta & Absalão, 1998. 41: paratype (MACN 34051); 42, 44 (MZSP 19301); 43 (IBUFRJ 7216). Figs 41, 42: whole shells (lengths: 41: 4.2 mm; 42: 4.0 mm); fig. 43: last whorl; fig. 44: protoconch. Scale bars: 200 μm.

Figs 45-48. Turbonilla aff. unilirata Bush, 1899. 45: holotype of T. unilirata (ANSP 79010); 46-48 (MZSP 30907). Figg. 45, 46: whole shells (lengths: 45: 2.8 mm, 46: 2.8 mm); 47: last whorl; 48: protoconch. Scale bars: 200 μm.

Figg. 41-44. Turbonilla zulmae Pimenta & Absalão, 1998. 41: paratipo (MACN 34051); 42, 44 (MZSP 19301); 43 (IBUFRJ 7216). Figg. 41, 42: conchiglie intere (lunghezza: 41: 4,2 mm; 42: 4,0 mm); fig. 43: ultimo giro; fig. 44: protoconca. Scala di riferimento: 200 μm.

Figg. 45-48. Turbonilla aff. unilirata Bush, 1899. 45: olotipo di T. unilirata (ANSP 79010); 46-48 (MZSP 30907). Figg. 45, 46: conchiglie intere (lunghezze: 45: 2,8 mm, 46: 2,8 mm); 47: ultimo giro; 48: protoconca. Scala di riferimento: 200 µm.

Remarks

Turbonilla pupoides (figs 55-60) is easily distinguished from the other *Turbonilla* species in the western Atlantic by its pupoid shell (figs 55-57) and spiral sculpture (fig. 58), with two wider striae, one in the middle of the whorl and another above the suture, and many fine, crowded spirals in the interspaces.

ABSALÃO & PIMENTA (1999) considered *Turbonilla pupoides* var. ischna Bush, 1899 as a synonym of *Turbonilla pupoides*. Based on the abundant material of this species studied herein, we could verify that the shell described by BUSH (1899) is part of an intraspecific variation of *T. pupoides*, especially the shape of the shell, which may be somewhat slender in some specimens (fig. 57).

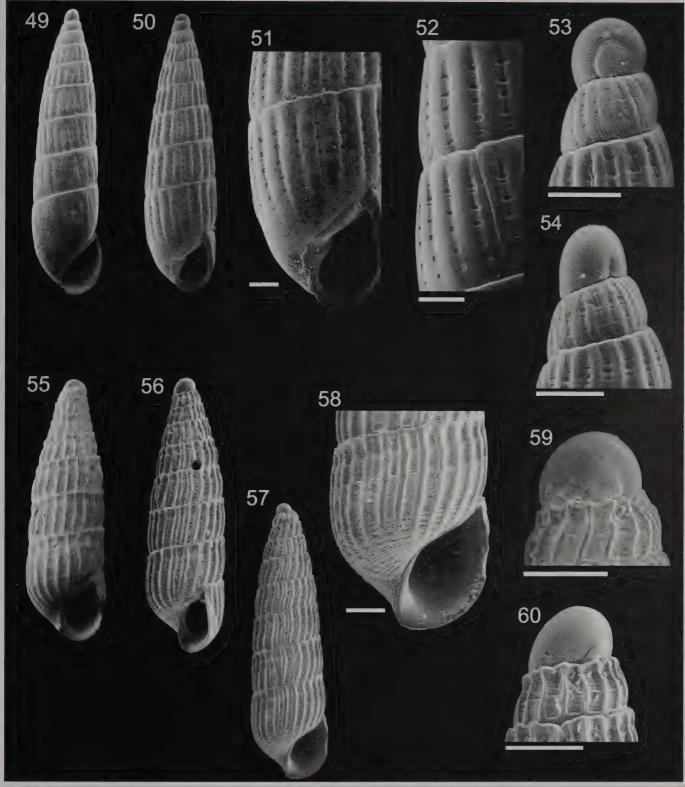
Turbonilla pupoides is very common along the Brazilian coast, but does not reach localities below 230 S. It is especially well represented off the coast of Bahia State, but is also present off the states of Rio de Janeiro and Espírito Santo. There are many other records from several localities in the Caribbean region (Bush, 1899; Verrill & Bush, 1900; Vokes & Vokes, 1983; Warmke & Abbott, 1964; Diaz & Puyana, 1994).

Turbonilla aracruzensis n. sp. (figs 61-65)

Description

Shell moderately tall, slender, conical; color light brown. Teleo-





Figs 49-54. *Turbonilla krebsii* (Mörch, 1875). 49: syntype (ANSP 19983); 50, 51 (IBUFRJ 8860); 52-54 (MNHN). Figs 49, 50: whole shells (lengths: 49: 4.1 mm, 50: 3.8 mm); fig. 51: last whorl; fig. 52: detail of sculpture; figs 53, 54: protoconch. Scale bars: 200 μm.

Figs 55-60. *Turbonilla pupoides* (d'Orbigny, 1842). 55: syntype (BMNH 1854.10.4.148). 56-60: (MORG 23908). Figs 55-57: whole shells (lengths: 55: 2.5 mm, 56: 3.0 mm, 57: 3.2 mm); fig. 58: last whorl, figs 59, 60: protoconch. Scale bars: 200µm.

Figg. 49-54. Turbonilla krebsii (Mörch, 1875). 49: sintipo (ANSP 19983); 50, 51 (IBUFRJ 8860); 52-54 (MNHN). Figure 49, 50: conchiglia intera (lunghezza: 49: 4,1 mm, 50: 3,8 mm); fig. 51: ultimo giro; fig. 52: dettaglio della scultura; figg. 53, 54: protoconca. Scala di riferimento: 200 μm.

Figg. 55-60. *Turbonilla pupoides* (d'Orbigny, 1842). 55: sintipo (BMNH 1854.10.4.148). 56-60: (MORG 23908). Figg. 55-57: conchiglie intere (lunghezze: 55: 2,5 mm, 56: 3,0 mm, 57: 3,2 mm); fig. 58: ultimo giro, figure 59, 60: protoconca. Scala di riferimento: 200 μm.



conch whorls slightly convex in profile. Suture shallow and straight. Protoconch heterostrophic planispiral; diameter about 360 µm. Axial ribs low, straight, orthocline or slightly opisthocline, becoming evanescent before reaching anterior suture; fairly distinct on first whorl; 20 on body whorl of holotype; interspaces narrow, heath the width of the ribs. Spiral sculpture formed by about 20 rows of very thin striae irregularly spaced. Base elongate, with evanescent ribs and very thin spirals. Aperture pyriform. Columella somewhat arcuate, without fold. Outer lip thin. No umbilical fissure.

Dimensions

Holotype with 8.5 teleoconch whorls; height 5.4 mm; width 1.2 mm.

Type material

Holotype: MNRJ 8928, Aracruz, Espírito Santo State, viii/1987; Paratypes: ANSP 410346, off Espírito Santo State, REVIZEE # VV16 (21°10'S, 040°27'W, 27.65 m), 26/ii/1996, NOAN coll.; IBUFRJ 11904, off Macaé, Rio de Janeiro State, REVIZEE # D1 (22°48'S, 041°09'W, 69 m), 23/ii/1996, NOAN coll.; MNHN; ZMA 4.02.022, Cabiúnas, Espírito Santo State, 20/iv/1993, NOAG coll.; MNRJ 8921, Camburi, Espírito Santo State, 12/ii/1987, Eq.Zoo. coll.; MORG 41041, Camburi, Espírito Santo State, 30/x/1986, Eq.Zoo. coll.; MNRJ 8929, type locality; MZSP 35850, Arquipélago de Santana, Macaé, Rio de Janeiro State, v/1993, NOAG coll.

Type locality

Off Aracruz, Espírito Santo State, southeast coast of Brazil.

Additional material

-- Espírito Santo State: IBUFRJ 10315 off Aracruz, viii/1987, V.Abud coll. [1]; IBUFRJ 8934 Camburi, 14/iii/1986, Eq.Zoo. coll. [1]; IBUFRJ 10298 off Cabiúnas, 20/iv/1993, NOAG coll. [5]; IBUFRJ 10307 off Cabiúnas, 21/iv/1993, NOAG coll. [3]; --Rio de Janeiro State: MORG 27884, off Cabo Frio, 2/xii/1986, NOAS coll. [1]; IBUFRJ 8931, Arquipélago de Santana, Macaé, v/1993, NOAG coll. [1]; --Rio Grande do Sul State: MORG 38603, off Rio Grande, (30-45 m), 1983, NOAS coll. [1]; --Uruguay: MORG 20068, off La Paloma (40 m), ii/1978, L.Alvarez coll. [1].

Distribution

Southeast and south coasts of Brazil (Espírito Santo, Rio de Janeiro and Rio Grande do Sul States); coast of Uruguay.

Etymology

The species is named after the city of Aracruz, type locality.

Remarks

Turbonilla aracruzensis (figs 61-65) shares with *T. krebsii* (figs 49-54), *T. pupoides* (figs 55-60), and *T. midas* n. sp. (see next species description) the same light brown, lustrous shell surface, but is distinguished by its shape, with a somewhat acuminate apex (fig. 61) and semi-pyriform whorls profile (figs 61-63).

Turbonilla midas n. sp. (figs 66-71)

Description

Shell tall, slender and conical; color light brown. Teleoconch whorls strictly straight in profile. Suture shallow, straight. Protoconch heterostrophic planispiral; diameter about 300 µm. Axial ribs low, straight, orthocline or slightly opisthocline; evanescent on anterior whorls; 26 on body whorl of holotype; interspaces about as wide as the ribs, with microscopic axial striae. Spiral sculpture formed by about 17 thin striae that cross the ribs on anterior whorls. Base elongate with evanescent axial ribs and very thin spiral striae. Aperture pyriform. Columella obliquely straight, with obsolete fold. Outer lip thin. No umbilical fissure.

Dimensions.

Holotype with 12 teleoconch whorls; height 8.5 mm; width 1.5 mm.

Type material. Holotype: MORG 41042 off Santos, São Paulo State (45 m), 24/ix/1969, NOAS coll.; Paratypes: MNRJ 8925; MNHN type locality; ANSP 410345; MORG 41043, off Rio Grande, Rio Grande do Sul State, (30-45 m), 1983, AS coll.; IBUFRJ 11905, Prainha, Arraial do Cabo, Rio de Janeiro State, 1989, T.Almeida coll.

Type locality

Off Santos (45 m), São Paulo State, southeast coast of Brazil.

Additional material

-- São Paulo State: MORG 15100, off Santos (45 m), 24/ix/1969, NOAS coll. [1]; --Rio de Janeiro State: IBUFRJ 6783 off Prainha, Arraial do Cabo, 1989, T. Almeida coll. [2]; IBUFRJ 9424 Arquipélago de Santana, Macaé, v/1993, NOAG coll. [1]; -- Rio Grande do Sul state: MORG 38570 off Rio Grande, # 4 (50 m), x/1983, NOAS coll. [1].

Distribution

Southeast and south coasts of Brazil (Rio de Janeiro, São Paulo and Rio Grande do Sul States).

Etymology

The species is named after Midas, king of Frígia, whose touch, according to the legend, changed anything into gold. An allusion to the somewhat golden shell surface.

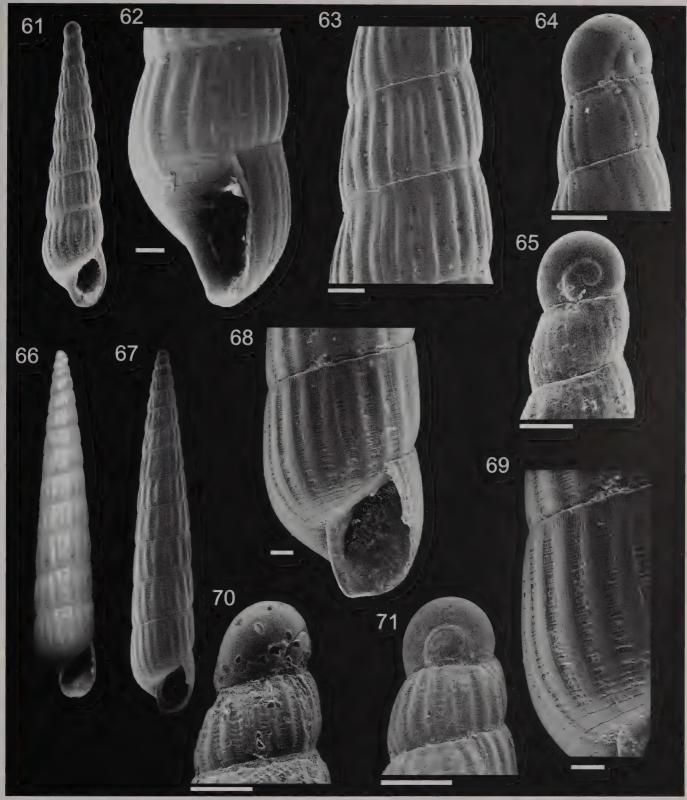
Remarks

Turbonilla midas (figs 66-71) has the same shell surface color as *T. aracruzensis* (figs 61-65) but with the addition of a light golden tone. It can also be distinguished by its shell shape, with a strictly straight whorls profile (figs 66-68); in *T. aracruzensis* the whorls profile is slightly convex. The spiral sculpturing (figs 68, 69) is stronger and numerous in *T. midas*.

Turbonilla portoricana Dall & Simpson, 1901 (figs 72-78)

Turbonilla portoricana Dall & Simpson, 1911: 414-415, pl. 53,





Figs 61-65. Turbonilla aracruzensis n. sp. 61-64: holotype (MNRJ 8928); 65: paratype (IBUFRJ 11904). Fig. 61: whole shell (length 5.4 mm); fig. 62: last whorl; fig. 63: detail of sculpture on last whorl; figs 64, 65: protoconch. Scale bars: 200 $\mu m.$

Figs 66-71. Turbonilla midas n. sp. 66, 71: paratype (MNHN); 65, 67-70: holotype (MORG 41042). Figs 66, 67: whole shell (lengths: 66: 8.2 mm, 67; 8.5 mm); fig.

68: last whorl; fig. 69: detail of sculpture on last whorl; figs 70, 71: protoconch. Scale bars: 200 μm.

Figg. 61-65. Turbonilla aracruzensis n. sp. 61-64: olotipo (MNRJ 8928); 65: paratipo (IBUFRJ 11904). Fig. 61: conchiglia intera (lunghezza 5,4 mm); fig. 62: ultimo giro; fig. 63: dettaglio della scultura dell'ultimo giro; figg. 64, 65: protoconca. Scala di riferimento: 200 µm.

Figg. 66-71. Turbonilla midas n. sp. 66, 71: paratipo (MNHN); 65, 67-70: olotipo (MORG 41042). Figg. 66, 67: conchiglia intera (lunghezza: 66: 8,2 mm, 67: 8,5 mm); fig. 68: ultimo giro; fig. 69: dettaglio della scultura dell'ultimo giro; figure 70, 71: protoconca. Scala di riferimento: 200 µm.



fig. 15; Warmke & Abbott (1962: 149, pl 29, fig. i); Jong & Coomans (1988: 128); Diaz & Puyana (1994: 238, fig. 952); Odé (1996: 52-53).

Type locality

Mayaguez, Porto Rico.

Types

Holotype: USNM 160204.

Material examined

The type and: --Espírito Santo State: IBUFRJ 8840, off Camburi, # 6 (60 m), 30/ix/1986, Eq.Zoo coll. [8]; IBUFRJ 8841, off Camburi (45 m), 15/i/1987, Eq.Zoo.coll. [1]; IBUFRJ 8842, off Camburi (60 m), 18/xi/19/86, Eq.Zoo.coll. [11]; IBUFRJ 8843, off Camburi (60 m), 25/xi/1987, Eq.Zoo.coll. [1]; IBUFRJ 8844, off Camburi (60 m), 12/ii/1987, Eq.Zoo.coll. [1]; --Rio de Janeiro State: IBUFRJ 9728, Arquipélago de Santana, Macaé, v/1993, NOAG coll. [35]; IBUFRJ 8836, off Cabo Frio, CFVII # 6147 (22°53.7'S, 041°50.5'W, 50 m), 24/iii/1983, NOAS coll. [22]; IBUFRJ 7197, CFVII # 6174 (23°16.8'S, 043°02.7'W, 92 m), 29/iii/1983, NOAS coll. [1]; IBUFRJ 7204, CFVII # 6197 (23°54.2'S, 044°12.4'W, 103 m), 02/iv/1983, NOAS coll. [5]; IBUFRJ 8837, CFVII # 6178 (23°39.7'S, 043°14.2'W, 119 m), 30/iii/1983, NOAS coll. [14]; IBUFRJ 8838, CFVII # 6172 (23°42.2'S, 043°01.1'W, 129 m), 28/03/1983, NOAS coll. [5]; IBUFRJ 8839, off Cabo Frio, CFVII # 6165 (23°02.8'S, 042°46'W, 56 m), 23/iii/1983, NOAS coll. [23]; MORG 11259, off Rio de Janeiro State, vi/1966, S. Paes coll. [2]; MORG 16557, off Rio de Janeiro State (100 m), xi/1969, 100 m, NOAS coll. [1]; MORG 19860, Praia dos Anjos, Cabo Frio, vi/1972, NOAS coll. [1]; MORG 27096, off Cabo Frio (55 m), xi/1988, NOAS coll. [1]; MZSP19451, off Cabo Frio (23°S, 042°10'W, 64 m), 10/iii/1971, NOWB coll. [4]; --Rio Grande do Sul State: MORG 23011, off Rio Grande # 13 (23 m), x/1983, AS coll. [1]; MORG 23064, off Rio Grande # 30, 26 m, x/1983, AS coll. [2]; MORG 23091, off Rio Grande # 38 (38 m), x/1983, AS coll. [10].

Distribution

Mayaguez, Porto Rico; West Indies; Colombian Caribbean; southeast and south coasts of Brazil (Espírito Santo, Rio de Janeiro and Rio Grande do Sul States).

Remarks

Although described from the Caribbean region, where it was recorded from several localities (Jong & Coomans, 1988; Diaz & Puyana, 1994), along the Brazilian coast, *T. portoricana* is restricted to localities below 19° S (Espírito Santo State). In spite of this apparently disjunct distribution, the Brazilian specimens (figs 74-78) doubtless belong to this species (holotype in figs 72, 73). The similarities in the shell shape, with a conical outline and straight whorl profiles (figs 72, 74), and in the sculpturing (figs 73, 75, 76) are remarkable. Also, the rhomboid aperture with a well-developed columellar fold and reflected inner lip (fig. 75) is exactly the same as in the holotype (figs 73). Indeed, the spiral sculpture formed by six or seven deep

rectangular furrows (fig. 76) is very constant in the several Brazilian exemplars studied.

Turbonilla maestratii n. sp. (figs 79-84)

Description

Shell moderately tall, slender, slightly conical to subcilindrical; fresh shells transparent, older ones white. Teleoconch whorls almost flat-sided in profile. Suture shallow, somewhat sinuous by ribs projection. Protoconch heterostrophic planispiral; diameter about 290 µm. Axial ribs slender, straight and orthocline; slightly projected over anterior suture; ending abruptly at periphery of last whorl; 19 on body whorl of holotype; interspaces as wide as the ribs. Spiral sculpture formed by about 17 rows of rectangular, deep furrows of regular width and spacing. Base rounded tending to elongate, with 4-5 thin spiral striae. Aperture slightly pyriform tending to rhomboid. Columella somewhat arcuate, with obsolete fold. Outer lip thin. No umbilical fissure.

Dimensions.

Holotype with 11 teleoconch whorls; height 5.3 mm; width 0.9 mm.

Type material

Holotype: MNRJ 8934 Praia do Despacho, Itaparica, Bahia State, 1984-99, P. Maestrati coll.; Paratypes: ANSP 410347; MNHN; ZMA 4.02.023; MORG 41044; MNRJ 8927; MZSP 35852; MNRJ 8933; IBUFRJ 11906, type locality.

Type locality

Praia do Despacho, Itaparica, Bahia State, northeast coast of Brazil.

Additional material

--Pará State: IBUFRJ 8905, off Pará, AMASSEDS # 4134, RVCI coll. [2]; --Pernambuco State: MMUFRPE, off Itaparica [8]; --Bahia State: MNHN, Praia do Despacho, Itaparica, 1984-99, P.Maestrati coll. [27]; IBUFRJ 9781, Bom Despacho, Itaparica, L.Trinchão coll. [2]; -- Espírito Santo State: IBUFRJ 9427, Camburi # 2 (45 m), 04/ix/1986, Eq.Zoo coll. [1]; --Rio de Janeiro State: IBUFRJ 9467, off Cabo Frio, CFVII # 6147 (22°53.7'S, 041°50.5'W, 50 m), 24/iii/1983, NOAS coll. [2]; IBUFRJ 10297, off Prainha, Arraial do Cabo, 1990, T.Almeida coll. [1].

Distribution

North, northeast and southeast coasts of Brazil (Pará, Bahia, Espírito Santo and Rio de Janeiro States).

Etymology

The species is named after Dr. Phillippi Maestrati (MNHN), who collected mollusks along the northeast coast of Brazil, including the material of this new species.

Remarks

Turbonilla maestratii (figs 79-84) shows some resemblance to T.





Figs 72-78. Turbonilla portoricana Dall & Simpson, 1901. 72, 73: holotype (USNM 160204); 74, 76, 78: (IBUFRJ 8837); 75: (IBUFRJ 8839); 77: (IBUFRJ 8836). Figs 72, 74: whole shells (lengths: 72: 4.4 mm; 74: 5.5 mm); figs 73, 75: last wholrs; fig. 76: detail of sculpture; figs 77, 78: protoconch. Scale bars: 200 μm. Figs. 72-78. Turbonilla portoricana Dall & Simpson, 1901. 72, 73: olotipo (USNM 160204); 74, 76, 78: (IBUFRJ 8837); 75: (IBUFRJ 8839); 77: (IBUFRJ 8836). Figs. 72, 74: conchiglie intere (lunghezze: 72: 4,4 mm; 74: 5,5 mm); figg. 73, 75: ultimo giro; fig. 76: dettaglio della scultura; figg. 77, 78: protoconca. Scala di riferimento: 200 μm.

portoricana (figs 72-78), especially in the conical shell shape, straight whorl profile, and axial ribs. It can be distinguished, however, by the spiral striae (figs 81, 82), much more numerous and narrow in *T. maestratii*, and by the aperture (fig. 81), which does not show the columellar fold. In addition, the axial ribs do not extend over the base (fig. 81), as they do in *T. portoricana* (figs 73, 75).

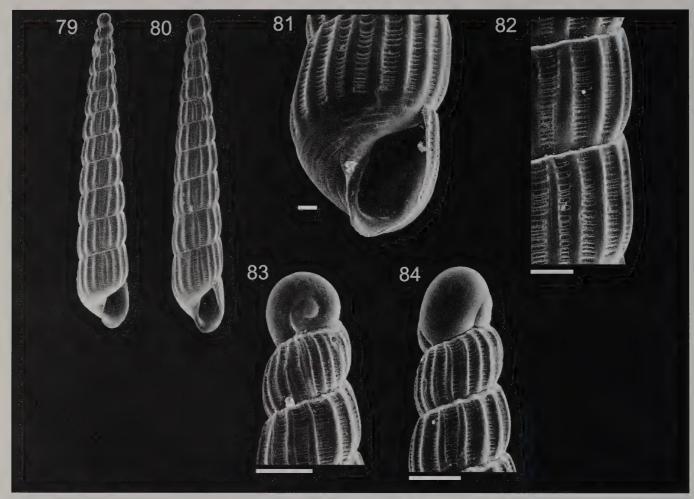
Turbonilla insularis Dall & Simpson, 1901 and *Turbonilla santodomingensis* Gabb, 1873 (figure in Pilsbry, 1922) are similar in their slender conical shell shape, but in both species the spiral sculpturing is composed of fewer striations than in *T. maestratii* (7 in *T. santodomingensis* and 11 in *T. insularis*, while there are about 15 in *T. maestratii*) (figs 81, 82). Moreover, *T. insularis* has more axial ribs on the last whorl (28), which invade the base; in *T. maestratii*, there are fewer axial ribs (19 on the last whorl of the holotype), which end abruptly before entering the base (fig. 81).

Turbonilla kaapor n. sp. (figs 85-89)

Description

Shell moderately tall, slender, slightly conical to subcilindrical; color white. Teleoconch whorls slightly convex in profile. Suture somewhat deep and sinuous by ribs projection. Protoconch heterostrophic planispiral elevated; diameter about 240 µm. Axial ribs well marked, broad, straight and slightly prosocline; projected over anterior suture; fairly distinct on first teleoconch whorl; ending abruptly at periphery of last whorl; 25 on body whorl of holotype; interspaces as wide as the ribs. Spiral sculpture formed by 6-7 rows of rectangular, deep furrows regularly spaced. Base rounded-elongate, with 4-5 thin spiral striae. Aperture pyriform tending to rhomboid. Columella somewhat arcuate, without fold. Outer lip thin. No umbilical fissure.





Figs 79-84. Turbonilla maestratii n. sp. 79, 81, 82: holotype (MNRJ 8934); 80: paratype (MNRJ 8927), 83: paratype (MNRJ 8927); 84: paratype (MNRJ 8927). Figs 79, 80: whole shells (lengths: 79: 5.3 mm, 80: 4.4 mm); fig. 81: last whorl; fig. 82:detail of sculpture; figs 83, 84: protoconchs. Scale bars: 200 µm. Figg. 79-84. Turbonilla maestratii n. sp. 79, 81, 82: olotipo (MNRJ 8934); 80: paratipo (MNRJ 8927), 83: paratipo (MNRJ 8927); 84: paratipo (MNRJ 8927). Figg. 79, 80: conchiglie intere (lunghezze: 79: 5,3 mm, 80: 4,4 mm); fig. 81: ultimo giro; fig. 82: dettaglio della scultura; figg. 83, 84: protoconche. Scala di riferimento: 200 µm.

Dimensions

Holotype with 7.25 teleoconch whorls; height 4.1 mm; width 0.9 mm.

Type material

Holotype: MZSP 35853, off São Paulo State, PADCT # 6631 (25°46'S, 045°28.8'W, 164 m), NOWB coll. Paratypes: MNRJ 8932; IBUFRJ 11907; ANSP 410348; MNHN, type locality; MZSP 35854, off Santa Catarina State, PADCT # 6611 (28°24.3'S, 047°21.6'W, 195 m).

Type locality

Off São Paulo State, PADCT # 6631 (25°46'S, 045°28.8'W, 164 m), southeast coast of Brazil.

Distribution

Southeast-south costs of Brazil (São Paulo and Santa Catarina States).

Etymology

This species is named after the Brazilian Indian tribe Kaapor.

Remarks

Turbonilla kaapor (figs 85-89) is somewhat similar to *T. portoricana* (figs 72-78) in the axial and spiral sculpture, but the axial ribs are faint in the first teleoconch whorl (figs 85, 88, 89), the shell is pupoid in outline (fig. 85), and the protoconch is elevated (figs 88, 89), with its whorl somewhat detached from the teleoconch (fig. 89). This kind of protoconch is also present in *Turbonilla stimpsoni* Bush, 1899 (figs 92, 93), which is also similar in having faint ribs on the first teleoconch whorl; but the shell of the latter is taller, with a more acuminate apex (figs 90, 91) and the spiral sculpture is more complex, with more numerous and irregular striae (fig. 94) than *T. kaapor*, which has about seven rectangular pits per interspace (figs 86, 87).

Turbonilla stimpsoni Bush, 1899 (Figs 90-94)

Turbonilla stimpsoni Bush, 1899: 156, pl. 8, fig. 7; Jong & Coomans (1988: 130, pls. 20, 26, fig. 677); Johnson (1989: 65); Odé (1996: 57); Absalão & Pimenta (1999: 82-83, figs. 16, 16a).



Type locality

Coasts of North and South Carolina.

Types

Holotype by monotypy: ANSP 72042.

Material examined

The type and: --Amapá State: IBUFRJ 8861, AMASSEDS # 3210 (01°52.45'N, 016.02'W, 47 m), 12/v/1990, RVCI coll. [18]; IBUFRJ 8939, AMASSEDS # 3209 (01°20.9'N, 048°00.2'W, 53 m), 12/v/1990, RVCI coll. [5].

Distribution.

Coasts of North and South Carolina; north coast of Brazil (Amapá State).

Remarks

For description, see Bush (1899) and Absalão & Pimenta (1999). This species is compared with T. kaapor (figs 85-89) in the section dealing with the latter.

Turbonilla cf. riisei (Mörch, 1875) (figs 95-100)

Chemnitzia riisei Mörch, 1875: 165.

Turbonilla riisei: Jong & Coomans (1988: 129, pl. 7fig. 673); Odé (1996: 56); Absalão & Pimenta (1999: 81-82, fig. 20); Redfern (2001: 150, pl. 67, figs 623A, 623B).

Turbonilla pilsbryi Bush, 1899: 151, pl. viii, fig. 9; Johnson (1989: 58).

Type locality.

Saint Thomas, West Indies.

Types

Types not located. Holotype of *T. pilsbryi* by monotypy: ANSP 72045.

Material examined

The type and: --Amapá State: MORG 38581, off Amapá State (56 m), 01/v/1968, NOAS coll. [1]; --Pará State: IBUFRJ 8979, off Pará State, RVCI coll. [2]; --Bahia State: IBUFRJ 9459, off Guarajuba, Itapuã, 1990, L.Trinchão coll. [1]; --Espírito Santo State: IBUFRJ 8975, off Espírito Santo State [7]; IBUFRJ 8976, off Aracruz, 01/viii/1988, V.Abud coll. [8]; IBUFRJ 9790, off Camburi, 19/iii/1993, Eq.Zoo. coll. [4]; --Rio de Janeiro State: IBUFRJ 8978, off Prainha, Arraial do Cabo, T.Almeida coll. [1]; MZSP 30906, off Bacia de Campos # 01 (21°21'05"S, 040°47'25"W, 20 m) [2].

Distribution

Bermuda; West Indies; north, northeast and southeast coasts of Brazil (Amapá, Pará, Bahia, Espírito Santo and Rio de Janeiro States).

Remarks

For descriptions and discussions, see Bush (1899), Jong &

Coomans (1988), and Absalão & Pimenta (1999). Jong & Coomans (1988) pointed out that *T. pilsbryi* Bush, 1899 is a junior synonym of *T. riisei*, a position followed by Odé (1996). This was based on the fact that the holotype of *T. pilsbryi* (photograph in Absalão & Pimenta, 1999: fig. 20) encompasses the variation found by Jong & Coomans (1988) in *T. riisei*.

The shell of *Turbonilla riisei* from the West Indies illustrated by Jong & Coomans (1988), and also the holotype of *T. pilsbryi*, are more inflated and wider than the Brazilian specimens. In fact, the Brazilian material shows wide variation in shell form (figs 95-97), with some specimens (fig. 95) almost as wide as the material from the Caribbean, but others (fig. 97) more slender. Since the spiral and axial sculptures are in the same pattern in the Brazilian specimens (fig. 98) and in the holotype, and we could not correlate the variation in shell form of the Brazilian specimens with any geographical distribution, we regard all of them as belonging to the taxon *T. cf. riisei*.

Turbonilla capixaba n. sp. (figs 101-106)

Description

Shell small, slightly conical to slightly subcilindrical; color white with yellow spiral band on middle of each whorl. Teleoconch whorls straight to slightly convex in profile. Suture somewhat deep, slightly sinuous by ribs projection. Protoconch heterostrophic planispiral; diameter about 290 µm. Axial ribs slender, straight or slightly sinuous, orthocline or slightly opisthocline; 21 on body whorl of holotype; interspaces about twice the width of the ribs, sculptured by microscopic axial striae. Spiral sculpture formed by about 16 rows of furrows of irregular width and spacing along interspaces. Base rounded, with evanescent ribs and very thin spiral furrows. Aperture pyriform tending to rhomboid. Columella obliquely straight, with very obsolete fold. Outer lip thin. No umbilical fissure.

Dimensions

Holotype with 7 teleoconch whorls; height 3.8 mm; width 1.1 mm.

Type material

Holotype: MNRJ 8926, off Espírito Santo State, REVIZEE # VV38 (19°28'S, 038°22'W, 71.4 m), 29/ii/1996, NOAN coll.; Paratypes: MNRJ 8937; MORG 41045; MZSP 35857; IBUFRJ 11908; ZMA 4.02.024; ANSP 410349; MNRJ 8938; MNHN, type locality.

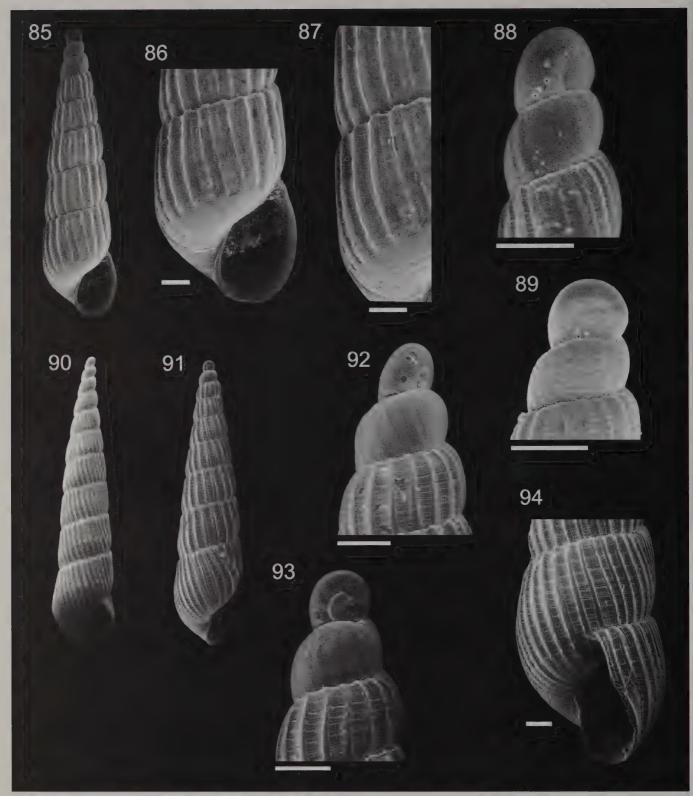
Type locality

Off Espírito Santo State, REVIZEE # VV38 (19°28'S, 038°22'W, 71.4 m), southeast coast of Brazil.

Additional material

-- Espírito Santo State: MORG 39059, REVIZEE # VV38 (19°28'S, 038°22'W, 71.4 m), 29/ii/1996, NOAN coll. [5]; IBUFRJ 9660, REVIZEE # VV38 (19°28'S, 038°22'W, 71.4 m), 29/ii/1996, NOAN coll. [37]; IBUFRJ 9409, REVIZEE # VV24 (20°S, 039°54'W, 45 m), 27/ii/1996, NOAN coll. [5]; -- Rio de Janeiro State: IBUFRJ 9768, off Bacia de Campos





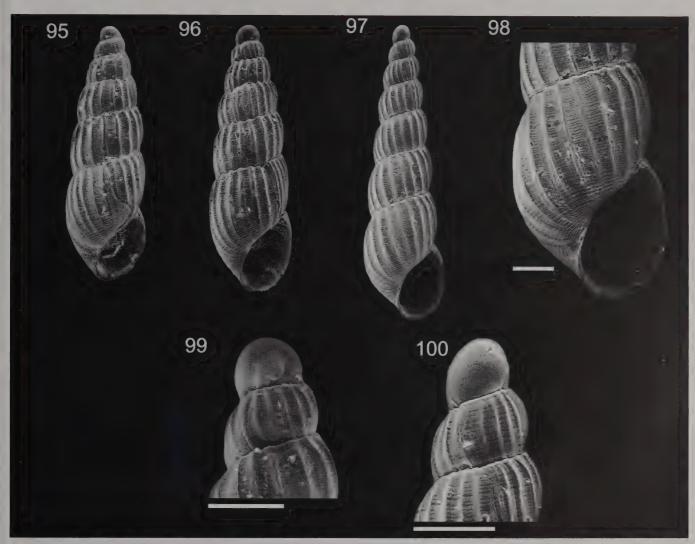
Figs 85-89. Turbonilla kaapor n. sp. 85-88: holotype (MZSP 35853); 89: paratype (MNRJ 8932). Fig. 85: whole shell (length 4.1 mm); fig. 86: last whorl; figs. 87: detail of sculpture on last whorl; figs 88, 89: protoconchs. Scale bars: 200 µm.

Figs 90-94. Turbonilla stimpsoni Bush, 1899. 90: holotype (ANSP 72042); 91-94: (IBUFRJ 8861). Figs 90, 91: whole shells (lengths: 90: 4.8 mm, 91: 4.7 mm); figs 92, 93: protoconchs; fig. 94: last whorl. Scale bars: 200 μm.

Figg. 85-89. Turbonilla kaapor n. sp. 85-88: olotipo (MZSP 35853); 89: paratipo (MNRJ 8932). Fig. 85: conchiglia intera (lunghezza 4,1 mm); fig. 86: ultimo giro; fig. 87: dettaglio della scultura dell' ultimo giro; figure 88, 89: protoconche. Scala di riferimento: 200 µm.

Figg. 90-94. Turbonilla stimpsoni Bush, 1899. 90: olotipo (ANSP 72042); 91-94: (IBUFRJ 8861). Figg. 90, 91: conchiglie intere (lunghezze: 90: 4,8 mm, 91: 4,7 mm); figg. 92, 93: protoconche; fig. 94: ultimo giro. Scala di riferimento: 200 µm.





Figs 95-100. Turbonilla cf. riisei (Mörch, 1875). 95: (IBUFRJ 8980); 96-100: (IBUFRJ 8975). Figs 95-97: whole shells (lengths: 95: 2.6 mm, 96: 2.9 mm, 97: 3.9 mm); fig. 98: last whorl; figs 99, 100: protoconchs. Scale bars: 300 µm.

Figg. 95-100. Turbonilla cf. riisei (Mörch, 1875). 95: (IBUFRJ 8980); 96-100: (IBUFRJ 8975). Figg. 95-97: conchiglie intere (lunghezze: 95: 2,6 mm, 96: 2,9 mm, 97: 3,9 mm); fig. 98: ultimo giro; figg. 99, 100: protoconche. Scala di riferimento: 300 μm.

(22°47'47"S, 040°45'32"W), NOAG coll. [1]; IBUFRJ 9410, off Cabo Frio, CFVII # 6165 (23°02.8'S, 042°46'W, 56 m), 23/iii/1983, NOAS coll. [5].

Distribution

Southeast coast of Brazil (Espírito Santo and Rio de Janeiro States).

Etymology

"Capixaba" is the general determination to those people that was born in Espírito Santo State, Brazil.

Remarks

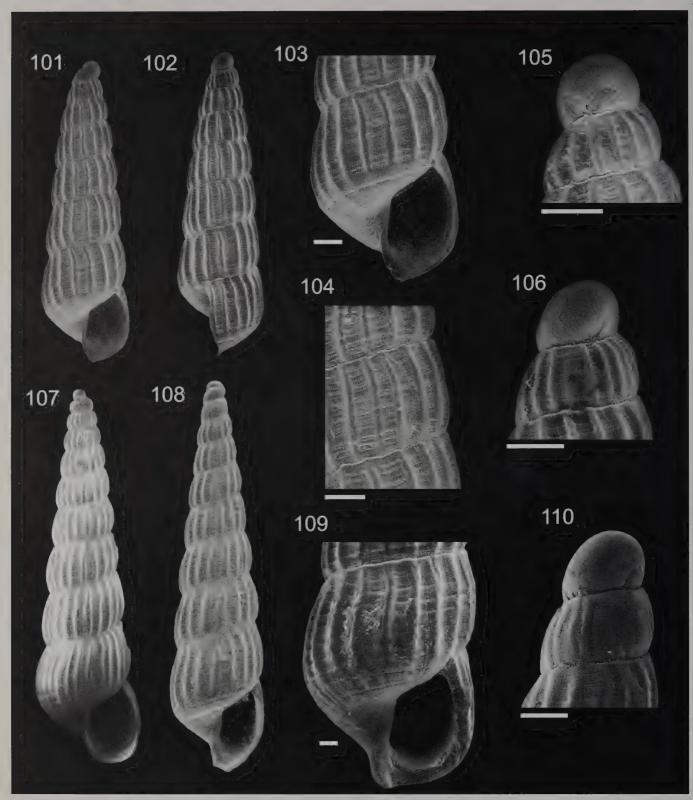
Turbonilla capixaba (figs 101-106) is somewhat similar in general shell shape to Turbonilla interrupta (Totten, 1835) and its many synonyms, such as Turbonilla areolata Verrill, 1873 and Turbonilla buteonis Bartsch, 1909, among others [complete lists of synonyms in Abbott (1974) and Odé (1996)]. However, there are differences in the more numerous spiral sculpture of T. capixaba (figs 103, 104), more marked axial ribs, and more con-

vex whorl outline (figs 101-103).

Turbonilla capixaba shows some similarity in spiral sculpture with Turbonilla rushii Bush, 1899 [holotype illustrated by PIMENTA & ABSALÃO (2001: 79, fig. 17)] and T. anira Bartsch, 1927 (fig. 107), but the shell is not as tall as in these two species, and the whorls outline is straighter. The most similar species, however, are Turbonilla lineolata Bush, 1899 and Turbonilla fasciata (d'Orbigny, 1840). Turbonilla lineolata also has a yellow spiral band and similar shell shape, but the shell is larger, less slender, with more regularly convex whorls, the spiral lines are more regularly spaced, lacking the finer lines, and the axial ribs are straighter. Turbonilla fasciata [figures in PIMENTA & ABSALÃO (2001: 77, figs 12-16)] is also similar, especially in the spiral sculpture pattern, but T. capixaba has more sinuous axial ribs (figs 103, 104) and a more conical shell (figs 101, 102) than T. fasciata, which has scalloped whorls in a somewhat pupoid shell shape.

Turbonilla cf. anira Bartsch, 1927 (figs 107-110) Turbonilla anira Bartsch, 1927: 84-85; ODÉ (1996: 35).





Figs 101-106. Turbonilla capixaba n. sp. 101-105: holotype (MNRJ 8926); 106: paratype (MNRJ 8937). Figs 101, 102: whole shell (length: 3.8 mm); fig. 103: last whorl; fig. 104: detail of sculpture; figs 105, 106: protoconchs. Scale bars: 200 µm.

Figs 107-110. Turbonilla cf. anira Bartsch, 1927: 107: holotype (USNM 108058); 108, 109: (IBUFRJ 9426); 110: (IBUFRJ 3151). Figs 107, 108: whole shells (lengths: 107:8.3 mm, 108: 7.8 mm); fig. 109: last whorl; figs 110: protoconch. Scale bars: 200 μm.

Figg. 101-106. Turbonilla capixaba n. sp. 101-105: olotipo (MNRJ 8926); 106: paratipo (MNRJ 8937). Figure 101, 102: conchiglia intera (lunghezza: 3,8 mm); fig. 103: ultimo giro; fig. 104: dettaglio della scultura; figg. 105, 106: protoconche. Scala di riferimento: 200 µm.

Figg. 107-110. Turbonilla cf. anira Bartsch, 1927: 107: olotipo (USNM 108058); 108, 109: (IBUFRJ 9426); 110: (IBUFRJ 3151). Figg. 107, 108: conchiglie intere (lunghezze: 107: 8,3 mm, 108: 7,8 mm); fig. 109: ultimo giro; fig. 110: protoconca. Scala di riferimento: 200 μm.



Type locality.

Off Fernandina, Florida (530 m).

Type material

Holotype: USNM 108058; 12 paratypes (?): USNM 360179.

Material examined

The holotype's photograph and: --Espírito Santo State: IBUFRJ 9415, off Espírito Santo State [5]; --Rio de Janeiro State: IBUFRJ 8720 off Bacia de Campos, NOAG coll. [1]; IBUFRJ 9417, CFVII # 6194 (24°03.6'S, 044°07.6'W, 134 m), 01/iv/1983, NOAS coll. [2]; IBUFRJ 3151, Prainha, Arraial do Cabo, 1989, T.Almeida coll. [2]; MZSP 28880, off Angra dos Reis, 18/ii/1968 [1]; IBUFRJ 9416, CFVII # 6178 (23°39.7'S, 043°14.2'W, 119 m), 30/iii/1983, NOAS coll. [4]; IBUFRJ 10313, REVIZEE # D3 (22°52'S, 041°09'W, 80 m), 23/ii/1996, NOAN coll. [2]; IBUFRJ 11071, off Bacia de Campos, NOAG coll. [1].

Distribution

Florida; southeast coast of Brazil (Espírito Santo and Rio de Janeiro States).

Remarks

The Brazilian specimens determined herein as *Turbonilla* cf. anira (figs 108-110) differ slightly from the type of *T. anira* (fig. 107). In the type there are more and more sinuous axial ribs, the first four whorls are wider (fig. 108), and the protoconch is smaller (fig. 107). In spite of these differences, the general shell shape and sculpture are very similar (figs 107, 108).

Turbonilla cf. anira most closely resembles T. rushii Bush, 1899 [holotype figured by ABSALÃO & PIMENTA (1999: fig. 17)], but the spiral sculpturing in T. rushii is more numerous and variable than in T. cf. anira (fig. 109).

Turbonilla scapulata n. sp. (figs 111-115)

Description

Shell moderately tall, conical with somewhat scaloned whorls; color white. Teleoconch whorls almost flat-sided, slightly concave on middle and shouldered at the summits. Suture somewhat deep and sinuous, with distinct subsutural shelf. Protoconch heterostrophic planispiral; diameter about 280 µm. Axial ribs slender, straight and orthocline; strongly shouldered apiacally, not reaching the anterior suture, leaving a spiral band that looks like a second suture; 23 on body whorl of holotype; interspaces much wider than the ribs, bearing microscopic axial growth lines. Spiral sculpture formed by about 20 rows of very thin and close furrows, with three wider furrows, below the shoulder of the whorls, on middle of whorl and just above the suture. Base rounded, with evanescent ribs and very thin spiral striae. Aperture rhomboid. Columella straight, without fold. Outer lip thin. No umbilical fissure.

Dimensions

Holotype with 8 teleoconch whorls; height 4.6 mm; width 1.3 mm.

Type material

Holotype: MZSP 35858, off São Paulo State, PADCT # 6579 (24°42.3'S, 045°18.8'W, 84 m), NOWB coll.; Paratypes: IBUFRJ 11909; ANSP 410350; MNHN; MNRJ 8920; MZSP 35862, type locality.

Type locality

Off São Paulo State, PADCT # 6579 (24°42.3'S, 045°18.8'W, 84 m), southeast coast of Brazil,

Distribution

Only known from type locality (off São Paulo State).

Etymology

This species is named after its shouldered whorls (*scapula*, L. = shoulder blade).

Remarks

Turbonilla scapulata (figs 111-115) shows some similarity to Turbonilla asperula Bush, 1899 [lectotype illustrated by ABSALÃO & PIMENTA (1999: figs. 17, 17a)] in the apically shouldered axial ribs (fig. 113) and whorls (figs 111, 112). However, it is clearly distinguished by its much more numerous and finer spiral sculpture (fig. 113).

Turbonilla paulinoi n. sp. (figs. 116-120)

Description

Shell small, conical; color white. Teleoconch whorls semi-pyriform in profile. Suture somewhat deep and sinuous by ribs projection. Protoconch heterostrophic planispiral; diameter about 270 µm. Axial ribs slender, slightly sinuous and strictly prosocline; strongly shouldered apiacally and projected over anterior suture; 24 on body whorl of holotype; interspaces a little bit wider than ribs. Spiral sculpture formed by seven rows of rectangular, deep furrows regularly spaced intercaling with spiral cords that cross the axial ribs; the second cord, below the suture is wider and form small nodules when crossing the ribs. Base rounded, with ribs that continue through the parietal region and rows of spiral furrows. Aperture pyriform. Columella somewhat arcuate, without fold. Outer lip thin. No umbilical fissure.

Dimensions

Holotype with 6.5 teleoconch whorls; height 2.9 mm; width 0.8 mm.

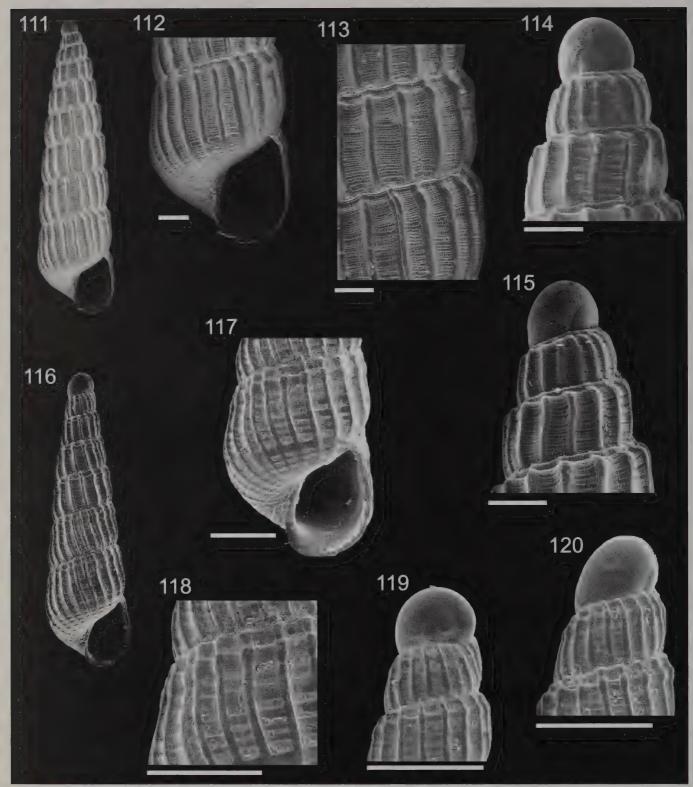
Type material

Holotype: MNRJ 8941, off Pará State, AMASSEDS # 3209 (01°20.9'N, 48°00.2'W, 53 m), 12/v/1990, RVCI coll.; Paratypes: ANSP 410351; MNHN; MORG 41046; ZMA 4.02.025; MZSP 35860, type locality; IBUFRJ 11910, off Amapá State, AMASSEDS # 3228 (03°25.1'N, 49°56.4'W, 64 m), 17/v/1990, RVCI coll.

Type locality

Off Pará state (01° 20.9' N / 48° 00.2' W, 53 m), north coast of





Figs 111-115. Turbonilla scapulata n. sp. 111-114: holotype (MZSP 35858); 115: paratype (MNRJ 8920). Fig. 111: whole shell (length 4.6 mm); fig. 112: last whorl; fig. 113: detail of sculpture; figs 114, 115: protoconch. Scale bars: 200 μm.

Figs 116-120. Turbonilla paulinoi n. sp. 116-119: holotype (MNRJ 8941); 120: paratype (MORG 41046). Fig. 116: whole shell (length 2.9 mm); fig. 117: last whorl; fig. 118: detail of sculpture; figs 119, 120: protoconchs. Scale bars: $400 \, \mu m$.

Figg. 111-115. Turbonilla scapulata n. sp. 111-114: olotipo (MZSP 35858); 115: paratipo (MNRJ 8920). Fig. 111: conchiglia intera (lunghezza 4,6 mm); fig. 112: ultimo giro; fig. 113: dettaglio della scultura; figg. 114, 115: protoconca. Scala di riferimento: 200 μm.

Figg. 116-120. Turbonilla paulinoi n. sp. 116-119: olotipo (MNRJ 8941); 120: paratipo (MORG 41046). Fig. 116: conchiglia intera (lunghezza 2,9 mm); fig. 117: ultimo giro; fig. 118: dettaglio della scultura; figg. 119, 120: protoconca. Scala di riferimento: 400 μm.



Brazil.

Additional material

--Pará State: IBUFRJ 8945, AMASSEDS # 3209 (01°20.9'N, 48°00.2'W, 53 m), 12/v/1990, RVCI coll. [2].

Distribution

Only known from two localities off Pará and Amapá states, north coast of Brazil.

Etymology

The species is named after Dr. Paulino J. S. de Souza Junior, Brazilian malacologist who collected the type material.

Remarks

The most similar species to *Turbonilla paulinoi* (figs 116-120) is *Turbonilla macaensis* Pimenta & Absalão, 2001. Both species bear a spiral cord of nodules below the sutures. However, in *T. macaensis*, the spiral cords form nodules on all the axial ribs that they cross, the strongest being the anterior one, which touches the suture above it; while in *T. paulinoi*, there are two spiral nodulose cords, the stronger being separated from the suture by a weaker cord of nodules (figs 117, 118).

Turbonilla rachialis n. sp. (figs 121-126)

Description

Shell stout, large, moderately conical; color light beige with white spiral band on middle of whorl. Teleoconch whorls broad, inflate and slightly convex in profile. Suture somewhat deep and sinuous by ribs projection. Protoconch heterostrophic planispiral; diameter about 320 µm. Axial ribs slender, somewhat sharp, straight and strictly orthocline; projected over anterior suture; 18 on penultimate whorl of holotype; interspaces very broad, about three times as wide as the ribs; entire surface of interspaces and tops of the ribs covered by axial micro striae. Spiral sculpture formed by numerous crowded micro striae that cover the interspaces and axial ribs and cross the axial striae forming a delicate reticulate pattern. Base elongate, with evanescent ribs and same spiral pattern as teleoconch whorls. Aperture pyriform. Columella somewhat arcuate, without fold. Outer lip thin. No umbilical fissure.

Dimensions

Holotype with 9 teleoconch whorls; height 10.4 mm; width 3.0 mm.

Type material

Holotype: MNRJ 8918, Prainha, Arraial do Cabo, Rio de Janeiro State, x/1998, P. de Sousa coll.; Paratypes: ANSP 410352, Prainha, Arraial do Cabo, Rio de Janeiro State, 26/ix/1998, P.M. Costa coll.; IBUFRJ 11915, off Piúma (21°10'S, 040°37'W, 16-18 m), Espírito Santo State, 1993, F. Pitombo coll.

Type locality

Prainha, Arraial do Cabo, Rio de Janeiro State, southeast coast

of Brazil.

Distribution

Only known from two localities in Rio de Janeiro and Espírito Santo States.

Etymology

The species is named after its type locality, "Prainha" (small beach). Rachialis, belonging to the beach; rachia, L. = beach.

Remarks

Turbonilla rachialis (figs 121-126) is one of the largest Turbonilla in Brazil, with a very inflated shell (fig. 121, 122) and very low, sharp axial ribs with wide interspaces (figs 122, 123). It is also unique in its sculpture pattern, with very fine and crowded spiral sculpture crossing the microscopic axial striae, both crossing the axial ribs (figs 123, 124). These characteristics distinguish T. rachialis from any other Brazilian species.

Turbonilla uaca n. sp. (figs 127-132)

Description

Shell stout, large, tall, conical, with acuminate apex, last whorls inflated; color light brown. Teleoconch whorls straight in profile. Suture deep, sinuous by ribs projection. Protoconch heterostrophic planispiral; diameter about 300 µm. Axial ribs strictly straight and slightly prosocline; projected over anterior suture; 28 on body whorl of holotype; interspaces as wide as the ribs. Spiral sculpture formed by seven rows of deep furrows irregular both in width and spacing. Base rounded, with evanescent ribs and rows of spiral striae. Aperture pyriform tending to rhomboid. Columella somewhat arcuate, with distinct fold. Outer lip thin. Parietal callus well developed. Umbilical present.

Dimensions

Holotype with 11 teleoconch whorls; height 7.2 mm; width 1.7 mm.

Type material

Holotype: MORG 41047, off Amapá State (56 m), 01/v/1968, NOAS coll.; Paratypes: MNRJ 8942; IBUFRJ 11911; MZSP 35863; ANSP 410353; MNHN; MORG 41052, type locality; MORG 41048, off Cabo Orange, Amapá State (85 m), 1970, NOAS coll.; ZMA 4.02.026, Prefil de Caviana, Pará State (56 m), 01/v/1968, NOAS coll.

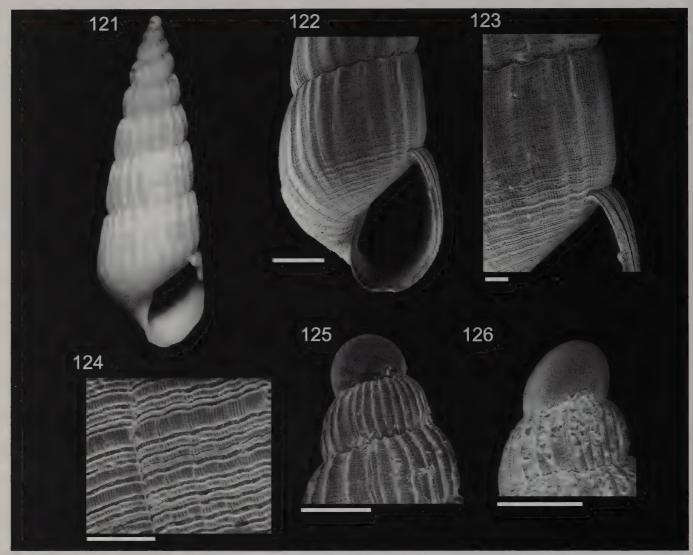
Type locality

Off Amapá State, north coast of Brazil.

Additional material

--Amapá state: MORG 15886, off Cabo Orange (85 m), 1970, NOAS coll. [1]; MORG 14811, off Amapá State (56 m), 01/v/1968, NOAS coll. [6]; --Pará State: IBUFRJ 8871, AMASSEDS # 4134, x/1991, RVCI coll. [1]; IBUFRJ 9455, AMASSEDS # 3210 (01°52.45'N, 16.02'W, 47 m), 12/v/1990, RVCI coll. [3].





Figs 121-126. Turbonilla rhachialis n. sp. 121: holotype (MNRJ 8918). 122-126: paratype (ANSP 410352). Fig. 121: whole shell (length: 10.4 mm); fig. 122: last whorl; figs 123, 124: details of sculpture; figs 125, 126: protoconchs. Scale bars: figs 123, 125,126: 300 μm; fig. 122: 1.0 mm; fig. 124: 100 μm.

Figg. 121-126. Turbonilla rhachialis n. sp. 121: olotipo (MNRJ 8918). 122-126: paratipo (ANSP 410352). Fig. 121: conchiglia intera (lunghezza: 10,4 mm); fig. 122: ultimo giro; figg. 123, 124: dettaglio della scultura; figure 125, 126: protoconca. Scala di riferimento: figg. 123, 125, 126: 300 μm; fig. 122: 1,0 mm; fig. 124: 100 μm.

Distribution

North coast of Brazil (Amapá and Pará States).

Etymology

This species is named after the Brazilian Indian tribe Uacá that inhabits an area in Amapá State.

Remarks

Turbonilla uaca (figs 127-132) is one of the largest Turbonilla species of the Brazilian coast. The shell of this species is very characteristic in its acuminate apex (fig. 127) and in having a very developed umbilicus and inner lip in some specimens (fig. 129). Although a slight umbilical fissure and somewhat reflected inner lip may be present in some species, these characters are uncommon and never (at least in the South American fauna) as strong as seen in some specimens of T. uaca. The most similar species is Turbonilla incisa Bush, 1899 [paratype illustrated by

ABSALÃO & PIMENTA (1999: fig. 18)], which has similar shell sculpture, though stronger in *T. uaca* (figs 128-130). They can be distinguished by the acuminate apex of *T. uaca*, in which there is a relatively greater increase in diameter from the fifth whorl on (fig. 127).

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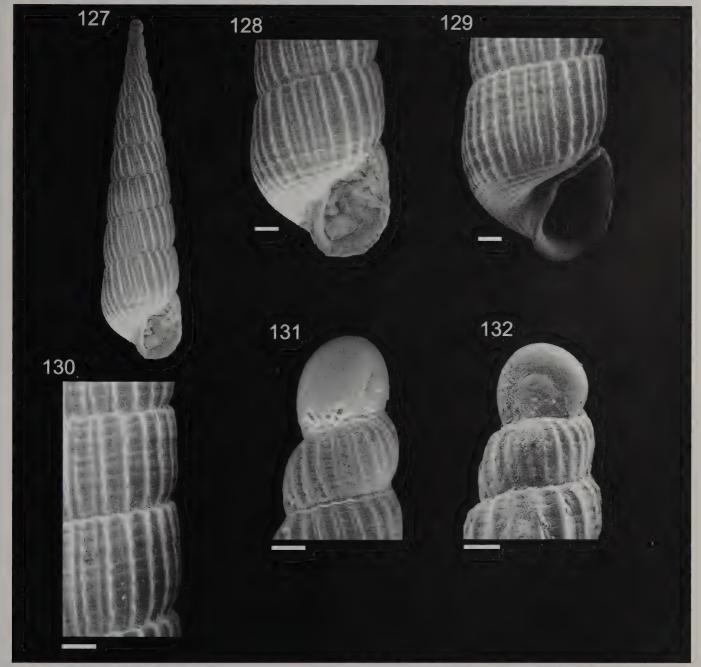
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Figs 127-132. Turbonilla uaca n. sp. 127, 128: holotype (MORG 41047); 129-132: paratype (MNHN). Fig. 127: whole shell (length: 7.2 mm); figs 128, 129: last whorls; fig. 130: detail of sculpture; figs 131, 132: protoconchs. Scale bars: figs 128-130: 300 μm; figs 131, 132: 100 μm.

Figg. 127-132. Turbonilla uaca n. sp. 127, 128: olotipo (MORG 41047); 129-132: paratipo (MNHN). Fig. 127: conchiglia intera (lunghezza: 7,2 mm); figure 128, 129: ultimo giro; fig. 130: dettaglio della scultura; figg. 131, 132: protoconche. Scala di riferimento: figure 128-130: 300 μm; figg. 131, 132: 100 μm.



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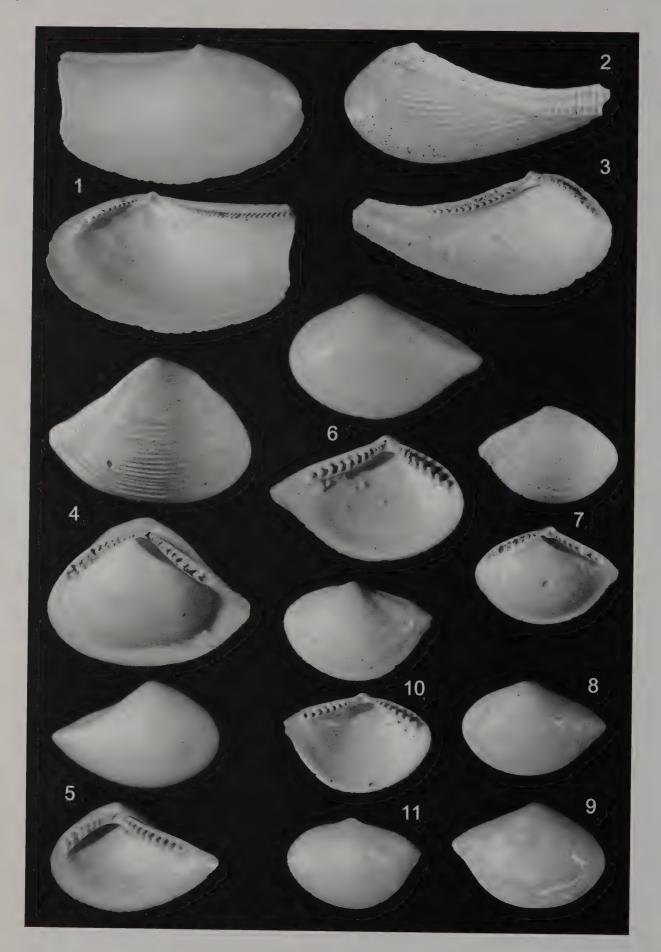
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I manoscritti dovranno essere presentati su fogli bianchi UNI-A4 di buona qualità, scritti con interlinea doppia, ed almeno 3 centimetri di margine ai lati. Tutte le pagine dovranno essere numerate consecutivamente. Tabelle, figure e didascalie dovranno essere stampate su fogli separati; la loro posizione approssimativa nel testo deve essere indicata a margine dello stesso, ma la composizione finale spetta alla Redazione.

I manoscritti dovranno essere organizzati come segue:

Prima pagina: contenente il titolo dell'articolo, il nome(i) per esteso dell'Autore(i), l'indirizzo cui inviare la corrispondenza (comprensivo di indirizzo di posta elettronica, se disponibile), le parole chiave (fino ad un massimo di dieci).

Seconda pagina: con l'Abstract (in inglese), Riassunto (in italiano), Resumen (in spagnolo) o Resume (in francese) a seconda della lingua utilizzata dall'Autore(i) nell'articolo. Un riassunto esteso, più dettagliato e contenente ogni utile riferimento per guidare il lettore ad una rapida comprensione dell'articolo (ad es. i caratteri diagnostici di una specie), deve seguire il riassunto classico. In questa sezione ogni riferimento a figure, tavole o citazioni bibliografiche deve essere evitato. Il riassunto esteso deve essere scritto in inglese od in italiano.

Pagine di testo: il testo deve essere composto da frasi chiare e brevi, possibilmente suddiviso in Introduzione, Materiali e Metodi, Risultati, Discussione, Conclusioni, Ringraziamenti, Riferimenti Bibliografici, Tabelle, Figure e didascalie (in pagine separate). Evitare le note se possibile. Le note indispensabili saranno indicate con un numero progressivo tra parentesi nel testo e collocate in fondo alla pagina cui si riferiscono. Tutte le abbreviazioni dovranno essere spiegate in una legenda. Solo e tutti i nomi di genere e specie devono essere in corsivo (o sottolineati). Ogni nome scientifico dovrà essere accompagnato da Autore ed anno di pubblicazione, la prima volta che viene citato nell'articolo. Tutte le figure devono essere numerate progressivamente con numeri arabi e devono essere citate nel testo. Le figure devono essere presentate su fogli a parte, ognuno con il nome dell'Autore(i) e titolo dell'articolo. I disegni al tratto dovranno essere chiari, tracciati con linee sottili di inchiostro nero di China su carta bianca di alta qualità o carta lucida ed essere almeno delle dimensioni finali di stampa. Quando possibile le figure dovranno essere raggruppate in tavole; la Redazione si riserva il diritto di ridurre o ingrandire gli originali in fase di composizione dell'articolo. Le indicazioni sulle figure (numeri e/o lettere) dovranno essere in caratteri Times New Roman minuscoli (Es. Fig. 1a, 1b, etc.), tenendo conto che nella loro versione finale questi caratteri dovranno avere un'altezza di 2,5-3 mm. Le stampe fotografiche dovranno essere nitide, ben contrastate non montate su cartoncini ma stampate su carta lucida fotografica, di dimensioni non inferiori rispetto a quelle finali. Due o più fotografie montate in una tavola devono avere toni simili. Illustrazioni a colori sono accettate solo se rilevanti scientificamente per il lavoro. Riproduzioni di illustrazioni protette da copyright dovranno essere accompagnate da un'autorizzazione scritta del proprietario del copyright. Si raccomanda agli Autori di inviare dapprima una bozza delle figure (nella loro taglia originale) con il manoscritto. Nella loro versione finale, ad accettazione avvenuta del manoscritto, le figure dovranno essere inviate esclusivamente in formato TIFF (con una risoluzione minima di 300 dpi) su di un disco ad alta capacità Zip 100 o su di un CD-rom. Le citazioni nel testo dovranno seguire uno dei seguenti esempi: "...Monterosato (1869) riportò...", "...Monterosato (1869, 1884) riportò...", "...Verrill & Bush (1900) descrissero...", "De Folin (1867a, 1867b)", "come noto in letteratura (De Folin, 1867a, 1867b; Monterosato, 1869, 1884; Verrill & Bush, 1900)","...du Golfe de Gascogne (Fischer et al., 1872)". Tutte e solo le opere citate nel testo devono essere elencate in ordine alfabetico e cronologico al termine del lavoro.

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Articoli

COGNOME ed iniziale del nome di tutti gli autori, anno. Titolo completo. Nome della rivista per esteso, Volume (fascicolo): prima ed ultima pagina del lavoro.

MONTEROSATO T.A., 1880. Conchiglie della zona degli abissi. Bullettino della Società malacologica italiana, 6 (1-4): 50-64.

Libri:

COGNOME ed iniziale del nome di tutti gli autori, anno. *Titolo completo*. Editore, Città di edizione, numero di pagine (e illustrazioni).

Es.:

WILEY E.O., 1980. Phylogenetics: the theory and practice of phylogenetic Systematics. Wiley, New York, 355 pp.

Capitoli di libri:

COGNOME ed iniziale del nome di tutti gli autori (del capitolo), anno. Titolo completo (del capitolo). In COGNOME ed iniziale dell'Editore(i) (Ed. or Eds), *Titolo del libro*. Città di edizione, Editore, pagine relative al capitolo (of the chapter).

BEDULLI D., CASTAGNOLO L., GHISOTTI F. & SPADA G., 1995 Bivalvia, Scaphopoda. In MINELII A., RUFFO S., & LA POSTA S. (Eds), Check list delle specie della fauna italiana. Bologna, Calderini, 17: 80-90.

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Title page: with title of the article, full name(s) of the Author(s), correspondence address (comprehensive of e-mail address, if available), and Key Words (up to ten).

Second page: with Abstract, Riassunto, Resumen or Resumé according to the language used by the Authors in the article. An Extended Abstract, more detailed and containing any useful reference to drive the reader to a quick comprehension of the article (e.g. the diagnostic characters of a species), must follow the Abstract. References to figures, tables and bibliographic citations however, are to be avoided. The extended abstract must be written in English or Italian.

Text Pages: the text should be presented in clear, short sentences and possibly arranged in Introduction, Material and Methods, Results, Discussion, Conclusions, Acknowledgements, References, Tables, Figures and Legends (in separate pages). Avoid footnotes if possible. If necessary, notes will be indicated by a number between parentheses in the text, and placed at the bottom of the relevant page. All abbreviations must be explained in a legend. Only and all the names of genus and species rank must be italicised (or underlined). Each scientific name will be accompained by its authorship and year of publication, the first time it is mentioned in the text. All figures must be numbered progressively with Arabic numerals, and must be cited in the text. Figures must be submitted in separate pages, each with the name of the Author(s) and the title of the paper. Line drawings should be clear and drawn with thin lines in black Indian ink on high-quality white paper, tracing paper, or plastic film, and be at least the final size. When possible the figures should be grouped in plates: the Redaction will operate the final enlargement/reduction in order to fit the iconography to the composed paper. Pictures in the plate must be labelled with Times New Roman lower case letters (Es. Fig. 1a, 1b, etc.). Labelling on the figures (letters and numbers) must be planned in order to have a final height of 2.5-3 mm. Photographs should be clear, with a sufficiently sharp contrast, printed on white paper with a glossy finish, no less than the final size, and should not be mounted on card. Two or more photographs combined in a plate should be similar in tone. Colour illustrations are accepted only if scientifically relevant to the paper. Reproduction of figures protected by copyright is allowed provided that a written permission by the holder of the copyright is furnished along with the manuscript. Authors are advised in the first instance to send drafts of figures (in original size) with the manuscript. In the final version of an acceped manuscript, figures must be sent exclusively in TIFF format (with a minimum resolution of 300 dpi) on a Zip 100 diskette or on a CD-rom. Citation in the text must follow one of the following examples: "...Monterosato (1869) reported...", "...Monterosato (1869, 1884) reported...", "...Verrill & Bush (1900) described...", "De Folin (1867a, 1867b)", "as known from literature (De Folin, 1867a, 1867b; Monterosato, 1869 1884: Verrill & Bush. 1900)"."...du Golfe de Gascogne (Fischer et al., 1872)". All and only the works cited in the text must be reported alphabetically and chronologically in the refer-

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E.g.:

MONTEROSATO T.A., 1880. Conchiglie della zona degli abissi. *Bullettino della Società malacologica italiana*, 6 (1-4): 50-64.

Books:

SURNAMES and initials of all authors, year. Complete Title. Publisher, place of issue, number of pages and plates.

E.g.

WILEY E.O., 1980. Phylogenetics: the theory and practice of phylogenetic Systematics. Wiley, New York, 355 pp.

Chapters in books:

SURNAMES and initials of all authors (of the chapter), year. Complete title (of the chapter). In Names and initials of the Editor(s) (Ed. or Eds), *Title of the book.* Place of issue, Publisher, number of pages (of the chapter).

E.g.

BEDULLI D., CASTAGNOLO L., GHISOTTI F. & SPADA G., 1995 Bivalvia, Scaphopoda. In MINELLI A., RUFFO S., & LA POSTA S. (Eds), Check list delle specie della fauna italiana. Bologna, Calderini, 17: 80-90.

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SOMMARIO

	B. ÖZTÜRK, G. BUZZURRO & H. AVNI BENLI - Marine molluscs from Cyprus: new data and checklist	49
2,3	C. Bogi & B. S. GALIL - The bathybenthic and pelagic molluscan fauna off the Levantine coast, eastern Mediterranean	79
-	V. Garilli - A new species of Ersilia (Caenogastropoda, Eulimidae) from the Plio-Pleistocene of the central Mediterranean area	_ 91
Kana	S. Gofas, V. Garilli & M.C. Boisselier-Dubayle - Nomenclature of the smaller Mediterranean Cerithium species	_ 95
	R. Rocchini - Prima segnalazione di <i>Mathilda barbadensis</i> Dall, 1889, in Mediterraneo (Heterostropha, Mathildidae)	_ 105
•	F. Boyer, P. Ryall, A. Wakefield - A Revision of Marginella reeveana Petit, 1851	_ 107
	P. Lozouet - Djeddilia and Escoffieria, abnormal specimens of Potamididae (Gastropoda: Caenogastropoda: Potamides, Tympanotonos)	_ 111
. 3	A. DIAS PIMENTA, R. SILVA ABSLAO - Fifteen new species and ten new records of <i>Turbonilla</i> Risso, 1826 (Gastropoda, Heterobranchia, Pyramidellidae) from Brazil	113
	R. La Perna - A causa della cattiva qualità tipografica, vengono ristampate le tavole del lavoro "The Quaternary deep-sea protobranch fauna from the Mediterranean: composition, depth-related distribution and changes" di La Perna R., pubblicato sul fascicolo 39(1-4) del Bollettino Malacologico. Ci scusiamo per il disguido.	25bis